

**WCO-Lite: online world catalogue of  
harvestmen (Arachnida, Opiliones).  
Version 1.0**

*Checklist of all valid nomina in Opiliones with  
authors and dates of publication up to 2018*



Warning: this paper is duly registered in ZooBank and it constitutes a publication *sensu* ICZN. So, all nomenclatural acts contained herein are effective for nomenclatural purposes. WCO logo, color palette and eBook setup all by AB Kury (so that the reader knows who's to blame in case he/she wants to wield an axe over someone's head in protest against the colors).

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*“Basta de castillos de arena, hagamos edificios de hormigón armado (con una piscina en la terraza superior).”*

*Miguel Angel Alonso-Zarazaga*

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







Mônica de Almeida Rocha - CRB7 2209



# World Catalogue of Opiliones

## WCO-Lite: online world catalogue of harvestmen (Arachnida, Opiliones). Version 1.0 – Checklist of all valid nomina in Opiliones with authors and dates of publication up to 2018

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# Chapter 1. Introduction and presentation of the project WCO

We are witnessing a raging storm on planet Earth. Greedy mankind is long past caring about the environment. Unprecedented wholesale **destruction of biodiversity** rampages in full swing, igniting a painful ordeal for the future generations. Reports of taxonomy's death have been exaggerated, but not by much. Nomenclature became a hermetic art, no longer mastered by zoologists. Kickstarting such a thing as a taxonomic catalogue may appear somewhat anachronistic. And yet... we, the authors of this work, are all cut from the same cloth: black kevlar. As such, we are holding up to the stereotype of survivors, struggling to keep our heads afloat by pulling off a **catalogue** in such a neglected line of work.

The **project *World Catalogue of Opiliones*** (Kury 2016~) is composed of three modules: (1) Kury's World Catalogue of Opiliones (**WCO**) a conventional paper, currently as an unfinished manuscript, (2) **WCO-Lite** an electronic online counterpart to WCO, and (3) **HarvEx**. It was **Norman Platnick** which came up with the *World Catalogue of Opiliones* form (as opposed to *Catalogue of Opiliones of the World* or *Catalogue of World Opiliones*) and the acronym WCO (pers. comm. 18 Nov. 2018), although he noted it also stands for something called the World Customs Organization.

The Microsoft Word **manuscript of WCO** has ca. 2000 pages. It has been originally created from (1) an exhaustive list compiled by Kury of all the literature

containing nomenclatural acts and geographic records of harvestmen; (2) importing Joel Hallan's checklist + Kury's 2003 New World Laniatores catalogue to serve as a scaffold. From there, the sources in "(1)" are being carefully combed and relevant data added to flesh out that scaffold. As a rule, past taxonomic decisions are retained while nomenclatural comments and mandatory corrections are added. It is a dynamic process as Kury himself is doing lots of research on the systematics of the order and publishing intermediate nomenclatural works such as a family-name catalogue (Kury 2018). There are a few **factors hampering a quick finish of WCO**: (a) obtaining obscure literature is not trivial, even with a decent network of arachno-friends and colleagues; (b) combing the early literature for nomenclatural acts can be tortuous, inconsistent, elusive and cryptic; (c) converting weird place names to recognizable modern toponyms takes time; (d) analysis of nomenclatural problems can be incredibly complex; (e) contemporary works never cease to pile up.

**WCO-Lite** (with the word "light" restyled as "lite") is a slenderer, more agile, online version of WCO, aiming to be a public-ready, authoritative reference for taxonomy of all subtaxa of harvestmen worldwide. WCO-Lite is being developed using an integrated web-based workbench for taxonomists called **TaxonWorks** (<http://taxonworks.org/>), created by **The Species File Group and many other contributors**. This PDF/A document is the **companion** text (for nomenclatural purposes) of the electronic version 1.0 uploaded to the internet.

This **Version 1.0 of WCO-Lite** contains a hierarchical arrangement of all **8707 valid nomina of taxa** in the arachnid order Opiliones, with author and year of publication, including suborders, infraorders, superfamilies, families, subfamilies, genera, subgenera, species and subspecies (more details in **Table 1**). Literature survey is meant to be **exhaustive up to December 2018**, and after that, other works are included as available, but coverage is not necessarily complete. Therefore, some papers up to early 2020 found their way here, especially papers

**Table 1.** Summary of number of **valid taxa** (both extant and fossil) by suborder contained in this **WCO-Lite Version 1.0**. Neither the total for Opiliones is equal to the sum of the subtaxa because of the taxa not included in any suborder (incertae sedis) nor the sum of all taxa here equals 8707 because there are intermediate ranks not in this table.

Suborder	valid families	valid subfamilies	valid genera	valid species	valid subspecies
Cyphophthalmi	6	3	40	229	13
Dyspnoi	11	4	49	409	25
Eupnoi	7	11	244	1806	78
Laniatores	39	63	1308	4189	129
Tetrophthalmi	1		2	2	
<b>Opiliones total</b>	<b>65</b>	<b>81</b>	<b>1645</b>	<b>6637</b>	<b>245</b>



published in leading journals such as Zootaxa, or widely advertised on ResearchGate.

The Fourth Edition of the International Code of Zoological Nomenclature is used here, and called simply the “Code”, and the articles are mostly cited as “Art.”. Here in WCO-Lite, mostly the bare minimum changes have been meant to be introduced, mainly to correct nomenclatural problems (see a synopsis in Table 2). Mandatory novelties established here include:

- (a) correction of grammatical inflections of adjectives, so that specific epithets match gender of generic nomina;
- (b) orthographical corrections in nomina formation as ruled by ICZN;
- (c) combinations of “orphan” species, which are currently assigned to invalid genera, with a valid genus nomen.

Since WCO-Lite is also a **vehicle for nomenclatural acts**, scattered synonymies and combinations including **external authors** were occasionally introduced here. To certify that due credit is given to those authors, those contributions are separately presented here in **Chapters numbered 4 to 12**.

Both in WCO and the WCO-Lite companion eBooks, a **hierarchical logonymy** is used as proposed by Dubois (2000), with **protonyms** (bulleted, no indentation), **aponyms** (indented, with colon) and **chresonyms** (more indented, with n-dash). That system is a refreshing departure from the chaotic mess of citations that is often and incorrectly called a “synonymy” in the literature. By using the **Dubois logonymy system**, the reader at a glance understands the taxonomic/nomenclatural structure of a given nomen. Here in this eBook **we keep it clean and simple**; only protonyms and some aponyms are deemed necessary to illustrate nomenclatural problems. There is no need to further clog the logonymy with numerous useless chresonyms.

Abbreviations of taxonomic and nomenclatural acts in Latin as used here are: translatio nova = **transl. nov.** (new transfer/assignment); combinātiō nova = **comb. nov.** (= new combination); combinātiō restaurata = **comb. rest.** (= reinstated or restored combination); nomen restitūtum = **nom. rest.** (= name revalidated or reinstated from synonymy), status promotus / status demotus = **stat. prom.** / **stat. dem.** (= promoted status / demoted status) synonymum novum = **syn. nov.** (= new synonym or new synonymy); synonymus resurrectus/restitutus = **syn. res.** (restored synonym). **It should be noted that comb. nov. and syn. nov. are not intended to express nomenclatural acts and their indication here is only meant to make taxonomic proposals clearer (Art.**

8.3). They should not carry unforeseen nomenclatural consequences such as inadvertently making nomina available.

**Why so many funny red things?** As a visual aid, to highlight categories and so that the readers can immediately grasp certain portions of the content, quite a few words in this eBook are marked with **visual enhancements**. The colored glyphs used in the setup of this work are: a bullet (•) denotes the **protonyms** (sensu Dubois 2000) and **unjustified emendations** (Art. 33.2.3); a globe icon (🌐) indicates general geographic distribution, a cat icon (🐱) denotes **type-species designations**, a gemini icon (♊) denotes statements of **homonymies and synonymies**; a scroll icon (📜) denotes miscellaneous nomenclatural notes (such as **incorrect spellings** and **incorrectly applied gender declinations**). Extinct taxa are marked with a traditional dagger (†). For those who frown upon the use of such symbols, let them be reminded that Roewer himself already in 1923 used similar visual resources, e.g., the sun symbol (☉) to denote unseen specimens.

## 1.1. Contents of future versions of this catalogue

We are already working to add many layers of relevant information to this project. As these are completed, new versions will be produced and released:

- (a) complete bibliographic sources and page numbers for all nomenclatural acts;
- (b) indication of type-species for all genera as well as designation mode; indication of type-specimens for all species with repositories (museum holdings);
- (c) cross-referenced invalid nomina (junior synonyms and homonyms);
- (d) cross-referenced ID for people, works, taxa, such as ORCID, Wikidata, Life Science Identifiers (LSID) of ZooBank for authors of works;
- (e) etymology and gender for all generic nomina; etymology and grammatical classification (if they are adjectives, participles, nouns) for all species epithets;
- (f) asserted geographic distribution of species;
- (g) indication of varied relationships among taxa;
- (h) alternative spellings and combinations;

**Table 2.** Organization of the sections 2 and 3 and chapters 4 to 13 (authored by external authors) in this eBook, their classification as nomenclatural or taxonomic matters and their relationship with the Articles of the Code when applicable.

Section	Subject of section	Classification of matters; articles
<b>Mandatory corrections</b>		
2.1	Grammatical adjective inflexions	Agreement in gender (Art. 31.2); Mandatory changes in spelling consequent upon changes in combination for species-group names (Art. 34.2)
2.2	Nomina incorrectly spelled based on mutilated adjectives	Assumption of Greek or Latin (Art. 26); Mandatory changes in spelling consequent upon changes in combination for species-group names (Art. 34.2)
2.3	Other issues of formation of nomina	Spellings that must be corrected (Art. 32.5); Unjustified emendation (Art. 33.2.3)
2.4	Replacement name for a secondary homonym	Secondary homonyms (Art. 57.3)
2.5	Synonymies originally proposed with inverted precedence	Priority applied to synonymy (Art. 23.3)
<b>Other nomenclatural and taxonomic acts</b>		
3.1	Genera hitherto without type-species	Genus-group names published after 1930 (Art. 13.3); Names published after 1999 not explicitly indicated as intentionally new (Art. 16.1); Names published after 1960 and treated as junior synonyms (Art. 11.6.3)
3.2	Valid species combined with invalid genera	Taxonomy
Ch 4	<i>Metaphalangium, Paropilio, Zachaeus</i>	Taxonomy
Ch 5	<i>Khazaddum</i>	Taxonomy
3.3	New synonymies, new allocations and new combinations	Taxonomy
Ch 6	<i>Acanthogonyleptes, Gonyleptes, Juticus, Gonyleptodes</i>	Taxonomy
Ch 7	<i>Tricommatum, Prosontes</i>	Priority applied to synonymy (Art. 23.3); Taxonomy
Ch 8	† Nemastomoididae, † Kustarachnidae, † Hastocularidae	Priority applied to synonymy (Art. 23.3); Taxonomy
Ch 9	Globipedidae	Taxonomy
Ch 10	Carunculata, Askawachidae	Taxonomy
Ch 11	<i>Liogyndulus, Acrographinotus, Parapachyloides</i>	Taxonomy
Ch 12	<i>Eubalta, Sadocus, Discocyrthus</i>	Taxonomy
Ch 13	<i>Bristoweia, Eopachylus, Gonyassamiinae</i>	Taxonomy
Ch 14	False <i>Gyndulus</i> , <i>Metagyndes</i>	Taxonomy



- (i) updates regarding the literature analysed;
- (j) migration of the database to the [Catalog of Life](#).

## 1.2. Conflicting rules from ICZN – inflection of specific epithets that are adjectives (ICZN Code Articles 26, 31, 32 and 34)

Let's start by citing a few articles of the International Code of Zoological Nomenclature (herein simply called the Code, ICZN (1999)):

**“26.** Assumption of **Greek or Latin** in scientific names. If the spelling of a scientific name, or of the final component word of a compound name [Art. 31.1], is the same as a Greek or Latin word, that name or that component is deemed to be a word in the relevant language unless the author states otherwise when making the name available.”

**“31.2.** Agreement in gender. A species-group name, if it is or ends in a Latin or latinized adjective or participle in the nominative singular, **must agree in gender with the generic name** with which it is at any time combined.”

**“31.2.2.** Where the author of a species-group name did not indicate whether he or she regarded it as a noun or as an adjective, and where it may be regarded as either and the evidence of usage is not decisive, it is to be **treated as a noun in apposition** to the name of its genus (the original spelling is to be retained, with gender ending unchanged; see Article 34.2.1).”

**“34.2.** Species-group names. The ending of a Latin or latinized adjectival or participial species-group name must agree in gender with the generic name with which it is at any time combined [Art. 31.2]; **if the gender ending is incorrect it must be changed accordingly** (the author and date of the name remain unchanged [Art. 50.3.2]).”

**“32.5.1.** If there is in the original publication itself, without recourse to any external source of information, clear evidence of an inadvertent error, such as a lapsus calami or a copyist's or printer's error, it must be corrected. **Incorrect transliteration or latinization, or use of an inappropriate connecting vowel, are not to be considered inadvertent errors.**”

Latin adjectives, in the same way as nouns, are inflected for gender into three forms: masculine, feminine and neuter.

First class adjectives have three endings in the nominative singular: mostly, they are inflected in the masculine (typically -us), the feminine (-a) and the neuter (-um). There is also the alternative ending in -er for the masculine. Examples: glaber, glabra, glabrum (hairless); niger, nigra, nigrum (black) and ornatus, ornata, ornatum (adorned, decorated).

Second class adjectives that have two endings have one form for the masculine and feminine, and a separate form for the neuter. The nominative singular ending for the masculine and feminine is -is, and the ending for the neuter is -e. Examples: inermis, inermis, inerme (unarmed, defenceless) and tenuis, tenuis, tenue (delicate, slender). There are exceptions, where the masculine ending is -er: silvester, silvestris, silvestre (wild); celer, celeris, celere (fast).

Third class adjectives with a single nominative ending for all genders. Examples: bicolor, bicolor, bicolor (having two colors, two-colored) and ferox, ferox, ferox (wild, fierce).

Authors of specific epithets who do not master Latin language (typically those who speak languages where adjectives have no genders, such as English) have often demonstrated at best an **incomplete, ultra-simplified view of gender matching**: everything masculine ends in -us, feminine in -a, neuter in -um. Therefore, several Latin adjectives were historically mutilated, creating anomalous aberrations.

Examples of mock-feminine and mock-masculine inflections:

*Paraconomma ovala* — Here we have two mistakes combined. *Paraconomma* is neuter; the authors mistook it for feminine probably because the ending in -a and created an inexistent mock-feminine inflection where it should have been ovale (3rd decl., two endings).

*Cosmetus pulchrus* — *Cosmetus* is masculine; author created an inexistent mock-masculine inflection where it should have been pulcher (1st decl., three endings).

In all cases above there are two **possible interpretations**, and strictly speaking, number 2 as used by Kury in the past does not hold under the present edition of the Code. The Code (ICZN 1999) still does not have a much needed article allowing for the correction of these mis-inflected or mis-constructed words:



The “cold hard” view (e.g., Alonso-Zarazaga, pers. comm., 2020): words such as *atrus*, *ovala* and *pulchrus* do not exist in Latin (Art. 32.5.1). Therefore they cannot be “guessed” as adjectives in spite of them being obviously mis-inflected adjectives. They should be considered as invariable nouns in apposition (Art. 31.2.2).

The “stretched” view (e.g., Kury 2003): words such as *atrus*, *ovala* and *pulchrus* are clearly crooked inflections of well-known adjectives, and we should not “look away” pretending we do not know what they are, just because they are not 100% “the same” (as in Art. 26). They should be treated for what they are, adjectives, and submitted to Latin grammar rules (Art. 34.2).

There is also a **third possibility**, which should be exercised with extreme caution:

The “non-conformist” view (as proposed herein): Although the current version of the ICZN Code does not fully support such corrections, while the ICBN (International Code of Nomenclature for algae, fungi, and plants) already and wisely does, we are doing them “extra-officially” anyway in the hope that ICZN soon introduces in its next versions the possibility of “officially” fixing such aberrations. If not, we can always try to make them qualify as **prevailing usage** thanks to the influence of this very catalogue and in 50 years those corrected forms can be the norm.

### 1.3. Author citation: personal names

Here in the WCO Project we unify spellings of author names across all of their publications regardless of the original form used. Eventually, all authors of WCO-Lite will be linked to permanent online IDs such as in ZooBank and Wikidata, so retrieving relevant information is maximized.

**Why unifying names?** In taxonomy, the author of a scientific name of a taxon is the person (or team) who first makes it available in the sense of the Code (many names are made available but formed incorrectly). That is most usually the author of the work where the nomenclatural act was published, but sometimes only a fraction of the authors of the work are responsible for naming the taxa and sometimes even an external person who does not have anything to do with the authorship of the work is deemed to be author of a taxon.

Names change. Every year we hear about a “new” country or province. Locality names often change, not

only small villages, not to mention misspelled names. **People’s names also change.** Motives vary. Sometimes it is an orthographic reform, or political influences that lead the author to use a specific language, or someone finds their name extremely common and wants to be recognized, people marry or they are Hispanophone, with special formatted names.

**There is no rule for standardization of author names in citations**, but as common practice in zoology, they are taken straight *ipsis litteris* from the publication. That means, if the name of the author appears differently in different publications, catalogers should write it differently every time. This makes sense if the reader needs to locate the relevant work. Not so much in an electronic catalogue. Especially if we aim to retrieve information about the author. The indexing systems today are unable to recognize if it is the same person if the name changes only a bit, especially now that we have unique identifiers for authors such as in ZooBank. This reverence for the original form of the name if taken to the extreme could lead to absurdities such as: one cannot cite “Adriano Kury (2007) did x” because that 2007 paper has “AB Kury” instead of Adriano Kury, therefore Adriano does not exist in AB Kury 2007.

As aptly put by others:

*“From the broader perspective I personally feel it’s better to **move towards unified approaches**. While that’s not saying what the approach is, it is suggesting that TaxonWorks will “encourage” people to forgo past idiosyncrasies so that future meaning is enhanced throughout the field. Formatting people’s names seems to be just such a case. Because we can hyperlink to the person, and ultimately show the full info for that person, we can check quickly in those (very few relatively speaking) edge cases where names are ambiguous between people.”* [TaxonWorks creator, Matthew Yoder, in litt. 2020].

*“Using URLs as unique identifiers (as Wikidata and many others do) is convenient in the short-term, but not wise for the long-term. I prefer using identifiers like **UUIDs as true “identifiers”**, then embed them within URLs as appropriate to represent resolvable/actionable identifiers.”* [Richard L. Pyle, Bernice Pauahi Bishop Museum, in litt. 2020].



### 1.3.1. POSITION OF PATERNAL SURNAME AND PERSONS WHO CHANGED NAMES DURING THEIR CAREER

Citations are the backbone of a scientist's survival. Authors are often led to change their names during their careers to ensure more accurate claim to their own work. The most common cases that harm authors' ability to gather their work together are: (1) when information retrieval systems have problems identifying the correct indexable surname from among the author's names and (2) when one's surname is extremely common and carries little discriminatory value.

How to proceed in such cases? Should we use different names for the same person according to the publications? Should we **retcon** (give **retroactive continuity** to) everything, ascribing a **standard name** to the person that we know is the same?

Welter-Schultes tackled this problem by saying: "The spelling of the author in a name of a taxon has never been a problem, until the age of electronic information where it has become necessary to connect information derived from various independent sources. Presence of various **different spellings** of authors in names of taxa (including the use of initials) **provide a serious obstacle for connecting information on species....** The situation in zoology is very complex, many authors were spelled in different ways, bibliographers and taxonomists are undecided which ones should be regarded as the preferred names." In spite of all these considerations, he ends up proposing that: "The spelling of the author(s) in a name of a taxon must be taken from the work in which the available name was established."

We herein decided to use an analogy taken from the Code itself. Article 11.9.3.2 says: "a species-group name is deemed to have been published in combination with the correct original spelling of the generic name, even if it was actually published in combination with an emendation or incorrect spelling of the generic name". The way we see it, we define a "correct spelling" for an author's name, and all other apparitions of this name

under varied shapes are retroactively deemed to be only incarnations of the same name.

After seeing nothing less than six different shapes that the name of Osvaldo Villarreal-Manzanilla took over his career (Fig. 2), we were definitely swayed to the dark side of the retcon. So, here, in the WCO project, whenever authors have their name changed mid career (1) because of different transliterations, (2) because of marriage, (3) to avoid confusion by being a Spanish maternal surname in the "wrong" place, or (4) just because the paternal surname is too common, they are all nevertheless standardized. This view is not universally spoused and maybe even loathed, but it has the immense advantage of allowing an author to be recognized as the same person for statistics, regardless of how his name is presented in press. Moreover, as WCO is organized, a person's name is an entity in a database, which is linked to a network of papers and taxa (see intro of this section 1.3). Alternative spellings/names would damage this information retrieval system.

Herein we treat **married ladies** with two name standards by the maiden name. As examples we have İlkay Çorak which gained the married surname Öcal and Ana Lúcia M. Tourinho, who married, adding the extra surname Davis, but subsequently divorced, returning to simply Tourinho. Those ladies are herein referred to as **Çorak, İ.** and **Tourinho, A.L.M.** As for ladies already married when started to publish, we just kept the husband's name such as in **Helia Soares**. We found that a powerful tool to improve easy retrieval of the same author with different names is registering as many **different aliases** as possible with **ZooBank**. We are using LSID for all authors in our database. Of course authors may choose whatever names they fancy; we are referring only to **our** use in the WCO Project. We would never follow the example of Eschmeyer, who once demanded that for authors of fishes, women be forced to publish eternally under the name of their divorced husband.

Information retrieval systems treat personal names as if they all belonged to the standard Western culture, where we have this pattern: First/given name + Middle/maternal surname + Last/Family/paternal surname.

**Table 3.** Frequency of personal surnames relevant for the present discussion (source: <https://forebears.io/surnames>). Indistinct, extremely common names compared to a moderately common name such as Soares and the much rarer name Sabattini.

Surname	nth most Common surname in the World	How many people bear this surname
<b>Zhang</b>	3rd	98,138,757
<b>da Silva</b>	47th	10,452,759
<b>González</b>	60th	8,753,528
<b>Silva</b>	103rd	5,449,154
<b>de Souza</b>	158th	3,458,455
<b>Soares</b>	278th	1,944,683
<b>Sabattini</b>	120,343rd	3,813

**American:** first name sometimes is doubled; middle name is almost always abbreviated (rarely spelled out, the entire form virtually unknown), rarely lacking = William A. Shear, Thomas S. Briggs, Clarence James Goodnight, which would be cited as: Shear WA, Briggs TS, Goodnight CJ. There is a more recent trend, where after marriage some authors hyphenate their birth last name with their spouse's, both in males and females (or same sex marriage).

**European:** given name often abbreviated (sometimes given names are multiple, especially in older names); middle name often lacking; last name sometimes is composed = Carl Ludwig Doleschall, Jürgen Gruber, A.S. Hirst, Jochen Martens, Octavius Pickard-Cambridge.

**Chinese:** family name comes first, and given name is often composed of two hyphenated parts = Chen Ssu-Li, Lian Wei-Guang, Shih Hsi-Te, which are here cited as Chen S-L, Lian W-G, Shih H-T.

**Russian:** the maternal surname is always lost when ladies marry, therefore they do not propagate to the new generation and are never used as surnames; there is also a patronymic (absent in other cultures), always abbreviated = Aleksey N. Chemeris, Dmitriy E. Kharitonov, Nataly Y. Snegovaya.

**Italian:** last name sometimes has prepositions, which should remain = Lodovico di Caporiacco. Indexed herein as di Caporiacco, L.

**Portuguese/Brazilian:** usually follow the American pattern, there is a frequent preposition "de" (which may be combined with definite articles "a, o, as, os"), they should not be indexed by preposition = Ludson Neves de Ázara, Cândido Firmino de Mello-Leitão, cited herein as Ázara LN de, Mello-Leitão CF de.

**Salvador de Toledo Piza Júnior:** this Brazilian Portuguese name has a compound non-hyphenated paternal name and a suffix. This author in arachnology is universally called Piza; the preposition "de" is discarded. Curiously, in entomology he is most often called Toledo Piza. Herein we use Piza, ST.

**Beatriz Santos de Souza:** this Brazilian author, as a member of a new generation, more alert to indexation issues and to the multiple homonym zoologists, decided from the start to use her paternal great grandmother's surname Sabattini instead of de Souza (compare incidence of both names in Table 3). Herein we use Sabattini, BS.

**Brazilian called da Silva (by far the commonest surname in Brazil) or Silva (also very common, see Table 3):** they either merge the preposition into the name (creating an otherwise non-existing name) or hyphenate

maternal and paternal names (often they do it mid-career when they realize their names are lost in an ocean of Silvas) = Márcio Bernardino DaSilva, Marília Pessoa-Silva. Whenever they change names mid-career they are retconned herein.

**Romanian:** the name Filimon Cîrdei after the 1953 orthographic reform replaced Cârdei (which occurred scarcely in the harvestmen output of this author); whenever the work is in Russian it was transliterated as Кырдеи (not Romanian Cyrillic, which was officially abandoned in 1860). Herein we use Cîrdei, F.

**Hispanophone (both European and South American):** the given name comes first and sometimes is doubled such as José Carlos. The paternal surname comes first, in the middle. The last name is usually the maternal surname (often excluded to avoid confusion) = as for example Gonzalo Giribet de Sebastián > Gonzalo Giribet, Andrés García Rincón > Andrés García; Miguel Medrano Leal > Miguel Medrano. A minority of authors, mostly in South America, use both surnames artificially connected by a hyphen to ensure the father's name is used in indexation = Manuel Ángel González-Sponga, Izaskun Merino-Sáinz.

**Carlos González Luque:** this Spanish name used to appear entire, but later the author chose to use a strange abbreviation of the paternal name, hardly obvious for a non-Spanish speaker: Carlos Glez. LUQUE. Finally, he decided to adopt the maternal surname and use only the initial of the paternal name (perhaps because González is an extremely frequent surname, see Table 3): Carlos G. Luque (Fig. 1). Herein we use Luque, CG.

**Hispanophone with father's and mother's names initially isolated and later in the career hyphenated:** Abel Pérez-González, Osvaldo Villarreal-Manzanilla. Those were variously cited in the literature under diverse monikers, take this example and we rest our case: (1) Osvaldo Villarreal, (2) Osvaldo M. Villarreal, (3) Osvaldo Villarreal M, (4) Osvaldo Villarreal Manzanilla, (5) Osvaldo Villarreal-Manzanilla. These names are all retconned herein, respectively to Pérez-González, A and Villarreal, O. We use here Villarreal as the perfect example of an indexation nightmare (Fig. 1). We wonder how this author manages to get his h-index calculated.

### 1.3.2. PERSONS SHARING A SURNAME – HOW TO DISTINGUISH THEM?

Usually, initials of given names are not included in names of taxon authors. However, in a few cases they are used in WCO to avoid ambiguity. **The TaxonWorks platform currently does not admit collectively labeling authors with initials:** these are always dropped when associated with taxon names (the underlying rationale is



## Descripción de *Trinella vigirima* sp. n. (Opiliones: Agoristenidae) de Venezuela

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Revalidation of *Obidosus* Roewer, 1913 and description of two new cave-dwelling species of *Protimesius* Roewer, 1913 from Brazil (Opiliones: Stygnidae)

Oswaldo Villarreal, Ludson Neves de Ázara & Adriano Brilhante Kury

**Rowlandius dumitrescoae** species group: new diagnosis, key and description of new cave-dwelling species from Brazil (Schizomida, Hubbardiidae)

Alessandro Ponce de Leão Giupponi<sup>1</sup>, Gustavo Silva de Miranda<sup>2</sup>,  
Oswaldo M. Villarreal<sup>3</sup>

Una nueva especie del género *Paravima* Caporiacco, 1951 (Opiliones, Laniatores, Agoristenidae) del tramo oriental de la cordillera de La Costa, Estado Miranda, Venezuela

Oswaldo Villarreal-Manzanilla y Carlos DoNascimento

ARQUENAS / Fauna Ibérica Subterránea: Cantabria

CARLOS GLEZ. LUQUE

**Los *Ischyropsalidoidea*  
de la Cornisa Cantábrica:  
Cantabria**

Три новых вида рода *Phalangium* Linnaeus, 1758 (Arachnida: Opiliones: Phalangidae) из Армении и Узбекистана

A.N. Tchemeris  
A.H. Чемерис

**New data on the harvestman genus *Nemaspela*  
Šilhavý, 1966 (Arachnida: Opiliones)**

Aleksey N. Chemeris  
Department of Invertebrate Zoology,  
Faculty of Biology and Soil Sciences,  
Tomsk State University,  
Lenin Av. 36, Tomsk 634010, Russia

Taxonomic notes on *Acanthomegabunus* Tsurusaki, Tchemeris & Logunov 2000 (Arachnida: Opiliones: Phalangidae), with a description of the new species *A. altaicus* sp. n. from the Altai Mountains of Russia and NE Kazakhstan

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VORLÄUFIGE MITTEILUNG ÜBER OPILIONEN-FAUNA  
VON RUMÄNIEN

von  
F. CÂRDEIU

CONTRIBUȚII LA CUNOAȘTEREA OPILIONIDELOR  
DIN ÎMPREJURIMILE MASIVULUI PIATRA CRAIULUI  
DE  
F. CÎRDEI

К ИЗУЧЕНИЮ СЕНОКОСЦЕВ (*OPILIONES*) ИЗ СЕВЕРО-ЗАПАДНОЙ  
ЧАСТИ РНР И ВЕРХНЕГО РЕЧНОГО БАСЕЙНА ПРУТА

Часть I \*)  
Ф. КЫРДЕЙ

NOI CONTRIBUȚII LA STUDIUL PHALANGHINELOR  
(SUBFAM. PHALANGHINAE, ORD. OPILIONES)  
DIN CARPAȚII ORIENTALI  
DE  
F. CÂRDEI și F. BULIMAR

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**A new cave-dwelling endemic *Ischyropsalis* C.L. Koch, 1839 (Opiliones: Dyspnoi: Ischyropsalidae) from the karstic region of Cantabria (Spain)**

CARLOS G. LUQUE & LUCIA LABRADA  
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**CONTRIBUCION AL CONOCIMIENTO DE LOS OPILIONES EPIGEOS E HIPOGEOS DE CANTABRIA (ESPAÑA). I APORTACIONES AL CATALOGO (ARACHNIDA: OPILIONES: PALPATORES)**

Carlos González Luque

Sección de Ciencias del C.A.E.A.P. y  
G.E.I.S. C/R., Apdo. de Correos 879,  
39080 Santander (España).

**Figure 1.** Examples of the varied spellings of the personal names of Aleksey Chemeris, Filimon Cîrdei, Carlos Luque and Oswaldo Villarreal in their own papers.

that no one needs initials because the individual author sheets are already linked, so there is no confusion as to who is who).

**Koch:** besides father and son Carl Ludwig Koch and Ludwig Carl Christian Koch, there was another contemporary unrelated (but also quasi-homonym) Koch: Carl Jakob Wilhelm Ludwig Koch. They are traditionally (and here in WCO) indicated along with some selected initials as: Carl Jakob Wilhelm Ludwig Koch = C. Koch; (2) Carl Ludwig Koch = C.L. Koch; (3) Ludwig Carl Christian Koch = L. Koch.

**Goodnight:** husband and wife Clarence James and Marie Louise Goodnight almost always (except for Goodnight & Goodnight 1976) published together and in the same sequence, so, denoting each of them by the initials would be mostly superfluous.

**Pickard-Cambridge:** uncle and nephew with similar names, Octavius and Frederick Octavius Pickard-Cambridge are to be distinguished by adding their initials.

**Soares:** husband and wife Benedicto Abilio Monteiro Soares and Helia Eller Monteiro Soares published in four different arrangements: Benedicto alone; Helia alone; Benedicto + Helia and Helia + Benedicto. In this case we in opilionology traditionally distinguish them as B. Soares and H. Soares.

### 1.3.3. USAGE OF LATIN EXTENDED ALPHABET

In older literature, especially **before the advent of UNICODE** and word-processing typing facilities, names of Brazilian, Czech, Serbian, Danish, Polish authors were **often adapted, mutilated, stripped of the diacritics**, as for example: Mello-Leitao, Silhavy, Hadzi, Sorensen, Starega. Sometimes diacritics do not appear in upper-case letters as in titles of works. Even the extremely frequent name Roewer appeared in the literature also as “Röwer”. Sometimes, names appeared declined as Wajgla, Leopolda for Wajgiel, Leopold. Here in WCO Project, the **extended Latin alphabet** is used to faithfully reproduce the original names. Examples:

(a) Czech = Bartoš; Bezděčka; Frič; Klimeš; Kratochvíl; Růžička; Šilhavý; Zavřel.

(b) Danish = Sørensen.

(c) French = Dugès; Guérin-Ménéville.

(d) German = Kästner; Müller.

(e) Latvian = Spuņģis; Tumšs.

(f) Norwegian = Solhøy.

(g) Polish = Kulczyński; Rozwałka; Staręga.

(h) Portuguese = Mello-Leitão.

(i) Romanian = Cîrdei; Roșca.

(j) Serbo-Croatian = Babić; Hadži; Mučalica.

(k) Spanish = González; Muñoz-Cuevas.

(l) Turkish = Çorak; Danişman; Yiğit .

(m) Bilingual countries = political constraints may lead to a change in the spelling of a name. Catalan is the most used language in Catalonia, but was not allowed to be used or written during the Franco dictatorship, thus many Catalan authors use their Catalan name even though their official name is different. This is why some of the earlier work of Maria Rambla spelled María in Spanish (with a acute), but later it was spelled in Catalan, without acute.

### 1.3.4. AUTHOR NAMES WRITTEN ORIGINALLY IN NON-LATIN SCRIPTS

WCO uses **Hepburn romanization** (Hebon-shiki) for **Japanese** characters employed in Japanese language and **BGN/PCGN Romanization** (United States Board on Geographic Names & Permanent Committee on Geographical Names for British Official Use) for the **Cyrillic script** as employed in modern **Russian**. There is a single exception here: the Cyrillic letter Ё ё (iotated vowel), which sounds more or less like [yo] in yo-yo. The BGN/PCGN system uses an unexplainable “yë”, but this is here transliterated herein as “yo”.

Examples of transliteration adopted in the project WCO :

仲辻耕治 (なかつじ こうじ) = never originally written in Latin script; it has been elsewhere romanized as Nakatsudi Kodi. Herein we use Nakatsuji Kōji, abbreviated as **Nakatsuji K.**

三好保徳 (みよしやすのり) = never originally written in Latin script; it has been elsewhere romanized as Miyosi Yasunori. Herein we use Miyoshi Yasunori, abbreviated as **Miyoshi Y.**

თამარ სევერიანეს ასული მხეიძე (Tamar Severianes asuli Mkheidze) = mostly written in original Georgian; often transcribed in Cyrillic as T. C. Мхеидзе (Tamara Severyanovna Mkheidze), it has been elsewhere romanized as Mcheidze. Herein we use Tamara S. Mkheidze, abbreviated as **Mkheidze TS.**



**Алексей Николаевич Чемерис** = this has been originally used in Cyrillic, and also transcribed by himself either as Chemeris or Tchemeris (as the author currently prefers). Herein we use Aleksey N. Chemeris.

**Борис Пименович Чевризов** = this has been originally used in Cyrillic, and also transcribed by himself as Chevrizov (not in paper titles). Herein we use Boris P. Chevrizov.

**Николай Иосифович Гриценко** = never originally written in Latin script; it has been elsewhere romanized as Gricenko. Herein we use Nikolay I. Gritsenko.

**Станислав Иванович Лёвшин** = never originally written in Latin script; it has been elsewhere romanized as Ljovuschkin. Herein we use Stanislav I. Lyovushkin.

**Дмитрий Евстратьевич Харитонов** = never originally written in Latin script; it has been elsewhere romanized as Charitonov. Herein we use Dmitry E. Kharitonov.

**Владимир В. Редикорцев** = originally used in Cyrillic, and also romanized by himself as Redikorzev (and Redikorcev elsewhere). Herein we use Vladimir V. Redikortsev.

### 1.3.5. SUMMARY OF CATEGORIES OF AUTHOR NAMES AS USED HEREIN (WITH EXAMPLES)

**Type 1a (“European-style”, ideal for indexation):** binominal, paternal only; maternal originally or artificially not used = Banks N; Gervais P; Gruber J; Hedin M; Kirby W; Martens J; Novak T; Say T; Simon E; Wijnhoven H.

**Type 1b:** same as 1a, but given names multiple = Doleschall CL; Goodnight CJ, Hara MR; Koch CL.

**Type 1c:** same as 1b, but given names hyphenated = Roewer C-F.

**Type 1d (“Russian-style”):** maternal name absent; patronymic always abbreviated, as a mock-middle-name = Chemeris AN; Kharitonov DE.

**Type 2 (“American-style”):** trinominal, middle-name (often maternal surname) abbreviated = Briggs TS; Cokendolpher JC; Kury AB; Mendes AC; Shear WA.

**Type 3:** maternal name absent; paternal name compound or augmented by other family name; originally or artificially hyphenated = Guérin-Ménéville FE; Mello-Leitão CF de; Pickard-Cambridge FO; Pinto-da-Rocha R.

**Type 4a:** binominal, Spanish with maternal discarded (sometimes real-life given name multiple simplified, also discarded) = García A; Giribet G; Medrano M; Rambla M; Villarreal O.

**Type 4b:** same as 4a, but original multiple given names remain = Acosta LE; Capocasale RM; Maury EA.

**Type 5a:** Spanish, with paternal + maternal artificially hyphenated (to enhance distinction or to ensure the paternal name was chosen for indexation) = Cruz-López JA; Franganillo-Balboa P; González-Sponga MA; Merino-Sáinz I; Muñoz-Cuevas A; Pérez-González A.

**Type 5b:** same as 5a, but Portuguese, with maternal + paternal artificially hyphenated (to enhance distinction) = Coronato-Ribeiro A; Pessoa-Silva M.

**Type 6:** Portuguese or Spanish, paternal name has preposition de (+ article), which is artificially accreted (to enhance distinction): DaSilva MB; DoNascimento C.

**Type 7a:** binominal Japanese and Chinese inverted = Suzuki S; Tsurusaki N; Zhang C.

**Type 7b:** same as 7a, but with Chinese composed names hyphenated = Chen S-L, Lian W-G, Shih H-T.

**Type 8:** miscellany of paternal names entirely or partially discarded = Luque CG; Piza ST.

## 1.4. Opening a can of worms: Identity of authors of taxa (Code Art. 50)

The Code Article 50 deals with authors of names and nomenclatural acts. However, it has many imprecisions, and **interpretation is not straightforward**. The author of a name or nomenclatural act is the person who first publishes it in a way that satisfies the criteria of availability, then again, publication in the meaning of the Code is a primary criterion of availability.

Art. 50.1.1. says: if **it is clear from the contents** that some person other than an author of the work is alone responsible both for the name or act and for satisfying the criteria of availability other than actual publication, then that other person is the **author of the name or act**.

The root of the problem here seems to be the expression “if it is clear”. What is it supposed to mean? The official interpretation of the Code on this matter (Sabrosky 1974) basically disavows any indications of authorship by an external party, which is in stark



**Figure 2.** Form in ZooBank: example of how trivial is it to have the authorship of nomina different from the authorship of the work.

contrast with the usual practice in Zoology (or at least in the harvestmen literature). We could easily include ourselves in this characterization: “Because most modern authors do not know this meaning of “author” in the Code’s sense, and they attribute their new names to incorrect authors.” [Welter-Schultes].

The ICZNwiki (<http://iczn.anasp.org/wiki/Article50>) has a great deal of comments on this article and its consequences for nomenclature, all comments are authored by **Francisco Welter-Schultes**, and in their lucidity they are herein considered as the prime guideline to our own interpretations. Furthermore, for the most difficult cases we invoked the invaluable assistance of **Francisco Welter-Schultes** himself to provide an analysis.

#### 1.4.1. A NAME IS ATTRIBUTED TO A SUBSET OF A MULTI-AUTHORED WORK

Earlier, most taxonomic works were authored by a single person, sometimes by two. The most prolific opil-io-author, Roewer, for example, never had a co-author. Nowadays, five or six authors are not infrequent, especially when distinct parts of the project are carried out by different people, all deserving authorship. In many cases only some of the work’s authors are responsible for the formal establishment of a taxon. This is something trivial, straightforward (Fig. 2). All cases listed in this section have authorship herein recognized exactly as it appeared in the original publications.

As a recent example in Opiliones, in a work of 2016 authored by Katya R. Jay, Zachary R. Popkin-Hall, Michelle J. Coblens, Jill T. Oberski, Prashant P. Sharma & Sarah L. Boyer, we have species described by different subteams, each case explicitly indicated:

- (a) Jay, Coblens & Boyer in Jay et al. 2016;
- (b) Jay, Oberski & Boyer in Jay et al. 2016;
- (c) Jay, Popkin-Hall, Coblens & Boyer in Jay et al. 2016;
- (d) Popkin-Hall, Jay & Boyer in Jay et al. 2016.

**Welter-Schultes example:** In 2011 Yanes and six co-authors published an article on five new species of the gastropod genus *Napaeus*. Next to the verbal descriptions the article contained also detailed tables in

which the morphological data of the new species were compared. The first new species was attributed to co-authors 2, 6 and 7, the third new species to co-authors 3, 4 and 5. The co-author who compiled the table was not specified, so in the absence of the clearcut attribution the table was under the responsibility of all seven co-authors. Since the table belonged to the descriptions, the attributed co-authorships were not representing the true responsibilities for the descriptions. They are accepted because the article was published after 1999.

Similar cases in Opiliones:

#### **Cyphophthalmi Simon, 1879**

##### ***Iberosiro rosae* Giribet, Merino-Sáinz & Benavides, 2017**

- *Iberosiro rosae* Giribet, Merino-Sáinz & Benavides **in** Giribet et al. 2017: 465.

##### ***Paramiopsalis anadonae* Giribet, Merino-Sáinz & Benavides, 2017**

- *Paramiopsalis anadonae* Giribet, Merino-Sáinz & Benavides, **in** Giribet et al. 2017: 469.

##### ***Paramiopsalis ramblae* Benavides & Giribet, 2017**

- *Paramiopsalis ramblae* Benavides & Giribet, 2017 **in** Giribet et al. 2017: 475.

##### ***Siro ligiae* Giribet, 2017**

- *Siro ligiae* Giribet, 2017 **in** Giribet et al. 2017: 479.

##### ***Siro richarti* Benavides & Giribet, 2017**

- *Siro richarti* Benavides & Giribet, 2017 **in** Giribet et al. 2017: 484.

#### **Dyspnoi Hansen & Sørensen, 1904**

##### ***Calathocratus hirsutus* Snegovaya, 2011**

- *Calathocratus hirsutus* Snegovaya **in** Snegovaya & Chumachenko 2011: 117, figs 2–14.

##### ***Calathocratus minutus* Snegovaya, 2011**

- *Calathocratus minutus* Snegovaya **in** Snegovaya & Chumachenko 2011: 117, figs 15–27.

##### ***Caucnemastoma martensi* Snegovaya, 2011**

- *Caucnemastoma martensi* Snegovaya **in** Snegovaya & Chumachenko 2011: 119, figs 37–47.



### *Giljarovia kratochvili* Snegovaya, 2011

- *Giljarovia kratochvili* Snegovaya in Snegovaya & Chumachenko 2011: 119, figs 28–36.

### *Oskoron* Shear, 2016

- *Oskoron* Shear in Shear & Warfel 2016: 56. 🐞 Type-species by original designation: *Taracus spinosus* Banks, 1894. 📖 This nomen is a **misspelling** of Οσορκων (Osorkon), a Hellenized name of Ancient Egyptian pharaoh *User-maat-ra Userken*, also known as Osorkon III, of Dynasty XXIII. However, as there is no evidence of an inadvertent error in the original publication itself, it should be left uncorrected (Code Art. 32.5.1).

### *Oskoron brevichelis* Shear, 2016

- *Oskoron brevichelis* Shear in Shear & Warfel 2016: 62, figs 1, 142–153.

### *Oskoron crawfordi* Shear, 2016

- *Oskoron crawfordi* Shear in Shear & Warfel 2016: 65, figs 154–163.

### *Taracus audisioae* Shear, 2016

- *Taracus audisioae* Shear in Shear & Warfel 2016: 50, figs 106–115, map 7.

### *Taracus carmanah* Shear, 2016

- *Taracus carmanah* Shear in Shear & Warfel 2016: 34, figs 45–52, map 2.

### *Taracus fluvipileus* Shear, 2016

- *Taracus fluvipileus* Shear in Shear & Warfel 2016: 40, figs 68–76, map 4.

### *Taracus marchingtoni* Shear, 2016

- *Taracus marchingtoni* Shear in Shear & Warfel 2016: 36, figs 53–67, map 4.

### *Taracus spesavius* Shear, 2016

- *Taracus spesavius* Shear in Shear & Warfel 2016: 45, figs 89–93, map 6.

### *Taracus taylori* Shear, 2016

- *Taracus taylori* Shear in Shear & Warfel 2016: 42, figs 77–88, map 6.

### *Taracus timpanogos* Shear, 2016

- *Taracus timpanogos* Shear in Shear & Warfel 2016: 47, figs 94–105, map 6.

### *Taracus ubicki* Shear, 2016

- *Taracus ubicki* Shear in Shear & Warfel 2016: 53, figs 116–125, map 7.

### Eupnoi Hansen & Sørensen, 1904

### Laniatores Thorell, 1876

### *Eulibitia victoriae* (Pinzón-M. & Townsend, 2017) **comb. nov.**

- *Platymessa victoriae* Pinzón-M. & Townsend in Pinzón-M., Townsend & Martínez-H. 2017: 61, figs 1–7.

**Comment.** This species was originally described in the genus *Platymessa* Mello-Leitão 1941, which is currently a junior synonym of *Eulibitia* Roewer, 1912, as established almost simultaneously by Medrano & Kury (2017). We do not think that this species fits exactly in Medrano & Kury diagnosis of *Eulibitia*, but for now, we



**Figure 3.** Snapshots of Roewer (1923) clearly showing measurements, a diagnosis and illustrations, all of which give availability to *Ischyropsalis taunica*, and all taken from Müller, with the sun symbol (☉) clearly apparent in the “type data” section, meaning that Roewer himself had not seen the material, which was in Müller’s collection in Frankfurt. The footnote 2 itself is of no consequence for our present analysis.



follow Pinzón-M. et al. decision, pending further study on those NW South American Cosmetidae.

### **Cryptomastriidae Derkarabetian & Hedin, 2018**

- Cryptomastriidae Derkarabetian & Hedin **in** Derkarabetian et al. 2018: 13.

### **Cryptomaster behemoth Starrett & Derkarabetian, 2016**

- *Cryptomaster behemoth* Starrett & Derkarabetian, **in** Starrett et al. 2016: 23.

### **Gnomulus baharu Schwendinger, 1998**

- *Pelitnus thorelli* Schwendinger 1992: 180, figs 14–26. **II** Junior primary homonym of *Pelitnus thorelli* Sørensen, 1932.
- *Gnomulus baharu* Schwendinger **in** Martens & Schwendinger 1998: 527. **I** Nomen novum for *Pelitnus thorelli* Schwendinger, 1992.

### **Paratakaoia minima Suzuki, 1986**

- *Paratakaoia minima* Suzuki **in** Suzuki & Stone 1986: 125, figs 1–9.

### **Stenophareus guerreroi Villarreal & DoNascimento, 2007**

- *Stenophareus guerreroi* Villarreal & DoNascimento, 2007 **in** Villarreal-M et al. 2007: 46, figs 8–14, 18–20.

### **Tingomaria Mello-Leitão, 1949**

- *Tingomaria* Mello-Leitão **in** Mello-Leitão & Feio 1949: 318. **I** Type-species by original designation: *Tingomaria hydrophila* Mello-Leitão, 1948.

### **Tingomaria hydrophila Mello-Leitão, 1949**

- *Tingomaria hydrophila* Mello-Leitão **in** Mello-Leitão & Feio 1949: 319.

## **1.4.2. A PERSON EXTERNAL TO THE WORK IS DEEMED TO BE AUTHOR OF A TAXON (CODE ART. 50.1.1)**

This case more often happened when someone retook abandoned projects or notes left by a deceased or departed colleague. Or else, someone contributed a punctual observation or conclusion to a larger paper. Here the **usual practice** among opilionologists **clashes** with a cold application of the Code. There are many examples of this in the opilionological literature.

### **1.4.2.1. Successful different-authorship attributions**

An iconic case is that of Kai Henriksen, who organized and published (1932) many annotations left prior by Sørensen when he died in 1916. The title of the work is ambiguous, and it is often attributed to Sørensen in

the literature. Our understanding here is that the work in question is authored by Henriksen, with many of the taxa authored by Sørensen, and some occasional ones explicitly attributed to Henriksen.

**Welter-Schultes example:** In 1846 Philippi established a new gastropod species *Bulimus cretensis* and attributed the name to “Pfr.” (= Pfeiffer) in the headline. A Latin diagnosis followed, then a locality, and then a German descriptive text. Behind all this in the last line was quoted “(Pfr.)”, expressing that Pfeiffer alone and not Philippi was responsible for the entire description. The name must be attributed to Pfeiffer.

Similar cases in Opiliones:



**Case taunica:** The name *Ischyropsalis taunica* was made available because a description was given: Roewer (1923) effectively published this species (with a diagnosis and figures all attributed to Müller). The posterior interpretation of Schönhofer (2013) that *I. taunica* was a nomen nudum is incorrect. For being a nomen nudum it would have been necessary that Roewer added an explicit disclaimer that the name should not be regarded as new here. Roewer originally attributed the authorship to Müller, stating that he knew of a manuscript containing the description: he wrote besides the species name “MÜLLER MS.” and in a footnote “Die Diagnose dieser Art nebst Abbildung wurde mir im März 1922 vom Autor als MS mitgeteilt; sie soll in nächster Zeit in den Zool. Jahrbüchern veröffentlicht werden.” The fact that Roewer stated that Mueller intended to publish another paper has no influence here. Most importantly, Roewer used his standard abbreviation of the sun symbol (☉) to indicate that **he had not seen the specimens**. That means that **all descriptive information came from Müller**. So in this case, the authorship should be attributed to Müller.

### **Ischyropsalis hellwigii hellwigii (Panzer, 1794)**

- *Phalangium Hellwigii* Panzer 1794: XIII, 18, unnumbered plate.
- *Ischyropsalis taunica* Müller **in** Roewer 1923: 683, fig. 850. **II** Junior subjective synonym of *Phalangium hellwigii* Panzer, 1794 by Martens (1965: 144).



**Case saxonicum:** Kästner (1928: 6) presented a description of *Nemastoma saxonica*, attributing it to Hnatewytch, with his consent. His figure 25 was prepared by Hnatewytch. The following year, Hnatewytch (1929) published this same species under the name *Nemastoma spinosa*. Roewer (1951: 146) incorrectly treated both names as being two separate species. We here concur with the analyses of Spoek (1963: 17) and Meijer (1973a: 120) that *N. spinosa* is a junior objective synonym of *N. saxonicum*. And the



correct forms of the specific epithets should be *saxonicum* and *spinosum* (as Kratochvíl in 1934 and Roewer in 1951 correctly wrote) and not *saxonica* because both *Nemastoma* and *Mitostoma* are of neuter gender.

#### ***Mitostoma chrysomelas* (Hermann, 1804)**

- *Phalangium chrysomelas* Hermann 1804: 108, pl. 8, figs 3, H, I, K, L (French vernacular “faucheur chrysomèle”).  
*Mitostoma chrysomelas*: Roewer 1951: 142.
- *Nemastoma saxonica* Hnatewytch in Kästner 1928: 1. **II** Junior subjective synonym of *Phalangium chrysomelas* Hermann, 1804 by Meijer (1973a: 125).  
*Mitostoma saxonicum*: Roewer 1951: 146, pl. 8, fig. 71.
- *Nemastoma spinosa* Hnatewytch 1929: 231, fig. 21. **II** Junior **objective** synonym of *Nemastoma saxonicum* Hnatewytch in Kästner, 1928, as noted by Spoek (1963: 17).  
*Nemastoma spinosum*: Kratochvíl 1934: 8, fig. 37.  
*Mitostoma spinosum*: Roewer 1951: 146, pl. 8, fig. 70.



**Case Grassatores:** Giribet et al. (2002) published the infraorder Grassatores, attributing the authorship to Kury. An infraorder is a **higher group** above the family-group. For such a higher-group name the Code provides some rules, for example it must be published, after 1757, it must be written in Latin script, it must be capitalized, it must not be published anonymously, no diacritics. Authorship is not regulated, there is no need for a description, the Principle of Priority does not apply, no typification is needed, homonymy can be disregarded. Those names are largely **based on common acceptance**. So it is convenient to take the name Grassatores from the first published source, Giribet et al. 2002, and attribute it to Kury as given in that original source. In spite of Grassatores having been originally called a nomen nudum by Giribet et al., **the term “nomen nudum” cannot be applied to a higher-group name** because this term is defined as a name that fails to conform to Arts. 12 or 13. These Articles are irrelevant for higher-group names.

#### **Infraorder Grassatores Kury, 2002**

- Grassatores **Kury in Giribet** et al. 2002: 31.



**Case ibericus:** Dresco (1967: 382) investigated a misidentification of *Nemastoma bacilliferum bacilliferum* by (Rambla 1959: 73), stating that it should be referred instead to a new species which he called “*Nemastoma ibericus* Rambla 1965, sp. nov. = *N. bacilliferum bacilliferum* (Rwr 1914), sensu Rambla 1959.” And a few lines below he specifically stated that he asked Rambla to give a name to this species that he recognized as new. Apart from the strange date of 1965 (presumably the year in which he corresponded with Rambla about the situation) and the incorrect inflection of the specific

epithet as *ibericus* instead of the correct *ibericum*, **we concur with his acts**. Only, the date should be 1967, as already noted by Starega (1987: 304). Prieto (2004: 117) mentioned the lack of type specimens, but for a species established before 2000 it is allowed (Code Art. 72.3).

#### ***Nemastomella iberica* (Rambla, 1967)**

- *Nemastoma ibericus* **Rambla in Dresco** 1967: 382.  
*Nemastomella iberica*: Starega 1987: 304.



#### **Taxa attributed to Sørensen in Lendl (1894)**

**Comment.** Adolf Lendl sent material from the Hungarian National Museum to Copenhagen for William Sørensen to study. After five years, Sørensen returned the collection, explaining that he could not finish his planned work due to his different and excessive preoccupations. He had only identified about 3/4 of the specimens. With Sørensen’s consent, Lendl presented the descriptions in his 1894 work. Lendl explicitly said: “the descriptions of the new species are mostly from Sørensen” and attributed the authorship of the correspondent species to him while others he attributed to himself. Our interpretation is that Lendl alone is responsible for some species descriptions and should be credited as such, while others should be credited as “Sørensen in Lendl”.

#### ***Astrobonus helleri* (Ausserer, 1867)**

- *Acantholophus Helleri* Ausserer 1867: 167, pl. 8, fig. 1.  
*Astrobonus Helleri*: Simon 1879b: 169.
- *Astrobonus croaticus* **Sørensen in Lendl** 1894: 21. **II** Junior subjective synonym of *Acantholophus helleri* Ausserer, 1867 by Martens (1978b: 389).

#### ***Carinostoma elegans* (Sørensen, 1894)**

- *Nemastoma elegans* **Sørensen in Lendl** 1894: 29.  
*Carinostoma elegans*: Martens 1978: 137, figs 201–207.

#### ***Egaenus convexus* (C.L. Koch, 1835)**

- *Opilio convexus* C.L. Koch 1835: plate and legend 128.19.
- *Egaenus maximus* **Sørensen in Lendl** 1894: 24. **II** Junior subjective synonym of *Opilio convexus* C.L. Koch, 1835 by Roewer (1911e: 17).

#### ***Lacinius horridus* (Panzer, 1794)**

- *Phalangium horridum* Panzer 1794: XVII, 21.  
*Lacinius horridus*: Thorell 1876a: 464, 480.
- *Acantholophus bellicosus* **Sørensen in Lendl** 1894: 25. **II** Junior subjective synonym of *Opilio hispidus* Herbst, 1798 by Roewer (1912e: 74).

#### ***Odiellus lendlii* (Sørensen, 1894)**

- *Acantholophus Lendlii* **Sørensen in Lendl** 1894: 26.

### ***Paranemastoma sillii sillii* (Herman, 1871)**

- *Nemastoma Sillii* Herman 1871: 28.
- *Nemastoma gigas* Sørensen in Lendl 1894: 29. II Junior subjective synonym of *Nemastoma sillii* Herman, 1871 by Staręga (1978: 205).
- *Nemastoma lineatum* Sørensen in Lendl 1894: 30. II Junior subjective synonym of *Nemastoma sillii* Herman, 1871 by Martens (1978b: 126).



**Case urnigerum:** Jean Frédéric Hermann died untimely at 25 due to a typhus epidemic when he was already finishing his important monograph on wingless insects (Hermann 1804) and his brother-in-law, **Frédéric-Louis Hammer** edited that work and saw it through press. The work was basically ready when Hermann died, so it is easy to ascertain that Hermann is the author of nearly everything. There is but one species which was clearly described in an **addendum of the editor**, clearly marked as such: “Addition de l’Éditeur”.

### ***Mitopus morio* (Fabricius, 1779)**

- *Phalangium Morio* Fabricius 1779: 340.  
*Mitopus morio*: Thorell 1876a: 465, 492.
- *Phalangium urnigerum* Hammer in Hermann 1804: 110, pl. 9, figs 2–3. II Junior subjective synonym of *Phalangium palliatum* Latreille, 1798 by Latreille (1806: 141); junior subjective synonym of *Opilio grossipes* Herbst 1799 by C.L. Koch (1847b: 23); revalidated by Meade (1855: 401); synonymy with *O. grossipes* reaffirmed by Grube (1859: 41); junior subjective synonym of *Phalangium morio* Fabricius, 1779 by Thorell (1876a: 492).



### **Taxa attributed to Sørensen in Henriksen (1932a)**

**Comment.** Henriksen (1932a) edited the written material left by Sørensen by occasion of his death. He provided a detailed introduction explaining where and how “Sørensen has the full credit and responsibility” for the major part the descriptions, while a few clearly marked were his own. There was no case of shared description as in Koch and Kollar. There is a great number of genera and species described in that work which should be credited to Sørensen (not cited exhaustively here). And there is only one case in which Henriksen correctly credited the first editor Carl Johannes With, who had also died.

### ***Asopella xanti* Sørensen, 1932**

- *Asopella xanti* Sørensen in Henriksen 1932a: 258.

### ***Cardwella atar* (Sørensen, 1932)**

- *Wintonia atar* Sørensen in Henriksen 1932a: 218.  
*Cardwella atar*: Roewer 1935a: 63.

### ***Cleombrotus* Sørensen, 1932**

- *Cleombrotus* Sørensen in Henriksen 1932a: 268. 🕸 Type-species by monotypy: *Cleombrotus minutus* Sørensen, 1932.

### ***Cleombrotus minutus* Sørensen, 1932**

- *Cleombrotus minutus* Sørensen in Henriksen 1932a: 268.

### ***Epedanus praedo* Sørensen, 1932**

- *Epedanus praedo* Sørensen in Henriksen 1932a: 254.

### ***Gnomulus thorelli* (Sørensen, 1932)**

- *Pelitus thorelli* Sørensen in Henriksen 1932a: 213.  
*Gnomulus thorelli*: Martens & Schwendinger 1998: 527.

### ***Metassamia variata* Sørensen, 1932**

- *Metassamia variata* Sørensen in Henriksen 1932a: 217.

### ***Meterginus simonis* (With, 1932)**

- *Rhaucus* (*Erginus*) *simonis* With in Henriksen 1932a: 350.

### ***Obidosus* Roewer, 1931**

- *Obidosus* Roewer 1931e: 154. 🕸 Type-species by monotypy: *Obidosus amplichelis* Roewer, 1931. II Junior subjective synonym of *Protimesius* Roewer, 1913 by Pinto-da-Rocha (1997a: 275); synonymy disclaimed by Villarreal et al. (2019: 978).
- *Ideostygnus* Sørensen in Henriksen 1932a: 288. 🕸 Type-species by monotypy: *Ideostygnus laevis* Sørensen, 1932. II Junior subjective synonym of *Pickeliana* Mello-Leitão, 1932 by Mello-Leitão (1935e: 112); junior subjective synonym of *Protimesius* Roewer, 1913 by Pinto-da-Rocha (1997a: 275); junior subjective synonym of *Obidosus* Roewer, 1931 by Villarreal et al. (2019: 978).

### ***Philacarus* Sørensen, 1932**

- *Philacarus* Sørensen in Henriksen 1932a: 266. 🕸 Type-species by monotypy: *Philacarus samoides* Sørensen, 1932.

### ***Philacarus samoides* Sørensen, 1932**

- *Philacarus samoides* Sørensen in Henriksen 1932a: 267.

### ***Seuthes simonis* (Sørensen, 1932)**

- *Paraselenca simonis* Sørensen in Henriksen 1932a: 219.  
*Seuthes simonis*: Roewer 1935: 111.

### ***Spinopilar anomalis* (Sørensen, 1932)**

- *Olynthus anomalis* Sørensen in Henriksen 1932a: 251.  
*Olynthus anomalus*: Mello-Leitão 1935e: 90.  
*Spinopilar anomalis*: Kury 2003a: 204.

### ***Tibangara* Mello-Leitão, 1940**

- *Tachus* Sørensen in Henriksen 1932a: 277. 🕸 Type-species by monotypy: *Tachus keyserlingii* Sørensen, 1932. II Junior homonym of *Tachus* Jurine, 1807 (Hymenoptera); homonymy first noted by Strand (1942).
- *Tibangara* Mello-Leitão 1940c: 100. 🕸 Type-species by original designation: *Tibangara nephelina* Mello-Leitão, 1940.



### *Tibangara keyserlingii* (Sørensen, 1932)

- *Tachus keyserlingii* Sørensen in Henriksen 1932a: 278.  
*Tachusina keyserlingii*: Strand 1942: 400.  
*Tibangara keyserlingii*: Kury 2014: 11, 24.

**Case *Cheiromachus*:** as in Koch & Berendt (1854). Georg Berendt started this project, and later called upon Carl Ludwig Koch for help with the second volume. Koch abandoned the project in 1849 and Berendt died in January 1850, so that Anton Menge undertook the task of editing the manuscript and seeing the book through press to publication. Some taxa are described, in footnotes, and explicitly signed by Menge, who is thus the author of those taxa.

### † *Cheiromachus* Menge, 1854

- *Cheiromachus* Menge in Koch & Berendt 1854: 102. 🐛 Type-species by monotypy: *Cheiromachus coriaceus* Menge, 1854.

### † *Cheiromachus coriaceus* Menge, 1854

- *Cheiromachus coriaceus* Menge in Koch & Berendt 1854: 102.

## 1.4.2.2. Subdivided responsibilities for descriptions: wasted chivalry

Whenever someone wishes to acknowledge use of notes/discoveries from prior researchers, it is common to make them authors or co-authors of those taxa. This practice is widespread in harvestmen and so far nobody questioned it. An early example is CL Koch, who attributed many of the species described in his series

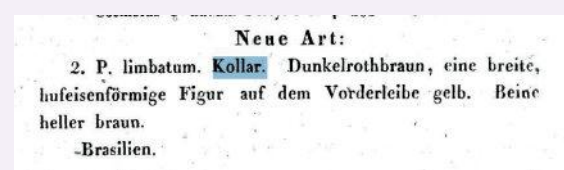
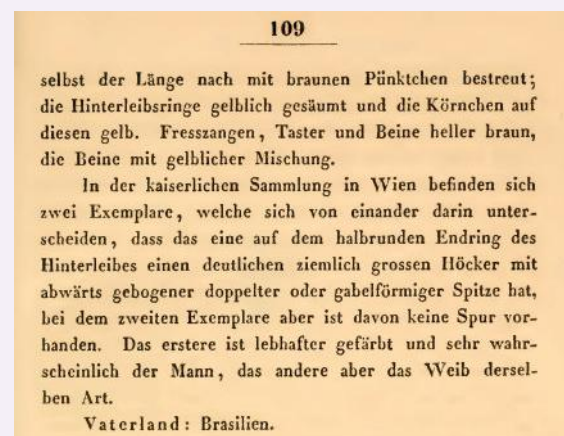
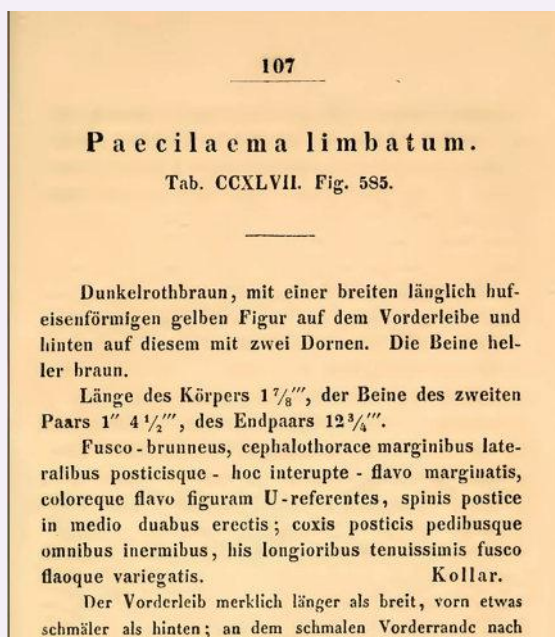
“Die Arachniden” to Vincenz Kollar, upon whose notes he partly based the descriptions (Fig. 3). Koch, courteously noted that Kollar was to be the author. And subsequent authors corroborated this idea.

Yet, under the cold scrutiny of modern nomenclature practice, this concept, even prevalent in Opilionology, does not hold.

We transcribe here three illuminating analyses by colleagues, who essentially arrived at the same conclusions:

(a) “It seems fairly straightforward that Koch is the author and not Kollar if these names had not been published previously. Simply placing an outside author’s name behind a novel nomen does not automatically make that person the author. There has to be evidence that that person provided more than just the name. In works where such a placement is made, either the descriptive characters are placed in quotes or mention is made in the work that that person provided descriptive characters. Looking at your attachment with Kollar’s name right after the description seems to imply that Kollar is the author of those characters, but as there is no evidence to prove it and there are no quotes, Koch is the author.” [Neal Evenhuis, pers. comm.].

(b) “What Koch considered regarding the authorship of these nomina, this is irrelevant. Authorship is determined not by what the authors of the paper think, but by the Rules of the Code. I see no evidence that the diagnoses of the new taxa, either in German or in Latin, were



**Figure 4.** Misguided common sense indications that Kollar is the author of some species: (a). Snapshot of Koch (1839b), showing Kollar’s name at the end of a Latin diagnosis; (b). Same, showing final part of German text of description, without any mention to Kollar; (c). Snapshot of subsequent work of Koch (1839c), showing the name of species with authorship clearly attributed to Kollar.

written by Kollar and copied by Koch. Such an evidence could be for example if these diagnoses were between quotation marks. Kollar might just have written labels in collections and not even written one word about these animals. Or he might have written complete descriptions or diagnoses that were copied by Koch. But for this latter interpretation, there should exist some evidence somewhere that this was the case.” [Alain Dubois, pers. comm.].

(c) The name as such was not explicitly attributed to Kollar in the original description, so we do not need to look closer on the responsibility for the description. If we look closer, then we see that only the Latin diagnosis was attributed to Kollar, but a German text containing additional components of the description was not, and we should assume that these were Koch’s own additions. In any case it is not “clear from the contents” that the German parts of the descriptions were authored by Kollar. **The criteria of Art. 50.1.1 for a “foreign” authorship are doubly not met**, because the name was not attributed to Kollar, and because Kollar was not “alone” responsible for the description. In such a case **the authorship must be attributed to Koch**, the author of the work. The observation that everybody in arachnology attributed the authorship to Kollar, has no influence on the correct application of the Code. [Francisco Welter-Schultes, pers. comm.]

This cold view by ICZN, which we have to follow here, **spoils the chivalry** of ladies and gentlemen who intended to pay homage to colleagues who treated the taxon before and which they feel should be somehow acknowledged. We consider it to be a **betrayal to the spirit** of the tributes paid by thoughtful authors.

**Opilio-example:** Regina Stefanini-Jim recognized a new species of *Goniosoma* in her unpublished thesis (1985). DaSilva & Gnaspini, in a review of the Goniosomatinae (2010) graciously acknowledged this by naming this species *Pyatan insperatum* DaSilva, Stefanini-Jim & Gnaspini, 2010. The authorship, however, is of DaSilva & Gnaspini.


**Welter-Schultes more or less parallel examples:** (1) Rossmässler 1835 established a new gastropod name *Clausilia cattaroensis* and attributed the name to Ziegler. The description was partly by Rossmässler and partly by Ziegler who was cited in quotation marks. The name must be attributed to Rossmässler because Ziegler was not alone responsible for the description. (2) Forster 1840 established a new gastropod name *Pupa sterrii* and attributed the name to Voith. The description was partly by Voith (the main description) and partly by Forster (who only gave a brief summary of Voith’s description in his own words). The name must be attributed to Forster because Forster’s text alone would equally have made the name available.

Similar cases in Opiliones:




**Case Kollar in Koch:** As abundantly explained above, all names of species which appeared in Koch (1839b) and were there attributed to Kollar must be attributed to Koch alone:


#### ***Pungoica decorata* (C.L. Koch, 1839)**

- *Ostracidium decoratum* C.L. Koch 1839b: 14, pl. 219, fig. 546.   
Originally attributed to Kollar.  
*Pungoica decorata*: Kury 2020c: xxx.


#### ***Stygnoplus triacanthus* (C.L. Koch, 1839)**

- *Stygnus triacanthus* C.L. Koch 1839b: 23, pl. 221, fig. 549.   
Originally attributed to Kollar.  
*Stygnoplus triacanthus*: Simon 1879a: 222.


#### ***Paecilaema u-flavum* (Perty, 1833)**

- *Cosmetus U-flavum* Perty 1833: 203.
- *Paecilaema limbatum* C.L. Koch 1839c: 21.  Originally attributed to Kollar. **II** Junior subjective synonym of *Cosmetus u-flavum* Perty, 1833 by Kury & Medrano (2018: 205).


#### ***Asarcus longipes* C.L. Koch, 1839**

- *Asarcus longipes* C.L. Koch 1839b: 68, pl. 234, fig. 569.   
Originally attributed to Kollar.


#### ***Mitogoniella badia* (C.L. Koch, 1839)**

- *Goniosoma badium* C.L. Koch 1839b: 65, pl. 233, fig. 568.   
Originally attributed to Kollar.  
*Mitogoniella badia*: DaSilva & Gnaspini 2010: 592.


#### ***Neoancistrotus bifurcatus* (C.L. Koch, 1839)**

- *Ancistrotus bifurcatus* C.L. Koch 1839b: 45, pl. 226, fig. 559.   
Originally attributed to Kollar.  
*Neoancistrotus bifurcatus*: Kury 1992a: 159.

#### ***Promitobates hexacanthus* (C.L. Koch, 1839)**

- *Ancistrotus hexacanthus* C.L. Koch 1839b: 48, pl. 226, fig. 560.   
Originally attributed to Kollar.  
*Promitobates hexacanthus*: Roewer 1913d: 286, fig. 114.

#### ***Discocyrtus flavigranulatus* B. Soares, 1944**

- *Gonyleptes curvipes* C.L. Koch 1839b: 36, pl. 224, fig. 555. **II** Junior primary homonym of *Gonyleptes curvipes* Guérin-Méneville, 1837, first detected by Kury et al. (2018: 358).   
Originally attributed to Kollar.
- *Discocyrtus flavigranulatus* B. Soares 1944b: 165, fig. 11. **II** Junior subjective synonym of *Gonyleptes curvipes* C.L. Koch, 1839 by Kury et al. (2018: 358). In virtue of the homonymy of *Gonyleptes curvipes* C.L. Koch, 1839, *Discocyrtus flavigranulatus* B. Soares, 1944 is the next oldest nomen from amongst the available synonyms.



### ***Eusarcus oxyacanthus* C.L. Koch, 1839**

- *Eusarcus oxyacanthus* C.L. Koch 1839b: 7, pl. 218, figs 543–544. Originally attributed to Kollar.

### ***Graphinotus ornatus* C.L. Koch, 1839**

- *Graphinotus ornatus* C.L. Koch 1839b: 10, pl. 219, fig. 545. Originally attributed to Kollar.

### ***Pachylus chilensis* (Gray, 1833)**

- *Gonoleptes chilensis* Gray 1833: 539, pl. 20, fig. 2. Subsequent incorrect spelling of generic nomen.
- *Pachylus granulatus* C.L. Koch 1839b: 20, pl. 221, fig. 548. Junior subjective synonym of *Gonyleptes chilensis* Gray 1833 by Sørensen (1895: 2). Originally attributed to Kollar.

### ***Pyatan insperatum* DaSilva & Gnaspini, 2010**

- *Pyatan insperatum* DaSilva & Gnaspini 2010: 560, figs 46, 47, 141–144, 201, 202. Originally attributed to DaSilva, Stefanini-Jim & Gnaspini.



**Case Guérin-Méneville in Gervais:** Gervais (1842) presented two new species attributed to Guérin-Méneville, and indicated in an explanatory note that Guérin-Méneville provided the information. Gervais also gave a reference to the plate explanation of the unpublished source which was published later in 1844 by Guérin-Méneville. Consulting the external 1844 source reveals that Guérin-Méneville had included a detailed species description on the plate caption, and that Gervais cited it almost verbatim. However Gervais did not indicate this unambiguously in 1842, and even experienced readers would not expect a detailed species description to have been cited verbatim from a figure caption. Explanations in figure captions usually referred to what was shown on the plate, while diagnoses and detailed species descriptions appeared separately in textual parts of works. The fact that an external source needs to be consulted is decisive for the authorship attribution. Because the external authorship attribution for the description was not unambiguously clear in the original source, the authorship for *Gonyleptes planiceps* and *Stygnus inflatus* must be attributed to Gervais alone. See Chapter 12 for details about *Eubalta planiceps*. See also section “Unintentionally stolen names” in Welter-Shultes (2012: 97).

### ***Eubalta planiceps* (Gervais, 1842)**

- *Gonyleptes planiceps* Gervais 1842: 2, pl. 2. Originally attributed to Guérin-Méneville.

### ***Stygnidius inflatus* (Gervais, 1842)**

- *Stygnus inflatus* Gervais 1842: 3, pl. 3. Originally attributed to Guérin-Méneville.  
*Stygnidius inflatus*: Simon 1879a: 223.



**Case Kollar in Doleschall:** Doleschall (1852) simply wanted to homage Kollar, who was then the director of the Natural History Museum in Vienna, and had collected the type specimen. There is not the slightest evidence that Kollar had anything to do with the description.

### ***Egaenus convexus* (C.L. Koch, 1835)**

- *Opilio convexus* C.L. Koch 1835: plate and legend 128.19.  
*Egaenus convexus*: C.L. Koch 1839c: 25.
- *Egaenus atomarius* Doleschall 1852: 650. Originally attributed to Kollar. **II** Junior subjective synonym of *Opilio convexus* C.L. Koch, 1835 by Roewer (1923: 813).



**Case Corti in Jaquet:** this case departs widely from the other cases here because it is simply a case of incorrect attribution of authorship of the work, nothing to do with attribution of a taxon nomen to an external author. Several works have been miscredited in this way, of which we give a few examples: (1) “Koch in Hahn & Koch” (1839), when it was a volume authored by Koch in a series that had started with Hahn. (2) “Sørensen in Koch” (1886), when it was simply a book chapter authored by Sørensen in series edited by Koch & Keyserling. This is exactly the same case: a work authored by Corti in a series authored by Jaquet. It is clearly abundant that Corti is the author not only from the subtitle, but also from phrases such as “I dedicate this species to Maurice Jaquet, who collected the specimens”. Therefore, the formula “Corti in Jaquet” is incorrect, and it should be cited as Corti [alone].

### ***Holoscotolemon jaqueti* (Corti, 1905)**

- *Phalangodes Jaqueti* Corti 1905: 224. Widely cited as “Corti in Jaquet”.  
*Holoscotolemon jaqueti*: Martens 1978b: 80, figs 103–106.



**Case C. Koch in Roewer:** Roewer (1911e: 50) described *Paropilio leiobuniformis*, but attributed authorship to “C.L. Koch, 1872” (which is clearly a lapse for C. Koch; see above for the three quasi-homonym Kochs). In the short paper by C. Koch (1872) there is only the description of *Opilio canariensis* (which should not be confused with *canariana*). Roewer also mentioned he examined Koch’s material, labeled as “*Opilio leiobuniformis* C. Koch. Typen”, which was then undescribed. It is clear that Roewer meant to acknowledge Koch’s prior identification of the species. However, the authorship should be of Roewer alone.

### ***Bunochelis canariana* (Strand, 1911)**

- *Phalangium canarianum* Strand 1911: 200.  
*Bunochelis canariana*: Staręga 1972b: 467, figs 6–10.
- *Paropilio leiobuniformis* Roewer 1911e: 50. Originally attributed to C. Koch. **II** Junior subjective synonym of

*Phalangium canarianum* Strand, 1911 by Starega (1972b: 467).



**Case Kollar in Roewer:** Roewer (1911e: 13) described *Phalangium pareissii*, but attributed the authorship to Kollar. He explained that Kollar had already discovered this species as new and even labeled it as *P. pareissii*. In spite of Roewer's ethics, authorship is Roewer's.

#### ***Phalangium punctipes* (L.Koch, 1879)**

- *Opilio punctipes* L.Koch 1879a: 63.  
*Phalangium punctipes*: Simon 1879b: 208.
- *Phalangium pareissii* Roewer 1911e: 13. Originally attributed to Kollar. **II** Junior subjective synonym of *Opilio punctipes* L.Koch, 1879a by Starega (1973a: 134).



**Case Hadži in Kratochvíl:** Kratochvíl (1934: 19) described the new species *Lacinius regis-alexandri* and attributed it to Hadži without any explanation whatsoever. We understand it here as a misguided homage, and the authorship is of Kratochvíl alone.

#### ***Lacinius horridus* (Panzer, 1794)**

- *Phalangium horridum* Panzer 1794: XVII, 21.  
*Lacinius horridus*: Thorell 1876a: 464, 480.
- *Lacinius regis-alexandri* Kratochvíl 1934: 19. Originally attributed to Hadži. **II** Junior subjective synonym of *Phalangium horridum* Panzer, 1794 by Šilhavý (1956a).



**Case Martens in Benavides et al.:** Martens (1969) has originally described a species of uncertain genus by using "Gen.?" to denote this. This nomen was unavailable for half a century, without being made available unintentionally before 2000 by any subsequent author, until Benavides et al. (2019) in a revision, decided to describe this species as new in the genus *Neogovea*. However, as Martens did not participate in anything, even the type material being freshly collected, authorship should be of Benavides et al.

#### ***Neogovea enigmatica* Benavides, Hormiga & Giribet, 2019**

- *Neogovea enigmatica* Benavides, Hormiga & Giribet 2019: 153, figs 42–44. Originally attributed to Martens.



# Chapter 2. Mandatory corrections proposed here

## 2.1. Grammatical adjective inflexions corrected herein (Code Art. 31.2)

**Comment.** These are mandatory changes referring to gender and inflection of adjectives. See also [Chapter 9](#) on *Metaphalangium/Paropilio*. The commission which is redacting the 5th Edition of the Code is considering to **relax the rules of gender agreement in cases on prevailing usage**. Generally, for most nomina in Opiliones (including all the cases listed here), their citations in the literature are so few that none deserves this status, therefore changing back to proper grammar is important to keep a semblance of Latin language in the scientific names.

### Assamiidae Sørensen, 1884

#### *Neobaeorix* Lawrence, 1962

- *Neobaeorix* Lawrence 1962: 71. 🐞 Type-species by original designation: *Neobaeorix cornuta* Lawrence, 1962.

🌍 East Tropical Africa.

**Etymology.** From Greek νέος (new) + pre-existing genus *Boeorix* misspelled as *Baeorix*. **Gender masculine.**

#### *Neobaeorix cornutus* Lawrence, 1962

- *Neobaeorix cornuta* Lawrence 1962: 72. 📖 Incorrect originally applied gender declination.

#### *Triaenopodium* Roewer, 1915

- *Triaenopodium* Roewer 1915c: 70. 🐞 Type-species by monotypy: *Triaenopodium tarsale* Roewer, 1915.

🌍 Malay Peninsula.

**Etymology.** From Greek τρίαινα (trident) + Latin *podium* = “foot”. **Gender neuter.**

#### *Triaenopodium pyriforme* Suzuki, 1976

- *Triaenopodium pyriformis* Suzuki 1976c: 33, figs 3H; 12A-I. 📖 Incorrect originally applied gender declination.

#### *Triaenopodium tarsale* Roewer, 1915

- *Triaenopodium tarsalis* Roewer 1915: 71, fig. 36. 📖 Incorrect originally applied gender declination.

### Cranidae Roewer, 1913

#### *Phareicranus* Roewer, 1913

- *Phareicranus* Roewer 1913d: 401. 🐞 Type-species by original designation: *Goniosoma calcariferum* Simon, 1879.

🌍 N South America.

**Etymology.** From the pre-existing genera *Phareus* and *Cranus*. Gender masculine (irrelevant in this case).

#### *Phareicranus x-albus* Roewer, 1932

- *Phareicranus x-albus* Roewer 1932: 301, fig. 17. 📖 Incorrect originally applied gender declination. Latin alphabet letters are neuter, and the adjective must refer to the letter, not to the genus ([Art. 32.5.2.4.3](#)).

### Globipedidae Kury & Cokendolpher, 2020

#### *Metopilio* Roewer, 1911

- *Metopilio* Roewer 1911e: 104. 🐞 Type-species by original designation: *Phalangium armigerum* Pickard-Cambridge, 1905.

**Etymology.** From Greek μετά (after) + pre-existing genus *Opilio*. **Gender masculine.**

#### *Metopilio armiger* (Pickard-Cambridge, 1905)

- *Phalangium armigerum* Pickard-Cambridge 1905: 579.  
*Metopilio armigerum*: Roewer 1911e: 105, pl. 1, figs 1–2. 📖 Incorrect subsequently applied gender declination.  
*Metopilio armigerus*: Roewer 1912: 265, pl. 1, fig. 16, pl. 4, fig. 18. 📖 Incorrect subsequently applied gender declination.

### Gonyleptidae Sundevall, 1833

#### *Triaenomerus* Roewer, 1913

- *Triaenomerus* Roewer 1913c: 223. Type-species by monotypy: *Triaenomerus olivaceus* Roewer, 1913.

**Etymology.** *Triaenomerus* from Greek τρίαινα (trident, three-pronged fish-spear) + μέρος (share, portion). **Gender neuter.**

#### *Triaenomerus olivaceus* Roewer, 1913

- *Triaenomerus olivaceus* Roewer 1913c: 224, fig. 93. 📖 Incorrect originally applied gender declination.

### Nemastomatidae Simon, 1872

#### *Mediostoma* Kratochvíl, 1958

- *Mediostoma* [subgenus of *Histicostoma*] Kratochvíl 1958b: 569. 🐞 Type-species by original designation: *Nemastoma graecum* Roewer, 1919.




*Mediostoma*: Gruber 1976: 799.


 Eurasia.

**Etymology.** *Mediostoma* from Latin *medio* (to divide in the middle) + truncation of pre-existing genus *Nemastoma*. **Gender neuter.** “The name probably refers to the position of pseudoarticulations in the middle of the leg femora. In the original composition of subgenera in *Histicostoma* (Kratohvil 1958) it was compared to the basal-positioned pseudoarticulations in *Basostoma*.” [Schönhofer 2013: 30].


### ***Mediostoma izmiricum* Snegovaya, Kurt & Yağmur, 2016**

- *Mediostoma izmirica* Snegovaya, Kurt & Yağmur 2016: 490, figs 2–11, 14, 17, 20, 23, 26–28, 31, 32, 37.  Incorrect originally applied gender declination.

### ***Mediostoma talischense* (Morin, 1937), nomen dubium**

- *Acropsopilio talischensis* Morin 1937: 210, figs 2A–B.  
*Mediostoma talischensis*: Kury & Snegovaya 2019: 14.  Incorrect subsequently applied gender declination.


### ***Paranemastoma* Redikortsev, 1936**

- *Paranemastoma* Redikortsev 1936: 40.  Type-species by original designation: *Paranemastoma superbum* Redikortsev, 1936.

 Eurasia.


**Etymology.** From Greek παρά (beside) + pre-existing genus *Nemastoma*. **Gender neuter.**

### ***Paranemastoma karolianum* Çorak, Bayram, Yiğit & Sancak, 2017**

- *Paranemastoma karolianus* Çorak, Bayram, Yiğit & Sancak 2017: 242, figs 1–4.  Incorrect originally applied gender declination. The specific epithet is an adjective honoring a person called Sevinç Karol.

### **Phalangiidae Latreille, 1802**


#### ***Graecophalangium* Roewer, 1923**

- *Graecophalangium* Roewer 1923: 755.  Type-species by subsequent designation of Crawford (1992: 22): *Graecophalangium atticum* Roewer, 1923.


 Eurasia.

**Etymology.** From Latin *graecus* (Greek) + pre-existing genus *Phalangium*. **Gender neuter.**

### ***Graecophalangium karakalense* Chemeris & Snegovaya, 2010**

- *Graecophalangium karakalensis* Chemeris & Snegovaya 2010: 69, figs 10–26, 35.  Incorrect originally applied gender declination.


### ***Phalangium* Linnaeus, 1758**

- *Phalangium* Linnaeus 1758: 618.  Type-species: *Phalangium opilio* Linnaeus 1758, by subsequent designation of Latreille (1810: 425, table), see ICZN (1910 and 1939). The type designation of *Phalangium parietinum* De Geer, 1778 by Thorell (1876a) is invalid.


 Eurasia.

**Etymology.** *Phalangium* from Greek φάλαγγξ, φάλαγγος (a clustered mass of infantry, presumably referring to venomous spiders which attack reapers in the fields, and which were in the early 1800s mixed-up for harvestmen). **Gender neuter.**

### ***Phalangium kitabense* Chemeris, 2012**


- *Phalangium kitabensis* Chemeris 2012: 20, figs 3–4, 8, 13–15, 20, 26–27, 30; map 2.  Incorrect originally applied gender declination.


### ***Phalangium kopetdaghense* Chemeris & Snegovaya, 2010**

- *Phalangium kopetdaghensis* Chemeris & Snegovaya 2010: 70, figs 27–35.  Incorrect originally applied gender declination.

### **Phalangodidae Simon, 1879**



#### ***Sitalcina* Banks, 1911**

- *Sitalcina* Banks 1911: 415.  Type-species by monotypy: *Sitalces californicus* Banks, 1893.


 W USA.

**Etymology.** From pre-existing genus *Sitalces* + feminine form of Latin suffix *-inus*. **Gender feminine.**

#### ***Sitalcina californica* (Banks, 1893)**


- *Sitalces californica* Banks 1893d: 151.  
*Sitalcina californica*: Banks 1911: 415, fig. 148B.  
*Sitalcina californicus*: Goodnight & Goodnight 1942f: 8, figs 28–30.  Incorrect subsequently applied gender declination.  
*Paramitraceras californicus*: Roewer 1912c: 155.  Incorrect subsequently applied gender declination.

#### ***Sitalcina lobata* Goodnight & Goodnight, 1942**

- *Sitalcina lobatus* Goodnight & Goodnight 1942f: 9, figs 37–39.  Incorrect originally applied gender declination.

### **Podoctidae Roewer, 1912**

#### ***Paramesoceras* Roewer, 1915**

- *Paramesoceras* Roewer 1915d: 219.  Type-species by monotypy: *Paramesoceras novoguineensis* Roewer, 1915.

 Papua.



**Etymology.** *Paramesoceras* from Greek παρά (beside) + pre-existing genus *Mesoceras*. **Gender neuter.**

***Paramesoceras novoguineense* Roewer, 1915**

- *Paramesoceras novoguineensis* Roewer 1915d: 220, pl. 4, fig. 4. Incorrect originally applied gender declination.

***Podoctops* Roewer, 1949**

- *Podoctops* Roewer 1949c: 277. Type-species by original designation: *Podoctops multimaculatum* Roewer, 1949.

Sunda.

**Etymology.** From pre-existing genus *Podoctis* + Greek ὤψ (eye; face; countenance). **Gender masculine (Code Art. 30.1.4.3).**

***Podoctops multimaculatus* Roewer, 1949**

- *Podoctops multimaculatum* Roewer 1949c: 277, figs 74a–b. Incorrect originally applied gender declination.

**Pyramidopidae Sharma, Prieto & Giribet, 2011**

***Conomma* Loman, 1902**

- *Conomma* Loman 1902: 186. Type-species by subsequent designation of Roewer (1949a: 34): *Conomma fortis* Loman, 1902.

Sub-Saharan Africa.

**Etymology.** From Greek κώνος (cone) + ὄμμα- (of the eye). **Gender neuter.**

***Conomma asperum* Lawrence, 1949**

- *Conomma asper* Lawrence 1949: 16. Incorrect originally applied gender declination.
- *Conomma aspera* Starega 1992: 289. Unjustified emendation.

***Conomma forte* Loman, 1902**

- *Conomma fortis* Loman 1902: 190, fig. 16. Incorrect originally applied gender declination.

***Conomma minimum* Roewer, 1912**

- *Conomma minima* Roewer 1912c: 125. Incorrect originally applied gender declination.

***Conomma orientale* Roewer, 1949**

- *Conomma orientale* Roewer 1949a: 38, figs 62a–d.
- *Conomma orientalis* Roewer 1961a: 38. Incorrect subsequently applied gender declination.

**Samoidae Sørensen, 1886**

***Mitraceras* Loman, 1902**

- *Mitraceras* Loman 1902: 186. Type-species by monotypy: *Mitraceras crassipalpus* Loman, 1902.

Seychelles.

**Etymology.** From Greek μίτρα (headband) + κέρασ (horn). **Gender neuter.**

***Mitraceras pulchrum* Rambla, 1984**

- *Mitraceras pulchra* Rambla 1984: 53, figs 18–19. Incorrect originally applied gender declination.

**Stygnopsidae Sørensen, 1932**

***Karos Goodnight & Goodnight, 1944***

- *Karos* Goodnight & Goodnight 1944a: 3. Type-species by original designation: *Karos barbarikos* Goodnight & Goodnight, 1944.

Mexico and Central America.

**Etymology.** From Greek κάρος (heavy sleep, torpor). **Gender masculine (Code Art. 30.1.2).**

***Karos tersus* Cruz-López & Francke, 2015**

- *Karos tersum* Cruz-López & Francke 2015: 852, figs 21–23, 59, 61C, 62A, 63G. Incorrect originally applied gender declination.

**Trianeaonychidae Sørensen, 1886**

***Parattahia* Roewer, 1915**

- *Parattahia* Roewer 1915b: 89. Type-species by monotypy: *Parattahia u-signatum* Roewer, 1915.

Tasmania.

**Etymology.** From toponym Parattah. Gender feminine (irrelevant in this case).

***Parattahia u-signatum* Roewer, 1915**

- *Parattahia U-signata* Roewer 1915b: 89. Incorrect originally applied gender declination. Latin alphabet letters are neuter, and the adjective must refer to the letter, not to the genus (Art. 32.5.2.4.3).

**Zalmoxidae Sørensen, 1886**


***Acanthominua* Sørensen, 1932**

- *Acanthominua* Sørensen in Henriksen 1932a: 248. Type-species by monotypy: *Acanthominua tricarinata* Sørensen, 1932.
- *Phalangodinella* Caporiacco 1951b: 5. Type-species by monotypy: *Phalangodinella roeweri* Caporiacco, 1951. Junior subjective synonym of *Acanthominua* Sørensen, 1932 by Pérez-González et. al. (2017: 15)


N South America.

**Etymology.** *Acanthominua* from Greek ἄκανθα (thorn) + pre-existing genus *Minua*. **Gender masculine.**

### ***Acanthominua aridus* (González-Sponga, 1987)**

- *Phalangodinella arida* González-Sponga 1987: 237, figs 277–282.  
*Acanthominua arida*: Pérez-González et. al. 2017: 15.  Incorrect subsequently applied gender declination.


### ***Acanthominua pilosus* (González-Sponga, 1987)**

- *Phalangodinella pilosa* González-Sponga 1987: 263, figs 321–326.  
*Acanthominua pilosa*: Pérez-González et. al. 2017: 15.  Incorrect subsequently applied gender declination.

### ***Acanthominua tricarinatus* Sørensen, 1932**

- *Acanthominua tricarinata* Sørensen in Henriksen 1932a: 249.  Incorrect originally applied gender declination.


### ***Acanthominua tropophylus* (González-Sponga, 1987)**

- *Phalangodinella tropophyla* González-Sponga 1987: 274, figs 339–344.  
*Acanthominua tropophyla*: Pérez-González et. al. 2017: 27.  Incorrect subsequently applied gender declination.


## 2.2. List of ill-formed nomina based on mutilated adjectives or substantives which are herein corrected

**Comment.** Following the reasoning put forth in Section (1.3), some articles of ICZN are in conflict. We then find justification to correct the following names, invoking Arts 26 and 34.2 and ignoring the myopic Art. 32.5.1 (the “non-conformist” view of page 7). Nevertheless, by strictly observing Art. 32.5.1 it would also be possible to regard those as arbitrary combinations of letters, not to be corrected.



### ***Gonyleptes ater* Mello-Leitão, 1923**

- *Gonyleptes atrus* Mello-Leitão 1923c: 140, fig. 16.  Incorrect originally applied gender declination. Species epithet is a Latin adjective of the 1<sup>st</sup> and 2<sup>nd</sup> declensions (āter, ātra, ātrum). Therefore, the nonexistent mock-masculine form “atrus” should be corrected to *ater*.


### ***Calicina brevis* (Briggs 1968)**

- *Sitalcina sierra breva* Briggs 1968: 21, figs 17, 47, 77.  Incorrect originally applied gender declination. Species epithet is a Latin adjective of the 3<sup>rd</sup> declension (brevis, brevis, breve). There is no \*brevus, \*breva, \*brevum in Latin. Therefore, the nonexistent mock-feminine form “breva” should be corrected to *brevis*.  
*Calicina breva*: Ubick & Briggs 1989: 119, figs 14a–c.



### ***Paraconomma ovale* Goodnight & Goodnight, 1942**

- *Paraconomma ovale* Goodnight & Goodnight 1942e: 5, figs 1–2.  Incorrect originally applied gender declination. Species epithet is a Latin adjective of the 3<sup>rd</sup> declension (ovālis, ovālis, ovāle). *Paraconomma*, in spite of ending in -a, is not a feminine name as mistakenly thought by the authors. There is no \*ovalus, \*ovala, \*ovalum in Latin. Therefore, the nonexistent mock-feminine form “ovala” should be corrected to *ovale*.  
*Paraconomma ovale*: Kury 2003a: 27.  Subsequent mandatory inflection of adjective to agree in gender with the neuter generic name (Code Art. 34.2).


### ***Gagrella pauper* With, 1905**

- *Gagrella paupera* With 1905: 7.  Incorrect originally applied gender declination. Species epithet is an invariable Latin adjective of the 3<sup>rd</sup> declension (pauper, pauper, pauper). Therefore, the nonexistent mock-feminine inflection “paupera” should be corrected to *pauper*.

### ***Cosmetus pulcher* Goodnight & Goodnight, 1942**

- *Cosmetus pulchrus* Goodnight & Goodnight 1942g: 4, fig. 18.  Incorrect originally applied gender declination. Species epithet is a Latin adjective of the 1<sup>st</sup> and 2<sup>nd</sup> declensions (pulcher, pulchra, pulchrum). Therefore, the nonexistent mock-masculine inflection “pulchrus” should be corrected to *pulcher*.  
*Cosmetus pulcher*: Kury 2003a: 40.  Subsequent mandatory inflection of adjective to agree in gender with the masculine generic name (Code Art. 34.2).

### ***Erginulus pulcher* Goodnight & Goodnight, 1942**

- *Erginulus pulchrus* Goodnight & Goodnight 1942b: 6, figs 12–13.  Incorrect originally applied gender declination. Species epithet is a Latin adjective of the 1<sup>st</sup> and 2<sup>nd</sup> declensions (pulcher, pulchra, pulchrum). Therefore, the nonexistent mock-masculine inflection “pulchrus” should be corrected to *pulcher*.


## 2.3. Formation of nomina

### **Phalangodidae Simon, 1879**

#### ***Ptychosoma* Sørensen, 1873**

 Mediterranean.

#### ***Ptychosoma espanoli* (Rambla, 1977)**

- *Scotolemon españolii* Rambla 1977a: 272, figs 2a–d.  
*Scotolemon espanoli*: Rambla & Juberthie 1994: 219.
- *Ptychosoma spagnoli*: Prieto 2008: 51.  **Unjustified emendation.**



**Comment.** For already established specific epithets, the “ñ” should not be transcribed into “gn”, but rather the tilde should be dropped (Code Art. 32.5.2.1).

## 2.4. Replacement name for a secondary homonym

### Epedanidae Sørensen, 1886

#### *Dibunus* Loman, 1906

SE Asia.

#### *Dibunus lagunae* Kury nomen novum

- *Triacudorsulum bakeri* Roewer 1926a: 555, pl. 2, fig. 2. **II** Junior secondary homonym of *Anacudorsum bakeri* Roewer, 1926; **first noted here.**

*Dibunus bakeri*: Goodnight & Goodnight 1957: 73.  
urn:lsid:zoobank.org:act:FB335E4B-94A1-464A-9937-523D-150013DC

**Comment.** In virtue of the generic synonymies of *Anacudorsum* Roewer, 1926 and *Triacudorsulum* Roewer, 1926 with *Dibunus* Loman, 1906 (both proposed by Goodnight & Goodnight 1957), *Triacudorsulum bakeri* Roewer, 1926 became a junior secondary homonym of *Anacudorsum bakeri* Roewer, 1926, but this homonymy has not been hitherto detected. Therefore, we here propose the new replacement name *Dibunus lagunae* for *Triacudorsulum bakeri* Roewer, 1926. The new replacement name derives from the province of Laguna, where the type locality of this species is situated. Because both names were published simultaneously in Roewer’s 1926 papers, we here act as the first reviser in determining which name has precedence over the other.

## 2.5. Synonymies originally proposed with inverted precedence and corrected here (Code Art. 23)

**Comment.** It is rare, but sometimes authors do not observe the Principle of Priority (Code Art. 23), by not choosing the oldest available nomen to denote a taxon.

### Gonyleptidae Sundevall, 1833

#### *Neopachylus incertus* (Mello-Leitão, 1935)

- *Huralvius incertus* Mello-Leitão 1935c: 382, fig. 12.

*Neopachylus incertus*: Kury 2003a: 178.

- *Nunduavius nebulosus* Mello-Leitão 1936b: 11, fig. 8. **II** Junior subjective synonym of *Huralvius incertus* Mello-Leitão, 1935 by Montemor, Bragagnolo & Pinto-da-Rocha (2015: 116), **with inverted precedence.**

#### *Neopachylus marginatus* (Mello-Leitão, 1931)

- *Gephyropachylus marginatus* Mello-Leitão 1931b: 14.  
*Neopachylus marginatus*: Kury 2003a: 178.
- *Neopachylus serrinha* Soares & Soares 1947d: 222, figs 9–10. **II** Junior subjective synonym of *Gephyropachylus marginatus* Mello-Leitão, 1931 by Montemor, Bragagnolo & Pinto-da-Rocha (2015: 118), **with inverted precedence.**

### Podoctidae Roewer, 1912

#### *Strandibalonius* Roewer, 1912 revalidated

- *Strandibalonius* Roewer 1912c: 199. Type-species by monotypy: *Ibalonius abnormis* Strand, 1911.
- *Metibalonius* Roewer 1915c: 32. Type-species **by subsequent designation, hereby proposed**: *Metibalonius longipalpis* Roewer, 1915; subsequent designation of *Ibalonius cervicornis* Strand, 1910 as type-species by Goodnight & Goodnight (1957: 77) is invalid because that species was not originally included. **II** Junior subjective synonym of *Strandibalonius* Roewer, 1912 by Goodnight & Goodnight (1957: 77), **with reversed precedence.**
- *Homibalonius* Roewer 1915c: 39. Type-species **by subsequent designation, hereby proposed**: *Homibalonius obscurus* Roewer, 1915. **II** Junior subjective synonym of *Metibalonius* Roewer, 1915 by Goodnight & Goodnight (1957: 77).

Melanesia, Micronesia.

**Comment.** Based on reasoning put forth by Kury & Machado (2018), *Strandibalonius* should be transferred from Ibaloniinae to Erecananinae, where it is situated closest to *Lomanius* Roewer, 1923.

**Comment.** Roewer (1915c: 32) described the new genus *Metibalonius* Roewer, 1915 to include two new species from Papua New Guinea – *Metibalonius longipalpis* Roewer, 1915 and *Metibalonius gracilipes* Roewer, 1915. However, neither he nor any subsequent author designated a type species. As *Metibalonius* dates from before 1931 it is available; nevertheless, *Metibalonius longipalpis* Roewer, 1915 is designated herewith as type, following ICZN Rec. 69A.9.

**Comment.** Roewer (1915c: 39) described the new genus *Homibalonius* Roewer 1915 to include two new species from Papua New Guinea – *Homibalonius obscurus* Roewer, 1915 and *Homibalonius scaber* Roewer, 1915. However, neither he nor any subsequent author designated a type species. As *Homibalonius* dates from before 1931 it is available; nevertheless, *Homibalonius obscurus* Roewer, 1915 is designated herewith as type, following ICZN Rec. 69A.9. *Homibalonius* is currently a

junior synonym of *Strandibalonius* (as per Goodnight & Goodnight 1957).

***Strandibalonius biantipalpis* (Roewer, 1915) comb. nov.**

- *Reclinobunus biantipalpis* Roewer 1915c: 37.  
*Metibalonius biantipalpis*: Goodnight & Goodnight 1957: 77.

***Strandibalonius cervicornis* (Strand, 1911) comb. nov.**

- *Ibalonius cervicornis* Strand 1911c: 4.  
*Sitalces cervicornis*: Roewer 1912c: 199, fig. 43.  
*Metibalonius cervicornis*: Roewer 1923: 167, figs 185a–b.

***Strandibalonius esakii* (Suzuki, 1941) comb. nov.**

- *Metibalonius esakii* Suzuki 1941c: 100, figs 2–3, pl. 7, figs 9–16.

***Strandibalonius femoralis* (Roewer, 1949) comb. nov.**

- *Fronticonus femoralis* Roewer 1949c: 257, figs 29a–f.  
*Metibalonius femoralis*: Goodnight & Goodnight 1957: 77.

***Strandibalonius gracilipes* (Roewer, 1915) comb. nov.**

- *Metibalonius gracilipes* Roewer 1915: 34, fig. 17.

***Strandibalonius longipalpis* (Roewer, 1915) comb. nov.**

- *Metibalonius longipalpis* Roewer 1915: 33, fig. 16.

***Strandibalonius obscurus* (Roewer, 1915) comb. nov.**

- *Homibalonius obscurus* Roewer 1915: 40.  
*Metibalonius obscurus*: Goodnight & Goodnight 1957: 77.

***Strandibalonius oppositus* (Roewer, 1927) comb. nov.**

- *Serratobunus oppositus* Roewer 1927a: 308, fig. 20.

***Strandibalonius spinatus* (Roewer, 1949) comb. nov.**

- *Manema spinata* Roewer 1949c: 258, figs 31a–b.  
*Metibalonius spinatus*: Goodnight & Goodnight 1957: 77.

***Strandibalonius spinulatus* (Roewer, 1915) comb. nov.**

- *Serratobunus spinulatus* Roewer 1915: 43.  
*Metibalonius spinulatus*: Goodnight & Goodnight 1957: 77.

***Strandibalonius strucki* (Goodnight & Goodnight, 1947) comb. nov.**

- *Metibalonius strucki* Goodnight & Goodnight 1947: 332, figs 4–5.

***Strandibalonius tenuis* (Roewer, 1949) comb. nov.**

- *Celebesia tenuis* Roewer 1949c: 257, fig. 28a–e.  
*Metibalonius tenuis*: Goodnight & Goodnight 1957: 77.

***Strandibalonius triceratops* (Kury & Machado, 2018) comb. nov.**

- *Trispinibunus abnormis* Roewer 1915: 46. **II** Junior secondary homonym of *Ibalonius abnormis* Strand, 1910, by virtue of the synonymy of *Trispinibunus* Roewer, 1915 with *Strandibalonius* Roewer, 1912; first noted by Kury & Machado (2018: 163).

*Metibalonius abnormis*: Goodnight & Goodnight 1957: 77.

*Metibalonius triceratops*: Kury & Machado 2018: 163.

***Strandibalonius yalomensis* (Suzuki, 1982) comb. nov.**

- *Metibalonius yalomensis* Suzuki 1982b: 206, figs 74–78.

**Comment.** Goodnight & Goodnight (1957: 77) synonymized nine genera with *Metibalonius* Roewer, 1915, including *Strandibalonius* Roewer, 1912. They **mistakenly** indicated the establishment of *Metibalonius* as of 1912, on page 39 of a paper that does not contain this nomen. Page 39 deals with species of *Assamia*. Therefore, *Strandibalonius* is the oldest available synonym to be used, if we decide to take Goodnight & Goodnight's taxonomic decision at face value. Furthermore, the authors only explicitly combined one species with *Metibalonius* (plus, of course the type-species of each synonymized genus), so a few species have to be formally combined with *Strandibalonius* here. Two of those nine genera, *Homibalonius* Roewer 1915, do not have type-species designated, and they are designated herein. Technically, none of the *Homibalonius* species has been combined with *Metibalonius*, because it has no type-species until now.

**Sclerosomatidae Simon, 1879**

**Genus *Psathyropus* L. Koch, 1878**

- *Psathyropus* L. Koch 1878: 785. **I** Type-species by monotypy: *Psathyropus tenuipes* L. Koch, 1878.
- *Metagagrella* Roewer 1910a: 95. **I** Type-species by original designation: *Gagrella sordidata* Thorell, 1889. **II** Junior subjective synonym of *Psathyropus* L. Koch, 1878 by Suzuki (1973d: 250), **with inverted precedence.**

 SE Asia.

**Comment.** *Metagagrella* Roewer, 1910 was considered a junior subjective synonym of *Psathyropus*, L. Koch 1878, by Suzuki (1973d: 250), who, however, inverted the precedence, as Koch's name is more than 30 years older than Roewer's. Crawford (1992: 40) commented on that, refraining however, from making the necessary new combinations: "Suzuki may have intended to reject the long unused name *Psathyropus* under ICZN Article 23b, then in force, but did not state so; under the current articles 23b and 79c .2(iv), *Psathyropus* is valid."

Therefore, hitherto we have several **valid** species combined with the **invalid** genus *Metagagrella*. Only the type-species, *Psathyropus tenuipes*, was combined with *Psathyropus*, and **the rest of the needed combinations are made here**. We are not assuming the monophyly of *Psathyropus* sensu Suzuki, but as the taxonomy of Gagrellinae is in very bad shape now, the new combinations reflect an educated guess as to the inclusion of



those species but more crucially, stabilise the currently chaotic taxonomy of this group.

***Psathyropus aurolucens* (Roewer, 1954) comb. nov.**

- *Metagagrella aurolucens* Roewer 1954c: 250, pl. 23, fig. 66.

***Psathyropus bengalensis* (Roewer, 1954) comb. nov.**

- *Metagagrella bengalensis* Roewer 1954c: 241, pl. 23, fig. 54.

***Psathyropus bilineatus* (Roewer, 1954) comb. nov.**

- *Metagagrella bilineata* Roewer 1954c: 242, pl. 23, fig. 56.

***Psathyropus bimaculatus* (Roewer, 1954) comb. nov.**

- *Metagagrella bimaculata* Roewer 1954c: 246, pl. 23, fig. 63.

***Psathyropus biseriatus* (Roewer, 1912) comb. nov.**

- *Metagagrella biseriata* Roewer 1912b: 37.

***Psathyropus cingulatus* (Roewer, 1954) comb. nov.**

- *Metagagrella cingulata* Roewer 1954c: 240.

***Psathyropus conjugatus* (Roewer, 1954) comb. nov.**

- *Metagagrella conjugata* Roewer 1954c: 245, pl. 23, fig. 61.

***Psathyropus conspicuus* (Roewer, 1954) comb. nov.**

- *Metagagrella conspicua* Roewer 1954c: 241, pl. 23, fig. 53.

***Psathyropus cuprilucidus* (Roewer, 1954) comb. nov.**

- *Metagagrella cuprilucida* Roewer 1954c: 242.

***Psathyropus distinctus* (Sato & Suzuki, 1938) comb. nov.**

- *Metagagrella distincta* Sato & Suzuki 1938b: 379, figs 5–6.

***Psathyropus formosa* (Roewer, 1911) comb. nov.**

- *Metagagrella formosa* Roewer 1911d: 594, pl. 16, figs 1–2.

***Psathyropus granulatus* (Roewer, 1954) comb. nov.**

- *Metagagrella granulata* Roewer 1954c: 246, pl. 23, fig. 62.

***Psathyropus granulosis* (Suzuki, 1977) comb. nov.**

- *Metagagrella granulosa* Suzuki 1977a: 65, figs. 28A-F, 50E-F.

***Psathyropus guttatus* (Roewer, 1954) comb. nov.**

- *Metagagrella guttata* Roewer 1954c: 242, pl. 23, fig. 55.

***Psathyropus hainanensis* (Wang, 1941) comb. nov.**

- *Metagagrella hainanensis* Wang 1941: 103, figs 4a–d.

***Psathyropus hirtus* (Roewer, 1915) comb. nov.**

- *Metagagrella hirta* Roewer 1915: 146.

***Psathyropus hsuehshanensis* (Suzuki, 1977) comb. nov.**

- *Metagagrella hsuehshanensis* Suzuki 1977b: 136, figs 6N-T.

***Psathyropus koyamai* (Suzuki, 1979) comb. nov.**

- *Metagagrella koyamai* Suzuki 1979: 142, figs 1–17.

***Psathyropus luteomaculatus* (Suzuki, 1970) comb. nov.**

- *Metagagrella luteomaculata* Suzuki 1969b: 66, fig. 36; pl. IV, figs 1–36.



**Figure 4.** *Psathyropus tenuipes* L. Koch, 1878, male, in vivo, from Japan, Shizuoka Prefecture. Image copyright © Ryosuke Kuwahara (Kumamoto University), used with permission, cropped.

***Psathyropus mandalayius* (Roewer, 1954) comb. nov.**

- *Metagagrella mandalayia* Roewer 1954c: 244, pl. 23, fig. 60.

***Psathyropus micans* (Roewer, 1926) comb. nov.**

- *Metagagrella micans* Roewer 1926c: 558, pl. 2, fig. 5.

***Psathyropus minax minax* (Thorell, 1889) comb. nov.**

- *Gagrella minax minax* Thorell 1889: 638.

***Psathyropus minax trispinosus* (Thorell, 1889) comb. nov.**

- *Gagrella minax trispinosa* Thorell 1889: 638.

***Psathyropus mysoreanus* (Roewer, 1954) comb. nov.**

- *Metagagrella mysoreana* Roewer 1954c: 240, pl. 22, fig. 50.

***Psathyropus niger* (Roewer, 1912) comb. nov.**

- *Metagagrella nigra* Roewer 1912b: 36.

***Psathyropus octomaculatus* (Roewer, 1954) comb. nov.**

- *Metagagrella octomaculata* Roewer 1954c: 240, pl. 23, figs 51–52.

***Psathyropus perakanus* (Roewer, 1954) comb. nov.**

- *Metagagrella perakana* Roewer 1954c: 245.

***Psathyropus pustulatus* (Roewer, 1910) comb. nov.**

- *Metagagrella pustulata* Roewer 1910a: 97, pl. 2, fig. 15.

***Psathyropus roeweri* (Suzuki, 1974) comb. nov.**

- *Metagagrella roeweri* Suzuki 1974e: 138, figs 1–8, 10–11.

***Psathyropus rufoscutum* (Suzuki, 1982) comb. nov.**

- *Metagagrella rufoscuta* Suzuki 1982d: 173, figs 4A–H.

***Psathyropus rufus* (Roewer, 1954) comb. nov.**

- *Metagagrella rufa* Roewer 1954c: 243, pl. 23, fig. 57.

***Psathyropus satarensis* (Roewer, 1954) comb. nov.**

- *Metagagrella satarensis* Roewer 1954c: 239, pl. 22, fig. 49.

***Psathyropus silvestrii* (Roewer, 1927) comb. nov.**

- *Metagagrella silvestrii* Roewer 1927c: 209.

***Psathyropus sinensis* (Schenkel, 1953) comb. nov.**

- *Metagagrella sinensis* Schenkel 1953: 91.

***Psathyropus sordidatus* (Thorell, 1889) comb. nov.**

- *Gagrella sordidata* Thorell 1889: 634.

***Psathyropus sulcatus* (Roewer, 1954) comb. nov.**

- *Metagagrella sulcata* Roewer 1954c: 243, pl. 23, fig. 59.

***Psathyropus tenuis* (Roewer, 1954) comb. nov.**

- *Metagagrella tenuis* Roewer 1954c: 243, pl. 23, fig. 58.

***Psathyropus tongkingensis* (Roewer, 1954) comb. nov.**

- *Metagagrella tongkingensis* Roewer 1954c: 246, pl. 23, fig. 64.

***Psathyropus versicolor* (Suzuki, 1964) comb. nov.**

- *Metagagrella versicolor* Suzuki 1964c: 123, figs 3, 4, 5A–D.



## Chapter 3. Other nomenclatural and taxonomic acts

We decided to keep new allocations, new synonymies and combinations to a minimum here. However, we included quite a few that were on the way as parts of scattered projects, some of them interrupted by the devastating fire of the Museu Nacional in 2018 (Kury et al., 2018). Several initial results from those projects are reflected here. We decided to implement those changes in the formal taxonomy in the best interest of advancing the taxonomy of Opiliones.

### 3.1. Genera hitherto without type-species

In this section we deal with subsequent designations of type-species for several genera. The cases may be grouped into six subsections:

#### 3.1.1. TWO GENERA ONLY MADE AVAILABLE HEREIN

There are two generic names described after 1931, which are unavailable, because there is neither original type species fixation or subsequent designation. They are herein made available by being described as new genera. There was no need to use the same original nomina, but it was deemed best, in the spirit of stability to keep them, because they occur in the literature and are already associated with the species nomina.

#### Family Epedanidae Sørensen, 1886

##### *Asopella* Kury gen. nov.

urn:lsid:zoobank.org:act:ADC1F734-E6F5-4C24-9CB6-06BD-5F4D0554

- *Asopella* Henriksen 1932a: 255. Unavailable, Code Art. 13.3.

**Comment.** Henriksen (1932a), based on the notes left by Sørensen, described the genus *Asopella* (credited to himself) as new to include three species – *Epedanus lutescens* Thorell, 1876 plus two new species (which he correctly attributed to Sørensen): *Asopella xanti* Sørensen, 1932 and *Asopella bicolor* Sørensen, 1932. None of the subsequent authors who treated the genus

(Roewer 1938b; Suzuki 1982b) designated a type-species. *A. bicolor* was later transferred to genus *Caletor* Loman, 1892 (Roewer 1938b: 128), and a fourth species, *A. robusta*, was described by Suzuki (1982b). *Asopella* is unavailable and a formal description is given herewith.

**Etymology.** Aiming to maintain stability, the same unimaginative nomen used by Henriksen is kept here. It comes from pre-existing genus *Asopus* + Latin diminutive suffix *-ella*. Gender feminine.

**Included species.** *Epedanus lutescens* Thorell, 1876 (type species, designated herewith), *Asopella xanti* Sørensen, 1932 and *Asopella robusta* Suzuki, 1982.

**Diagnosis (adapted from Henriksen, 1932, Roewer 1938 and Suzuki, 1982).** Sarasinicinae: Ocularium with median spine of variable length. Mesotergum divided into 4 disjoint unarmed scutal areas. Pedipalpus: Femur substraight, ventrally with 6-8 and mesal-apical with 1–2 megaspines, Patella ventro-mesal with 2 and ventro-ectal with 1 megaspine. Legs: Tarsal counts 8–14(3) / 20–28 (3) / 7–9 / 8–10. Tarsal claws of third and fourth tarsi each with 1 inner prong.

##### *Sinistus* Kury gen. nov.

urn:lsid:zoobank.org:act:BE545E06-9C1B-46F4-A636-6B-1FB3484C42

- “*Sinistus*” Roewer 1938: 151. Unavailable, Code Art. 13.3.

**Comment.** Roewer (1938) described the new genus *Sinistus* Roewer 1938 in Phalangodidae: Sarasinicinae to include two new species from Borneo – *Sinistus maculatus* Roewer, 1938 and *Sinistus fuscus* Roewer, 1938 – without choosing a type species from among these two. However, Code article 13.3 states: “To be available, every new genus-group name published after 1930... must... be accompanied by the fixation of a type species in the original publication.” There is no further citation of the nomen *Sinistus* in the literature. Therefore, *Sinistus* is unavailable and a formal description is given herewith.

**Etymology.** Aiming to maintain stability, the same nomen used by Roewer is kept here. It comes from Latin *sinistus*, which was the office of chief priest among the Burgundians (from Ammianus Marcellinus, *Rerum gestarum*). Gender masculine.

**Included species.** *Sinistus maculatus* Roewer, 1938 (type species, designated herewith) and *Sinistus fuscus* Roewer, 1938.

**Diagnosis (adapted from Roewer 1938).** Sarasinicinae: Ocularium with long median spine. Scutal areas I to IV unarmed. Pedipalpus: Femur straight, ventrally with 7 and mesal-apical with 4 megaspines, Patella ventro-mesal with 2 and ventro-ectal with 1 megaspine. Legs: Tarsal counts 10–11(3) / 25–31 (2) /



7–11 / 8–15. Tarsal claws smooth, without accessory prongs.

### 3.1.2. AVAILABLE GENERIC NOMINA DESCRIBED BEFORE 1931 BUT WHICH LACK A TYPE-SPECIES

**Comment.** There is a fairly large amount of generic names described **before 1930**, which are **available** (and either valid or invalid), but which have **originally included two or more species**. However, none of those species has been selected as type by either the original author or any subsequent author. The designation of type-species for those genera is made herewith following **Code Arts. 66–69** and applying all items of Recommendation 69A. In some cases, these designations affect synonymy. Because these genera are already available, a type-species may be designated, regardless of they being invalid. It is paramount for a genus to have a type-species, among other things because if its original species are someday deemed not to be congeneric, one has to know to which **onomatophore** the nomen is attached.

#### Family Assamiidae Sørensen, 1884

##### *Chilon* Sørensen, 1896

- *Chilon* Sørensen 1896: 187. 🐛 Type-species by subsequent designation of Staręga (1992): *Chilon robustus* Sørensen, 1896.
- *Parachilon* Roewer 1923: 280. 🐛 Type-species **by subsequent designation, hereby proposed**: *Chilon ferrugineus* Roewer, 1912, currently in synonymy with *Chilon robustus* Sørensen, 1896. II Junior subjective synonym of *Chilon* Sørensen, 1896 by Staręga (1992: 300).

##### *Hypoxestus* Loman, 1902

- *Hypoxestus* Loman 1902: 192. 🐛 Type-species by monotypy: *Hypoxestus levis* Loman, 1902.
- *Amasis* Sørensen 1910: 76. 🐛 Type-species **by subsequent designation, hereby proposed**: *Amasis patellaris* Sørensen, 1910. II Junior homonym of *Amasis* Leach, 1817 (Hymenoptera), first noted by Strand (1911d: 14). II Junior subjective synonym of *Hypoxestus* Loman, 1902 by Roewer (1912c: 88).

##### *Macrodampetrus* Roewer, 1915

- *Macrodampetrus* Roewer 1915: 73. 🐛 Type-species **by subsequent designation, hereby proposed**: *Macrodampetrus bicoloripes* Roewer, 1915.

##### *Maracandus* Simon, 1879

- *Maracandus* Simon 1879: 186. 🐛 Type-species **by subsequent designation, hereby proposed**: *Maracandus macei* Simon, 1879.

##### *Mosoia* Roewer, 1912

- *Mosoia* Roewer 1912c: 17. 🐛 Type-species **by subsequent designation, hereby proposed**: *Maracandus albiceps* Loman, 1906.

##### *Senarba* Roewer, 1927

- *Senarba* Roewer 1927a: 377. 🐛 Type-species **by subsequent designation, hereby proposed**: *Senarba rudicoxa* Roewer, 1927.

#### Family Cosmetidae Koch, 1839

##### *Metacynortoides* Roewer, 1912

- *Metacynortoides* Roewer 1912d: 65. 🐛 Type-species **by subsequent designation, hereby proposed**: *Cynorta scabrosa* Banks, 1909. II Junior subjective synonym of *Cynorta* C.L. Koch, 1839 by Goodnight & Goodnight (1953b: 38); synonymy disclaimed by Kury (2003a).

#### Family Epedanidae Sørensen, 1886

##### *Dibunus* Loman, 1906

- *Dibunus* Loman 1906b: 5. 🐛 Type-species by monotypy: *Dibunus pseudobiantes* Loman, 1906.
- *Anacudorsum* Roewer 1926c: 551. 🐛 Type-species **by subsequent designation, hereby proposed**: *Anacudorsum bakeri* Roewer, 1926. II Junior subjective synonym of *Dibunus* Loman, 1906 by Goodnight & Goodnight (1957: 73).
- *Dibunellus* Roewer 1927a: 341. 🐛 Type-species **by subsequent designation, hereby proposed**: *Dibunellus albitarsus* Roewer, 1927. II Junior subjective synonym of *Dibunus* Loman, 1906 by Goodnight & Goodnight (1957: 73).

##### *Heteroepedanus* Roewer, 1912

- *Heteroepedanus* Roewer 1912c: 233. 🐛 Type-species **by subsequent designation, hereby proposed**: *Epedanus monacantha* Roewer, 1911.

##### *Metathyreotus* Roewer, 1913

- *Metathyreotus* Roewer 1913b: 206. 🐛 Type-species **by subsequent designation, hereby proposed**: *Metathyreotus aborensis* Roewer, 1913.

##### *Metepedanus* Roewer, 1912

- *Metepedanus* Roewer 1912c [May]: 234. 🐛 Type-species **by subsequent designation, hereby proposed**: *Epedanus venator* Roewer, 1911.
- *Parabiantes* Hirst 1912 [August]: 82. 🐛 Type-species by monotypy: *Parabiantes longipalpis* Hirst, 1912. II Junior subjective synonym of *Metepedanus* Roewer, 1912 by Roewer (1923).

##### *Opelytus* Roewer, 1927

- *Opelytus* Roewer 1927a: 330. 🐛 Type-species **by subsequent designation, hereby proposed**: *Opelytus vepretum* Roewer, 1927.



**Comment.** *Opelytus* is probably formed from Greek ὀπή (hole) + λυτός (that may be untied, loosed or dissolved), gender masculine. It should have been *Lytope* (M. Alonso-Zarazag, pers. comm.). It possibly refers to the circle formed by the clenched chelicerae of male as depicted in Roewer's figure 30b, meaning that the hole disappears when the animal opens the chelicerae. As for the specific epithet, Latin *vēprētum* (thornhedge, bramble-thicket) is a noun in apposition, referring to the thorny aspect of the pedipalps.

### ***Pseudobiantes* Hirst, 1911**

- *Pseudobiantes* Hirst 1911b: 632. Type-species by original designation: *Pseudobiantes japonicus* Hirst 1911.
- *Strisilvea* Rower 1927c: 197. Type-species **by subsequent designation, hereby proposed**: *Strisilvea cavicola* Roewer, 1927, a junior synonym of *Epedanellus tuberculatus* Roewer, 1911 by Suzuki (1973d: 231). Junior subjective synonym of *Pseudobiantes* Hirst, 1911 by Suzuki (1985d: 56).

### **3.1.3. A MORE COMPLEX CASE: THE FORGOTTEN GENUS *HETEROBIANTES***

The designation of a type species further affects nomenclature in this case: *Epedanus geniculatus* Pocock, 1903 is herein designated as type species of *Heterobiantes* Roewer, 1912, and, accordingly, *Epedanestus* Roewer 1938 is considered a **junior objective synonym** of *Heterobiantes* Roewer, 1912.

#### **Family Epedanidae Sørensen, 1886**

#### **Subfamily Acrobuninae Roewer, 1912**

#### ***Heterobiantes* Roewer, 1912**

- *Heterobiantes* Roewer 1912c: 219. 🐞 Type-species **by subsequent designation, hereby proposed**: *Epedanus geniculatus* Pocock, 1903.
- *Epedanestus* Roewer 1938: 108. 🐞 Type-species by monotypy: *Epedanus geniculatus* Pocock, 1903. II Junior objective synonym of *Heterobiantes* Roewer, 1912, **first noted here**.

**Etymology.** From Greek ἕτερος (of another kind, different) + pre-existing genus *Biantes*. Gender masculine.

**Placement.** *Heterobiantes* originally in Phalangodidae: Acrobuninae.

**Comment.** With the *imbroglio* created by Roewer (1938), the name *Heterobiantes* was entirely forgotten, but it has priority over *Epedanestus* in the case of *E. geniculatus* being the type-species. The history of *Pseudobiantes*, *Heterobiantes* and related genera of Epedanidae is complex, and may be summarized as follows:

1) Hirst (1911b) described the new genus *Pseudobiantes* Hirst, 1911 without reference to family, but said it was closest to *Epedanus* Thorell, 1876, therefore including it in Epedanidae, sensu Roewer (1911b). Hirst originally included three species in *Pseudobiantes*, two of them described as new: *Pseudobiantes japonicus* Hirst, 1911 (which he designated as type species), *Pseudobiantes insulanus* Hirst, 1911 and *Epedanus geniculatus* Pocock, 1903. However, curiously, in his introduction, he misspelled the new genus as "*Heterobiantes*". Incidentally, these were the first Laniatores to be described from Japan.

2) Roewer (1912c: 216) described the new subfamily Acrobuninae Roewer, 1912 in Phalangodidae to include three genera: *Acrobunus* Thorell, 1891, *Pseudobiantes* Hirst, 1911 and the **new genus *Heterobiantes* Roewer, 1912** (perhaps using the idea of the name coined by Hirst). He left only *Pseudobiantes japonicus* in *Pseudobiantes*, and removed the other two species — *Epedanus geniculatus* Pocock, 1903 and *Pseudobiantes insulanus* Hirst 1911 — to form *Heterobiantes*. However, he designated neither as the type species for the genus.

3) Roewer (1915c) described in Sarasinellinae the monotypic **genus *Kilungius***, along with its type species *Kilungius bimaculatus* Roewer, 1915, from Taiwan.

4) Roewer (1923: 190, ff.) kept the same arrangement of *Pseudobiantes* with one Japanese species and *Heterobiantes* with two species — one from Okinawa and the other from Hong Kong.

5) Roewer (1938b: 109) stated that he finally was able to examine the types of *Epedanus geniculatus* Pocock, 1903 and accordingly, he transferred the species from the Acrobuninae to the Epedaninae, creating the **new monotypic genus *Epedanestus*** for this species. He cited *Heterobiantes* and *Pseudobiantes* only as "part" in the synonymy of *Epedanestus*.

6) A few pages below, Roewer (1938b: 138–139) transferred *Pseudobiantes insulanus* Hirst 1911 to *Kilungius* in Sarasinicinae.

7) By moving both original species of *Heterobiantes* to different genera, without establishing any generic synonymy, ***Heterobiantes* as a genus was entirely forgotten** by Roewer.

8) This makes *Heterobiantes* a senior synonym either of *Epedanestus* or *Kilungius*, **depending on which species is designated as type**.

9) We herewith pick *Epedanus geniculatus* Pocock, 1903 as the type species of *Heterobiantes*, which makes the scantily cited *Epedanestus* a junior synonym of this,

thus preserving the name *Kilungius*, which has been considerably more cited in the literature.

### 3.1.4. GENERIC NOMINA SUBSEQUENTLY MADE AVAILABLE BETWEEN 1931 AND 1999 (CODE ART. 13.3 AND 16.1)

**Comment.** Several generic nomina were established after 1930 without the fixation of a type species from among the two or more included species. This renders them *nomina nuda* (Code Art. 13.3). However, the subsequent action of choosing one of originally included species as the type-species before 2000 makes them available even if no intention of authors to establish new nominal taxa was explicit (Code Art. 16.1). There are several generic nomina that were made available after their original descriptions, mostly inadvertently, between 1931 and 1999 under these conditions (after 1999 the author should explicitly state that a new taxon was established). These nomina are listed here to make sure of their correct authorship citations in the future, because such nomenclatural acts can be easily overlooked. Some of them were already listed in Kury & Alonso-Zarazaga (2011) and Kury et al. (2014: 14).

#### Family Triaenonychidae Sørensen, 1886

##### ***Austromontia* Starega, 1992**

“*Austromontia*” Lawrence 1931: 398. 🚫 Unavailable because published after 1930 without fixation of a type species from among the three originally included (Code Art. 13.3): *Austromontia caledonica* Lawrence, 1931, *Austromontia capensis* Lawrence, 1931 and *Austromontia silvatica* Lawrence, 1931.

- *Austromontia* Starega 1992: 282 [first made available]. 🐞 Type-species by original designation: *Austromontia silvatica* Lawrence, 1931.

##### ***Biacumontia* Starega, 1992**

“*Biacumontia*” Lawrence 1931: 403. 🚫 Unavailable because published after 1930 without fixation of a type species from among the four originally included (Code Art. 13.3): *Biacumontia cornuta* Lawrence, 1931, *Biacumontia fissidens* Lawrence, 1931, *Biacumontia paucidens* Lawrence, 1931 and *Biacumontia truncatidens* Lawrence, 1931.

- *Biacumontia* Starega 1992: 283 [first made available]. 🐞 Type-species by original designation: *Biacumontia paucidens* Lawrence, 1931.

##### ***Callihamus* Hickman, 1958**

“*Callihamus*” Roewer 1931d: 161. 🚫 Unavailable because published after 1930 without fixation of a type species from among the two originally included (Code Art. 13.3): *Callihamus badius* Roewer, 1931 and *Callihamus inermis* Roewer, 1931.

- *Callihamus* Hickman 1958: 49 [first made available]. 🐞 Type-species by original designation: *Callihamus badius* Roewer, 1931.

##### ***Graemontia* Starega, 1992**

“*Graemontia*” Lawrence 1931: 413. 🚫 Unavailable because published after 1930 without fixation of a type species from among the two originally included (Code Art. 13.3): *Graemontia bifidens* Lawrence, 1931 and *Graemontia dentichelis*.

- *Graemontia* Starega 1992: 285 [first made available]. 🐞 Type-species by original designation: *Graemontia bifidens* Lawrence, 1931.

##### ***Larifugella* Starega, 1992**

“*Larifugella*”: Lawrence 1933: 226. 🚫 Unavailable because published after 1930 without fixation of a type species from among the two originally included (Code Art. 13.3): *Larifugella afra* Lawrence, 1933 and *Adaeulum natalense* Lawrence, 1931.

- *Larifugella* Starega 1992: 279 [first made available]. 🐞 Type-species by original designation: *Larifugella afra* Lawrence, 1933.

##### ***Mensamontia* Starega, 1992**

“*Mensamontia*” Lawrence 1931: 381. 🚫 Unavailable because published after 1930 without fixation of a type species from among the two originally included (Code Art. 13.3): *Mensamontia melanophora* Lawrence, 1931 and *Mensamontia morulifera* Lawrence, 1931.

- *Mensamontia* Starega 1992: 286 [first made available]. 🐞 Type-species by original designation: *Mensamontia morulifera* Lawrence, 1931.

##### ***Monomontia* Starega, 1992**

“*Monomontia*” Lawrence 1931: 416. 🚫 Unavailable because published after 1930 without fixation of a type species from among the two originally included (Code Art. 13.3): *Monomontia atra* Lawrence, 1931 and *Monomontia rattrayi* Lawrence, 1931.

- *Monomontia* Starega 1992: 287 [first made available]. 🐞 Type-species by original designation: *Monomontia atra* Lawrence, 1931.

##### ***Rostromontia* Starega, 1992**

“*Rostromontia*” Lawrence 1931: 388. 🚫 Unavailable because published after 1930 without fixation of a type species from among the four originally included (Code Art. 13.3): *Rostromontia capensis* Lawrence, 1931, *Rostromontia granulifera* Lawrence, 1931, *Rostromontia lisposoma* Lawrence, 1931 and *Rostromontia truncata* Lawrence, 1931.

- *Rostromontia* Starega 1992: 288 [first made available]. 🐞 Type-species by original designation: *Rostromontia truncata* Lawrence, 1931.



## Phalangodidae Simon, 1879

### *Bunofagea* Starega, 1992

“*Bunofagea*”: Lawrence 1959: 82. 📄 Unavailable because published after 1930 without fixation of a type species from among the two originally included (Code Art. 13.3): *Bunofagea gracilipes* Lawrence, 1959 and *Bunofagea remyi* Lawrence, 1959.

- *Bunofagea* Starega 1992: 295. 🐛 Type-species by original designation: *Bunofagea gracilipes* Lawrence, 1959.

## Assamiidae Sørensen, 1884

### *Cereatta* Starega, 1992

“*Cereatta*” Roewer 1935a: 78. 📄 Unavailable because published after 1930 without fixation of a type species from among the two originally included (Code Art. 13.3): *Cerea celeripes* Loman 1910 and *Cereatta elegans* Roewer, 1935.

- *Cereatta* Starega 1992: 299 [first made available]. 🐛 Type-species by original designation: *Cerea celeripes* Loman, 1910.

### *Cryptopygoplus* Starega, 1992

“*Cryptopygoplus*” Lawrence 1931: 360. 📄 Unavailable because published after 1930 without fixation of a type species from among the three originally included (Code Art. 13.3): *Cryptopygoplus africanus* Lawrence, 1931, *Cryptopygoplus damaranus* Lawrence, 1931 and *Cryptopygoplus rhodesianus* Lawrence, 1931.

- *Cryptopygoplus* Starega 1992: 301 [first made available]. 🐛 Type-species by original designation: *Cryptopygoplus africanus* Lawrence, 1931.

“*Lawrenciola*” Roewer 1935: 65. 📄 Unavailable because published after 1930 without fixation of a type species from among the two originally included (Code Art. 13.3): *Cryptopygoplus damaranus* Lawrence, 1931 and *Cryptopygoplus rhodesianus* Lawrence, 1931.

- *Lawrenciola* Starega 1992: 309 [first made available]. 🐛 Type-species by original designation: *Cryptopygoplus damaranus* Lawrence, 1931. II Junior subjective synonym of *Cryptopygoplus* Lawrence, 1931 by Starega & Snegovaya (2009)].

### *Erecella* Starega, 1992

“*Erecella*”: Roewer 1935a: 53. 📄 Unavailable because published after 1930 without fixation of a type species from among the two originally included (Code Art. 13.3): *Erecella flava* Roewer, 1935 and *Erecella lutea* Roewer, 1935.

- *Erecella* Starega 1992: 304 [first made available]. 🐛 Type-species by original designation: *Erecella lutea* Roewer, 1935.

### *Marua* Starega, 1992

“*Marua*” Roewer 1935a: 104. 📄 Unavailable because published after 1930 without fixation of a type species from among the two originally included (Code Art. 13.3): *Marua schenkeli* Roewer, 1935 and *Marua spinosa* Roewer, 1935.

- *Marua* Starega 1992: 311 [first made available]. 🐛 Type-species by original designation: *Marua spinosa* Roewer, 1935.

### *Rhabdopygella* Starega, 1992

“*Rhabdopygella*” Roewer 1935a: 18. 📄 Unavailable because published after 1930 without fixation of a type species from among the two originally included (Code Art. 13.3): *Rhabdopygella ferruginea* Roewer, 1935 and *Rhabdopygella laevis* Roewer, 1935.

- *Rhabdopygella* Starega 1992: 317 [first made available]. 🐛 Type-species by original designation: *Rhabdopygella laevis* Roewer, 1935.

## Cranidae Roewer, 1913

### *Angistrisoma* Soares & Soares, 1948

“*Angistrisoma*”: Roewer 1932: 338. 📄 Unavailable because published after 1930 without fixation of a type species from among the two originally included (Code Art. 13.3): *Angistrisoma atroluteum* Roewer, 1932 and *Angistrisoma fuscum* Roewer, 1932.

- *Angistrisoma* Soares & Soares 1948b: 587 [first made available]. 🐛 Type-species by original designation: *Angistrisoma fuscum* Roewer, 1932.

## Cosmetidae C.L. Koch, 1839

### *Prasiana* Strand, 1942

“*Prasia*” [subgenus of *Cynorta*] Sørensen in Henriksen 1932a: 379.

📄 Unavailable because published after 1930 without fixation of a type-species from among the six originally included (Code Art. 13.3): *Cynorta (Prasia) clypeata* Sørensen, 1932, *Cynorta (Prasia) fallax* Sørensen, 1932, *Flirtea phalerata* C.L. Koch, 1840, *Cosmetus pictus* Perty, 1833, *Cynorta quadripustulata* Simon, 1879 and *Cynorta (Prasia) venezuelana* Sørensen, 1932.

- *Prasia* Mello-Leitão 1933c: 108, 113 [first made available]. 🐛 Type-species by original designation: *Cynorta (Prasia) fallax* Sørensen, 1932. II Junior homonym of *Prasia* Stål, 1863 (Hemiptera)
- *Prasiana* Strand 1942: 399. 🐛 Type species automatically (ICZN Code Art. 67.8) the same as for *Prasia*. 📄 Nomen novum for *Prasia* Sørensen, 1932.

### *Zaraxolia* Strand, 1942

“*Zarax*”: Sørensen in Henriksen 1932a: 321. 📄 Unavailable because published after 1930 without fixation of a type-species from among the two originally included (Code Art. 13.3): *Zarax aenescens* Sørensen, 1932 and *Zarax devians* Sørensen, 1932.

- *Zarax* Mello-Leitão 1933c: 114 [first made available]. 🐛 Type-species by monotypy: *Zarax devians* Sørensen, 1932. II Junior homonym of *Zarax* Pascoe, 1867 (Coleoptera); junior subjective synonym of *Paecilaema* C.L. Koch, 1839 by Goodnight & Goodnight (1953b).
- *Zaraxolia* Strand 1942: 400. 📄 Nomen novum for *Zarax* Mello-Leitão, 1933. 🐛 Type species automatically (Code Art. 67.8) the same as for *Zarax* [the subsequent designation of *Z. aenescens* Sørensen, 1932 by Roewer (1947) is invalid because Mello-Leitão had already fixated *Zarax devians* as type].

## Sclerosomatidae Simon, 1879

### *Chasenella* Roewer, 1955

“*Chasenella*” Roewer 1933b: 46. 🚫 Unavailable because published after 1930 without fixation of a type-species from among the two originally included (Code Art. 13.3): *Chasenella luma* Roewer, 1933 and *Chasenella pakka* Roewer, 1933.

- *Chasenella* Roewer 1955a: 73 [first made available]. 🐛 Type-species by original designation: *Chasenella luma* Roewer, 1933.

### 3.1.5. UNAVAILABLE GENERIC NOMINA WHICH CANNOT BE MADE AVAILABLE HERE BY DESIGNATION OF A TYPE-SPECIES BECAUSE THEY ARE CURRENTLY TREATED AS JUNIOR SYNONYMS (CODE ART. 11.6.3)

### *Holoscotolemon* Roewer, 1915

“*Scotolemoniscus*” Roewer 1935b: 42. 🐛 Type-species: none designated. 🚫 Unavailable because published after 1930 without fixation of a type species from among the two originally included (Code Art. 13.3): *Scotolemoniscus austriacus* Roewer, 1935 and *Scotolemoniscus styriacus* Roewer, 1935. II Junior subjective synonym of *Holoscotolemon* Roewer, 1915 by Martens (1978b: 76).

“*Brigestus*” Roewer 1935b: 45. 🐛 Type-species: none designated. 🚫 Unavailable because published after 1930 without fixation of a type species from among the two originally included (Code Art. 13.3): *Metascotolemon granulatus* Roewer, 1915 and *Brigestus robustus* Roewer, 1935. II Junior subjective synonym of *Holoscotolemon* Roewer, 1915 by Martens (1978b: 76).

### *Ischyropsalis* C.L. Koch, 1839

“*Odontopalpa*” [subgenus of *Ischyropsalis*] Hadži 1931: 113. 🐛 Type-species: none designated. 🚫 Unavailable because published after 1930 without fixation of a type species from among the three originally included (Code Art. 13.3): *Ischyropsalis bosnica* Roewer 1914, *Ischyropsalis dentipalpis* Canestrini, 1872 and *Ischyropsalis (Odontopalpa) triglavensis* Hadži 1931. II Junior subjective synonym of *Ischyropsalis* C.L. Koch, 1839 by Martens (1969: 189).

### *Neosadocus* Mello-Leitão, 1926

“*Bunoweyhia*” Mello-Leitão 1935d: 18. 🐛 Type-species: none designated. Subsequent designation of *Bunoweyhia variabilis* Mello-Leitão, 1935 by Kury (2003a) is invalid because he treated it as a junior synonym (Code Art. 11.6.3). 🚫 Unavailable because published after 1930 without fixation of a type species from among the two originally included (Code Art. 13.3): *Bunoweyhia minor* Mello-Leitão, 1935 and *Bunoweyhia variabilis* Mello-Leitão, 1935. II Junior subjective synonym of *Neosadocus* Mello-Leitão, 1926 by B. Soares (1944e: 248).

### *Obidosus* Roewer, 1931

“*Fonteboatus*” Roewer 1931e: 156. 🐛 Type-species: none designated. Subsequent designation of *Fonteboatus coxalis* Roewer, 1931 by Pinto-da-Rocha (1997a: 275) is invalid because he treated it as a junior synonym (Code Art. 11.6.3). 🚫 Unavailable because published after 1930 without fixation of a type-species from among the two originally included (Code Art. 13.3): *Fonteboatus coxalis* Roewer, 1931 and *Fonteboatus palpalis* Roewer, 1931. II Junior subjective synonym of *Protimesius* Roewer, 1913 by H. Soares (1978b: 75); junior subjective synonym of *Obidosus* Roewer, 1931 by Villarreal et al. (2019: 978).

### *Promitobates* Roewer, 1913

“*Batomites*” Mello-Leitão 1931c: 140. 🐛 Type-species: none designated; subsequent designation of *Batomites difficilis* Mello-Leitão, 1931 by Kury (2003a) is invalid because he treated it as a junior synonym (Code Art. 11.6.3). 🚫 Unavailable because published after 1930 without fixation of a type species from among the two originally included (Code Art. 13.3): *Batomites difficilis* Mello-Leitão, 1931 and *Batomites spitzi* Mello-Leitão, 1931. II Junior subjective synonym of *Promitobates* Roewer, 1913 by B. Soares (1943f: 57).

### *Stygnoplus* Simon, 1879

“*Ilhastygnus*” Roewer 1943: 63. 🐛 Type-species: none designated. Subsequent designation of *Ilhastygnus dominicanus* Roewer, 1943, by Pinto-da-Rocha (1997a: 197) is invalid because he treated it as a junior synonym (Code Art. 11.6.3). 🚫 Unavailable because published after 1930 without fixation of a type species from among the two originally included (Code Art. 13.3): *Ilhastygnus antiguanus* Roewer, 1943 and *Ilhastygnus dominicanus* Roewer, 1943. II Junior subjective synonym of *Stygnoplus* Simon, 1879 by Pinto-da-Rocha (1997a: 197).

### *Peltonychia* Roewer, 1935

“*Kratochviliola*” Roewer 1935b: 64. 🐛 Type-species: none designated. 🚫 Unavailable because published after 1930 without fixation of a type species from among the three originally included (Code Art. 13.3): *Kratochviliola cavernicola* Roewer, 1935, *Kratochviliola navarica* Roewer, 1935 and *Kratochviliola parolinia* Roewer, 1935. II Junior subjective synonym of *Hadziana* Roewer, 1935, which was a junior subjective synonym of *Peltonychia* Roewer, 1935 by Martens (1978b: 70); briefly revalidated by Kury & Mendes (2007: 14) who thought *Peltonychia* was unavailable; finally synonymized with *Peltonychia* by Kury et al. (2014: 13).



## 3.2. Correction of valid species combined with invalid genera

Sometimes, when a genus *Bus* is synonymized, not all of its members end up referred to a valid genus *Aus*. The remaining species are often noted in the literature by writing “*Bus*” between quotation marks. This indicates that the authors know the usage might be incorrect but prefer not to commit to seeking a solution at that moment. In a given ergotaxonomy which does not recognize that genus, those species then lie suspended in limbo, creating a situation where we have valid species combined with invalid genera. Electronic indexing systems deal with this in their own way: “Leaving those names under deliberately incorrect usage may lead to other kinds of problems. Electronic resources for taxonomy mostly do not distinguish between a **true usage** of a generic nomen and a **loose usage** of the same name in another family between quotation marks, and then the system **assumes they are homonyms**, thus creating an error message.” [Kury & Snegovaya 2019].

Here (and also in chapters 4, 5, 6 and 11), a new combination is made for each of those valid species so they are linked to valid genera. All **based on new taxonomic decisions**, which were not covered in the original works.

Imagine a taxonomic publication “A” where the author synonymizes genus *Bus* (with 10 species) with genus *Aus*, but treating only the type-species of *Bus* – called *Bus xus* – which they newly combine as *Aus xus*. However, if none of the nine remnant valid species of *Bus* has been treated in work “A”, this generates a situation where all of them are **linked to the invalid genus *Bus***, at least in the light of the ergotaxonomy adopted there. In the case of a text-based catalogue, one could simply write “*Bus*” *yus*, “*Bus*” *zus*, etc, the **quotes** denoting that assignment of those species to genus *Bus* is **inertial and noncommittal**. However, in the case of an electronic catalogue, the system does not allow linking a valid species to an invalid genus. Therefore, we herein present new combinations for those species which do not necessarily share the fate of the type-species. The rationale for these combinations proposed here are **educated guesses as to the likely taxonomic allocations of those species** based on the present accumulated knowledge of Opiliones.

### Phalangiidae Latreille, 1802

#### *Egaenus laevipes* (Caporiacco, 1935) **comb. nov.**

- *Diabunus laevipes* Caporiacco 1935: 245.

**Comment.** *Diabunus* Thorell 1876 was considered a junior subjective synonym of *Egaenus* C.L. Koch, 1839 by Staręga (1978: 222), who did not mention this species. We concur with this synonymy, and accordingly, we herewith formally transfer it also to *Egaenus* creating the new combination *Egaenus laevipes* (Caporiacco, 1935) **comb. nov.**

### Sclerosomatidae Simon, 1879

#### *Pectenobunus argentinus* (Ringuelet, 1959) **comb. nov.**

- *Caiza argentina* Ringuelet 1959: 218.

**Comment.** The genus *Caiza* was synonymized with *Pectenobunus* by Tourinho (2004a), but this species has not been treated there, so it is herewith also tentatively ascribed to *Pectenobunus*, based on Tourinho’s diagnosis.

### Trogulidae Sundevall, 1833

#### *Calathocratus kyrgyzicus* (Chemeris, 2013) **comb. nov.**

- *Trogulocratus kyrgyzicus* Chemeris 2013: 43, figs 7- 16, map 2.

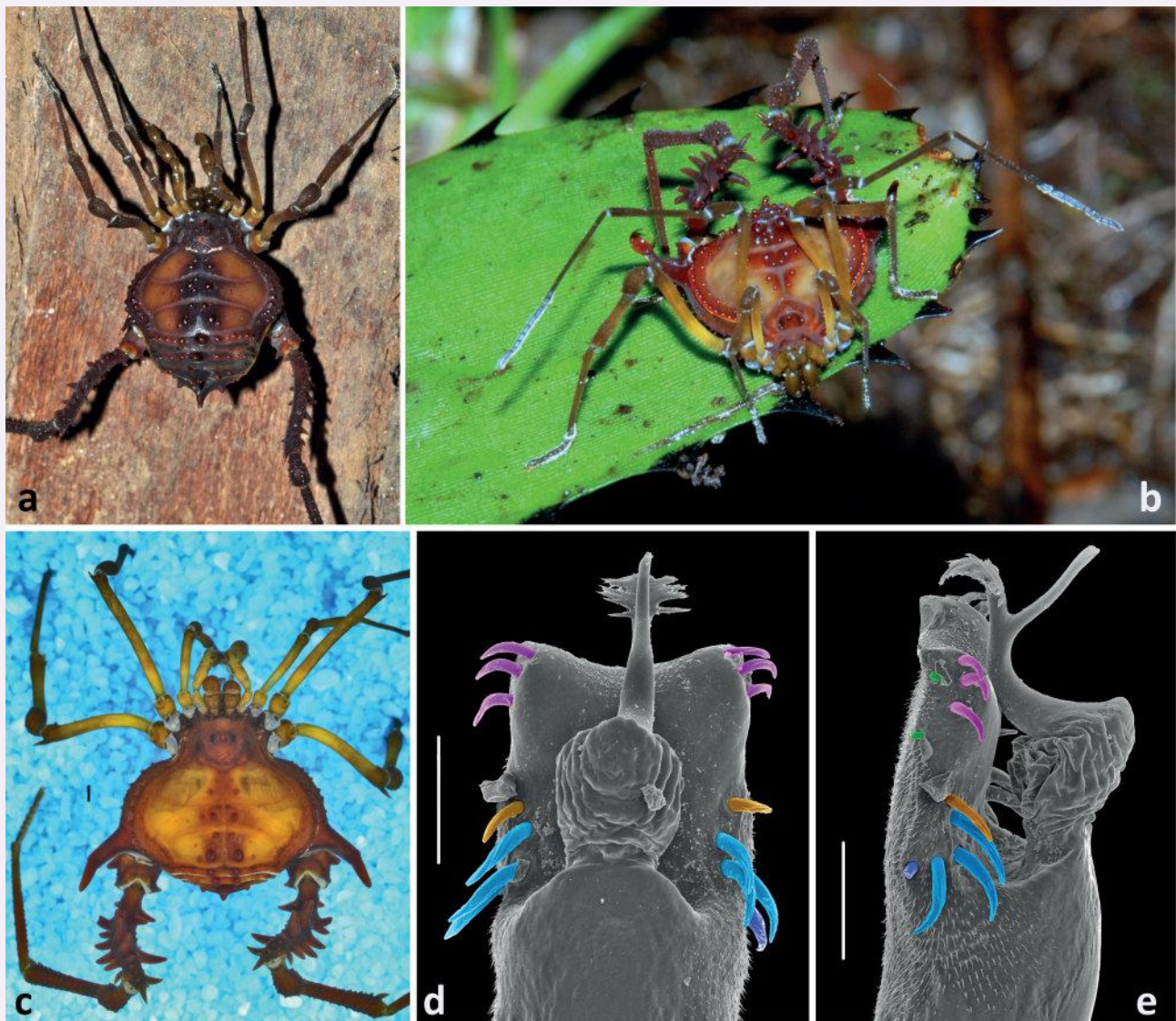
**Comment.** *Trogulocratus* Roewer, 1940 was considered a junior subjective synonym of *Calathocratus* Simon, 1879 by Schönhofer & Martens (2010: 71). Three years later, Chemeris (2013: 43) described a new species from Kyrgyzstan in *Trogulocratus*. No subsequent authors dealt with this species. The presentation of a name as a synonym is not a nomenclatural act in zoology. So one cannot say a name was qualified as a synonym in 2010 and subsequently it is not possible to describe a new species in such a synonymous genus. The 2013 paper is an independent publication and authors are allowed to adopt a totally different classification, completely independent from the 2010 work. Nevertheless, we are constrained by the ergotaxonomy that we are adopting here. So, **we have to transfer *Trogulocratus kyrgyzicus* to an adequate genus that is valid here in WCO.** Chemeris did not formally propose a reinstatement of *Trogulocratus*. After studying Chemeris description, and accepting Schönhofer & Martens classification with *Trogulocratus* as a junior synonym of *Calathocratus*, we herein transfer this species to *Calathocratus*, creating the new combination *Calathocratus kyrgyzicus* (Chemeris, 2013) **comb. nov.**

**Gonyleptidae Sundevall, 1833*****Pachylibunus nigripes* (Mello-Leitão, 1942) comb. nov.**

- *Caldasius nigripes* Mello-Leitão 1942a: 162, fig. 4.

**Comment.** *Caldasius* Roewer 1930 was considered a junior subjective synonym of *Bourguyia* Mello-Leitão, 1923 by Yamaguti & Pinto-da-Rocha (2009: 333), with which we concur. However, this species is not congeneric with the type-species. Already Kury (2003a: 153) listed this as “Pachylinae incertae sedis” and remarked that it was definitely not a member of the family Bourguiiinae.

Examination of the type material in MNRJ (now destroyed) and comparison with several species of bona fide *Pachylibunus* Roewer, 1913 allowed the inclusion of this species in *Pachylibunus*, which had species already known from Brazilian MG and RJ states. Therefore, we transfer *Caldasius nigripes* to the genus *Pachylibunus* creating the new combination *Pachylibunus nigripes* comb. nov.



**Figure 5.** *Pachylibunus nigripes* (Mello-Leitão, 1942) comb. nov.: (a). Female from Domingos Martins, ES, in vivo; (b). Male from Castelo, ES, in vivo; (c). Male (UFMG 13218) habitus, dorsal view; (d). (MNRJ 4547) male genitalia, distal part, macrosetae artificially colored, dorsal view; (e). Same, sinistrolateral view. Photographs Copyright ©: A.B. Kury (a, c, d, e), Pedro Henrique Martins (b). All used with permission. Artificial color conventions for ventral plate macrosetae are: A (blue), B (dark blue), C (magenta), D (golden yellow), E (green).



### 3.3. Other new combinations and synonymies

#### Phalangiidae Latreille, 1802

##### *Metaplathybunus obliquus* (C.L. Koch, 1839) **comb. nov.**

- *Opilio obliquus* C.L. Koch 1839c: 33.

**Comment:** Simon (1884d) said this species probably belongs to *Dasylobus*. Later, Martens (1978b) [our translation] commented: "Roewer (1912) sees *Opilio obliquus* CL Koch, 1839 Greece as a synonym of *Opilio glacialis* Heer 1845. However, this assignment cannot be right, because Koch (1849) represents the armature of pedipalps differently in both species, namely with without a medial apophysis at Pt and Ti in *obliquus* and *glacialis*. The oldest available and unique name, *glacialis* Heer must be used for the alpine species. The Greek species *obliquus* CL Koch probably belongs to the genus *Metaplathybunus*." This species is here placed in *Metaplathybunus* based on that observation by Martens. Thus, the new combination *Metaplathybunus obliquus* (C.L. Koch, 1839) **comb. nov.** is hereby formally proposed.

#### Podoctidae Roewer, 1912

##### *Ibalonius dubius* (Goodnight & Goodnight, 1948) **comb. nov.**

- *Philibalonius dubius* Goodnight & Goodnight 1948a: 7, figs 1–2.
- *Ibalonius rainbowi* Forster 1949b: 146, figs 7–9. **Syn. nov.**

**Comment:** *Ibalonius rainbowi* Forster 1949b: 146, figs 7–9 newly considered a junior subjective synonym of *Philibalonius dubius* Goodnight & Goodnight 1948: 7, figs 1–2 **syn. nov.**, which is herewith transferred from *Philibalonius* to *Ibalonius* in virtue of the synonymy of both genera proposed by Suzuki (1977a: 29). The result is *Ibalonius dubius* (Goodnight & Goodnight, 1948) **comb. nov.** A.B. Kury and G. Machado examined material of this species and carefully studied both descriptions; besides being sympatric, there is no significant difference between them.



# Chapter 4. A ghost species of “*Metaphalangium*” (Eupnoi: Phalangidae)

by AB Kury

It is a common misconception among taxonomists that when someone combines the type-species of genus *Aus* with older genus *Bus*, proposing thus a synonymy involving both, all the other species of *Aus* should also **automatically** enter into *Bus*. They do not. Rather, they should be deliberately placed somewhere. This is a taxonomic, not nomenclatural decision, and has to be made case by case.

The following example illustrates one of such cases, which cumulates with other nomenclatural faulty procedures.

1) Nosek (1905: 150) described from “Kleinasien, near Nigde” [= Turkey, Niğde, Niğde] the new species *Phalangium strandi* Nosek, 1905.

2) Roewer (1911e: 49) transferred *P. strandi* to *Paropilio*, where it remained to date (with at least 5 citations, 1923–2012). The last citation of this combination was Snegovaya (2012: 5).

3) Caporiacco (1949: 83) described from Rhodes the new species *Metaphalangium strandi* Caporiacco, 1949.

4) Both Martens (1965) and Gruber (1978) suggested that *Metaphalangium strandi* Caporiacco, 1949 was a junior subjective synonym of *Zachaeus crista* (Brullé, 1832).

5) Martens (1965: 62): “...liegt der Verdacht nahe, daß es sich um den auf Rhodos häufigen und sehr variablen *Zacheus crista* (Brulle) gehandelt hat.” [= it is reasonable to suspect that it was the *Zacheus crista* (Brulle), which is common and very variable on Rhodes.] (Fig. 6).

6) Gruber (1978: 571): “Von Rhodos beschrieb Di Caporiacco (1948) nach wenigen Stücken ein *Metaphalangium strandi*, das — wie schon MARTENS (1965) annahm — mit obiger Art identisch sein dürfte” [= Di Caporiacco (1948) based on a few specimens



**Figure 6.** *Zachaeus crista*, male, in vivo, from Sithonia, Greece. Image copyright © Walter Pfliegler, used with permission.



described from Rhodes *Metaphalangium strandi*, which – as Martens (1965) assumed – should be identical to the above species].

7) Staręga (1984: 42) considered *Paropilio* Roewer 1911 to be a junior subjective synonym of *Metaphalangium* Roewer, 1911, but did not combine any other *Paropilio* other than the type-species, *Phalangium abstrusum* L. Koch, 1882, under *Metaphalangium*.

8) Özdikmen et al. (2010), assumed that once *Paropilio* was synonymized with *Metaphalangium*, all species of the former became combined with the latter. In that case, *Metaphalangium strandi* Caporiacco 1949 would be indeed a junior secondary homonym of *Phalangium strandi* Nosek, 1905. Özdikmen and collaborators then did the formal combination for the latter and proposed a **nomen novum** for the former.

9) The nomen *Metaphalangium rhodesensis* has formation problems. The correct classical adjectives for Rhodes Island (Ῥόδος) are **rhodius**, **rhodiacus** or **rhodensis**, otherwise it may be confused with *rhodesiensis*, from Rhodesia, Africa. However, this mis-formation cannot be emended (Art. 32.5.1). *Rhodesensis* is herein considered a Neo-Latin adjective and as such should be declined, if this name is in use in the future (Art. 34.2).

10) In spite of the implicit proposal by Özdikmen and collaborators that *Paropilio strandi* should accompany *Paropilio abstrusus* in the transfer to *Metaphalangium*, I decide herein to follow the suggestions of Martens and Gruber, and to **formalize the synonymy** of Caporiacco's species with *Zachaeus crista* (Fig. 6).

11) Besides *Phalangium strandi* Nosek 1905, there are three other *Paropilio* species which should, in principle, belong to *Metaphalangium* as shown implicitly by the synonymy of both genera. However, this did not make available the combinations. Those three species were pending to be explicitly combined and they are herein formally coupled with *Metaphalangium*.

### ***Metaphalangium* Roewer, 1911**

- *Metaphalangium* Roewer 1911e: 70. 🐛 Type-species by original designation: *Phalangium propinquum* Lucas, 1846.
- *Paropilio* Roewer 1911e: 48. 🐛 Type-species by original designation: *Phalangium abstrusum* L. Koch, 1882. II Junior subjective synonym of *Metaphalangium* Roewer, 1911 by Staręga (1984).

🌐 Circum-Mediterranean, Balkans, Near East, Caucasus.

**Etymology.** From Greek μετά (after) + pre-existing genus *Phalangium*. **Gender neuter.**

### ***Metaphalangium monticola* (Mkheidze, 1952) comb. nov.**

- *Paropilio monticola* Mkheidze 1952a: 547, fig. 3.

### ***Metaphalangium punctatum* (Roewer, 1956) comb. nov.**

- *Paropilio punctatus* Roewer 1956b: 297, pl. 41 figs 162–165.

### ***Metaphalangium spinipes* (Roewer, 1956) comb. nov.**

- *Paropilio spinipes* Roewer 1956: 300, pl. 41 figs 166–169.

### ***Metaphalangium strandi* (Nosek, 1905)**

- *Phalangium Strandii* Nosek 1905: 150.  
*Paropilio Strandii*: Roewer 1911e: 49.  
*Metaphalangium strandi*: Özdikmen et al. 2010: 45.

### ***Zachaeus* C.L. Koch, 1839**

- *Zachaeus* C.L. Koch 1839a: 152. 🐛 Type-species by monotypy: *Zachaeus mordax* C.L. Koch, 1839. 📄 Made available before 1931 as a combined description and without being stated as new (Code Art. 12.2.6).

🌐 Circum-Mediterranean, Balkans, Near East, Caucasus.

### ***Zachaeus crista* (Brullé, 1832)**

- *Phalangium crista* Brullé 1832: 60, pl. 28, fig. 12.  
*Zachaeus crista*: Simon 1879c: lxxi [71].
- *Metaphalangium strandi* Caporiacco 1949: 83. II Junior secondary homonym of *Phalangium strandi* Nosek, 1905, only if included in *Metaphalangium*, for which there is no current support. **Syn. nov.**
- *Metaphalangium rhodesensis* Özdikmen, Kurt & Demir 2010: 46. 📄 Incorrect originally applied gender declination. 📄 Superfluous nomen novum for *Metaphalangium strandi* Caporiacco, 1949. **Syn. nov.**

🌐 Adriatic, SE Appenines, Balkans, Carpathians, Aegean islands, W Anatolia, Anatolian Isles.

# Chapter 5. A Brazilian “*Daguerreia*” (Laniatores: Gonyleptidae)

by RN Carvalho, AB Kury & MR Hara

Subfamily Roeweriinae Carvalho & Kury, 2018

Genus *Khazaddum* gen. nov.

urn:lsid:zoobank.org:act:E07B3F33-44CD-4FD8-96F1-33AA2F-8FB543

**Etymology.** From the fictional language Khuzdul, created by J. R. R. Tolkien in his work “The Lord of the Rings” Khazad-dûm (the fabulous subterranean city-kingdom of the Dwarves), in reference to the caves where it is often found. Gender neuter (as specified here, as it is unknown if the original language Khuzdul possessed genders, Code, Art. 30.2.2).

**Included species.** *Daguerreia inermis* Soares & Soares 1947 (type-species).

**Diagnosis.** *Khazaddum* gen. nov. resembles the other Roeweriinae species due to: VP basal portion twice as wide as the distal; macrosetae (MS) C1–C3 elongated; stylus depressed, with apical flaps; the presence of two spines on the bulla cheliceral ectal portion; coxa IV with a retroventral apophysis, without a geminated branch.

*Khazaddum* gen. nov. can be separated from the other Roeweriinae due to: VP with four MS A (A1–A4); MS B elongated, with almost equal size of the MS A1–A4; stylus and axis of its ventral process fused basally, forming a large pedestal; ventral process with a half (or a third) of the length of the stylus, with a spiny flabellum; pair of spines on the posterior face of the ocularium; scutal areas I–IV unarmed, coxa IV short and femur IV without remarkable armature; coxa IV prodorsal distal apophysis as a reduced spine (Figs. 7a, b).

**Geographic distribution.** BRAZIL: states of Paraná and São Paulo. Biogeographic areas used here are from the WWF Terrestrial Eco-regions of the World (names starting with “NT”; Olson et al. 2001, Fig. 9).

*Khazaddum inermis* (Soares & Soares, 1947) comb. nov.

• *Daguerreia inermis* Soares & Soares 1947d: 217, fig. 4;

**Type data.** *Daguerreia inermis*: ♂ holotype, MHNCI 3629 (not examined) from BRAZIL, state of Paraná, Bocaiúva do Sul, Gruta de Campinhos.

**Records.** BRAZIL: state of Paraná: Almirante Tamandaré: Água Boa cave. Bocaiúva do Sul: Jesuítas cave. Campo Largo: Pinheirinho cave. Castro: Olhos d’Água cave. Colombo: Bacaetava cave. Rio Branco do Sul: Itacolombo cave; Lancinha cave (Pinto-da-Rocha, 1993). BRAZIL: state of São Paulo: Apiaí: Aranhas cave; Pescaria cave. Guapiara: Rio Preto cave system. Iporanga: Águas Quentes cave; Água Suja cave; Alambari de Cima cave; Barra Bonita cave; Casa da Pedra cave; Colorida cave; Floido cave; Gurutuva cave; Tatu cave (Gnaspini & Trajano, 1994). BRAZIL: state of São Paulo: Apiaí: Chapéu cave. Iporanga: Abismo da Chuva; Alambari de Baixo cave; Arataca cave; Espírito Santo cave; Jane Mansfield cave; Laje Branca cave; Morro Preto I cave; Paivas cave; Pérolas cave; Resurgência das Areias; Santana cave, Temimina II cave (Pinto-da-Rocha, 1995). BRAZIL: state of Paraná: Arapoti; Campo Largo: Canavial cave. Telêmaco Borba: Harmonia. BRAZIL: state of São Paulo: Iporanga: Fendão cave; Jerivassal cave; Queijo Suíço cave (Pinto-da-Rocha, 1996).

**Material examined.** BRAZIL: state of Paraná: Rio Branco do Sul: 1 ♂ (MNRJ-HS 0890), 1 ♀ (MNRJ-HS 0891), Gruta da Lancinha, 31.v.1987, Pinto da Rocha, R. leg. BRAZIL: state of São Paulo: Apiaí: 1 ♂ (MNRJ 17877), Gruta das Aranhas, 25.vi.1989, Trajano, E. leg.; 2 ♀ (MNRJ 17878), Gruta do Chapéu, 25.vi.1989, Trajano, E. leg. Iporanga: 1 ♂ (MNRJ-HS 0868), Caverna Águas Quentes, Godoy, N. M. leg.; 1 ♂ 1 ♀ (MNRJ-HS 0898), idem, 02.v.1986, Gnaspini Netto, P. leg.; 1 ♂ (MNRJ-HS 0867), 1 ♀ (MNRJ-HS 0874), Caverna Água Suja, Trajano, E. leg.; 1 ♀ (MNRJ 17427), Caverna Alambari de Baixo; 1 ♂ (MNRJ-HS 0865), 1 ♀ (MNRJ-HS 0866), 1 ♂ (MNRJ-HS 0893) idem, Trajano, E. leg.; 1 ♀ (MNRJ-HS 0863), idem, 04.ii.1984, Trajano, E. leg.; 1 ♂ (MNRJ-HS 0872), Caverna Alambari de Cima, 17.ii.1985, Trajano, E. leg.; 1 ♂ 1 ♀ (MNRJ 6786), Fazenda Intervalles, 20.iii.1989, Kury, A. B. & Baptista, R. L. C. leg.; 1 ♂ 2 ♀ (MNRJ 6803), Gruta Casa de Pedra, 18.iv.1991, Baptista, R. L. C. leg.; 1 ♂ 1 ♀ (MNRJ-HS 0896), idem, Gnaspini Netto, P. leg.; 1 ♂ (MNRJ-HS 0897), Gruta Colorida, Gnaspini Netto, P. leg.; 1 ♂ (MNRJ-HS 0894), Gruta do Fendão, Gnaspini Netto, P. leg.; 2 ♂ (MNRJ-HS 0895), Gruta do Floido, Gnaspini Netto, P. leg.; 1 ♂ (MNRJ 8893), Gruta Laboratório I, 06.iv.2012, Ferreira, R. L. leg.; 1 ♂ 1 ♀ (MNRJ 2520), Gruta da Pescaria, 05.vi.2011, Bichuette, M. E. et al. leg.; 1 ♂ (MNRJ-HS 0822), Caverna Santana, viii.1981, Trajano, E. leg.

**Geographic distribution.** BRAZIL: Paraná: Almirante Tamandaré, Arapoti, Bocaiúva do Sul, Castro, Colombo,



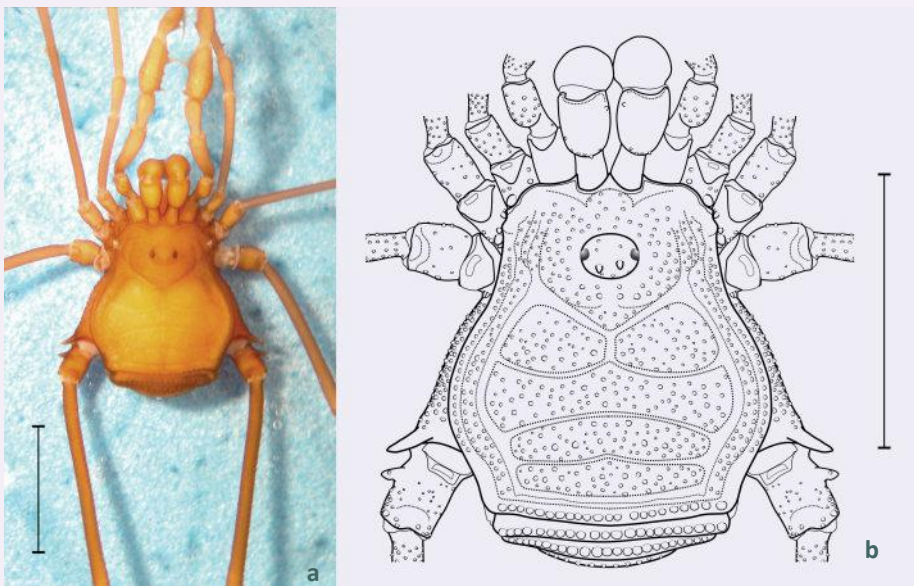
Rio Branco do Sul, Telêmaco Borba. São Paulo: Apiaí, Guapiara, Iporanga.

**Diagnosis.** As for genus.

**Description.** The original description of the female by Soares & Soares (1947d: 217, fig. 4), supplemented by the description of the male by Pinto-da-Rocha (1996c: 834, figs 1–7) are enough to characterize this species adequately. We present here more detailed images of the male external body and genitalia.

**Historical taxonomic remarks.** Soares & Soares (1947d: 217) described “the second species of *Daguerreia*, being the first from Brazil” based on a single

female from a cave in Paraná, Southern Brazil. In spite of their non-enunciation of characters which would lead to this generic assignment, it is fairly easy to conclude that it was based on features used in the **Roewerian system** (such as armature of ocularium, scutal areas and free tergites and tarsal counts), which back then was the current paradigm. It was not surprising, then, to discover that Soares’ *Daguerreia inermis* did not share any special similarity with the original Argentinean *Daguerreia* (which later was synonymized with *Pachyloides*). Already Acosta (1996) and Kury (2003) **recognized that *D. inermis* was not a member of *Pachyloides*** (as opposed the type-species of *Daguerreia*), but they chose not to assign it to any genus.



**Figure 7.** *Khazaddum inermis* (Mello-Leitão, 1935) comb. nov., (MNRJ-HS 867), male, specimen in alcohol: (a). Habitus, dorsal view, picture of the burned specimen; (b). Same, dorsal view, vectorized drawing. Scale bars: a–b, 5 mm.



**Figure 8.** *Khazaddum inermis* (Soares & Soares, 1947) comb. nov., (MNRJ-HS 867), penis, distal part: (a). Dorsal view; (b). Lateral view; (c). Ventral view; (d). Stylus, apical view, showing ventral process. Scale bars: 0.1 mm (a, c–d); 0.02 mm (b).

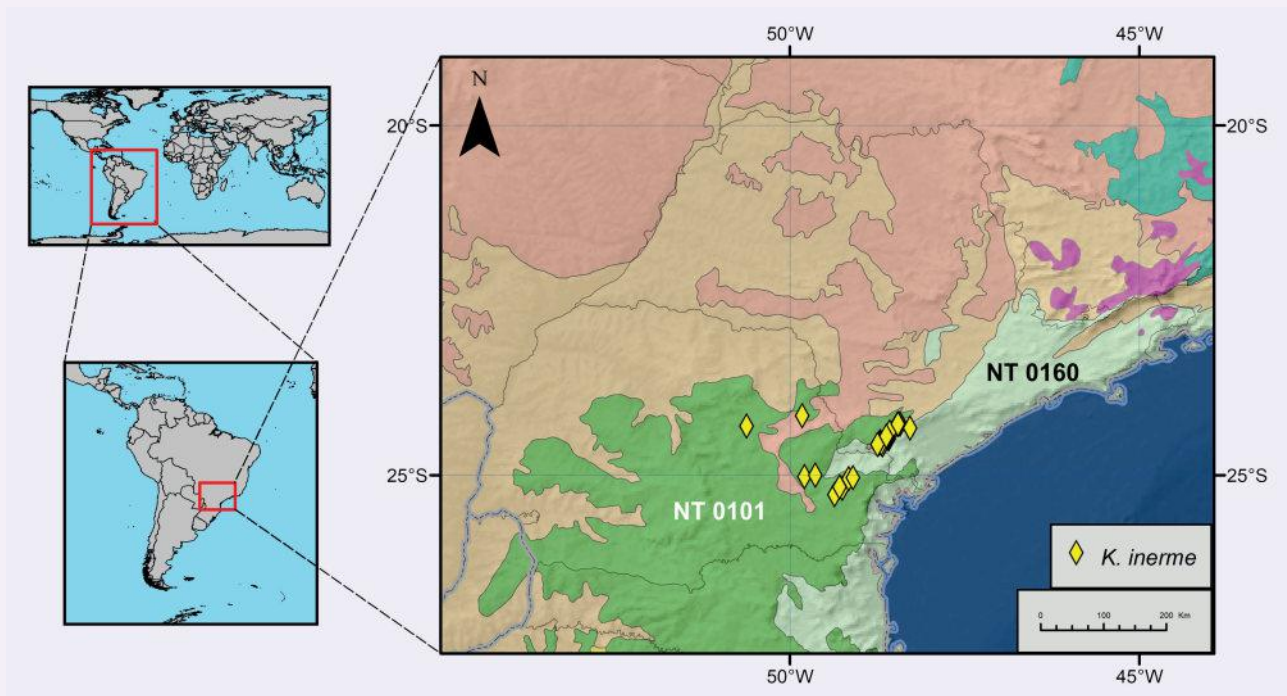
Four decades later, H. Soares (1978a: 78, figs 18–24) described the male “allotype” of *Planiphalangodus robustus* Roewer, 1929. However, we consider it as a misidentification of *K. inermis* comb. nov. or some very closely related species (the male genitalia is not a perfect match), hitherto undescribed. Males of *Planiphalangodus robustus*, examined by us, are different from them. Only twenty years later, the male of *D. inermis* was made known to the scientific world (Pinto-da-Rocha 1996), including illustrations of the male genitalia.

In an ongoing project that deals with an expansion of the Roeweriinae concept (Carvalho, Kury & Hara in prep.), we performed a phylogenetic analysis to assess the placement of some species of “Pachylinae” suspected of being Roeweriinae, including *K. inermis* comb. nov. We decided to anticipate to this eBook the combination of *D. inermis* because this configured an untenable situation of a valid species linked with an invalid genus (as other similar cases presented here in Section 3.2).

Based on the results of the above mentioned cladistic analysis, we herein propose the transfer of the hitherto incertae sedis “*Daguerreia*” *inermis* to Roeweriinae. However, *D. inermis* did not join any individual genus, rather being sister-group to a cluster of

other genera. This requires the creation of a new genus, which is formalized here as *Khazaddum* gen. nov. and a new combination *Khazaddum inermis* comb. nov.

This species does not share unambiguous synapomorphies with the rest of Roeweriinae, but can be recognized by the lack of armature on the mesotergum and femur IV (Fig. 7), contrasting all the remaining diversity of Roeweriinae. However, some of its genital features (e.g., VP, stylus and MS C) are similar to those presented in the original diagnosis of this subfamily (Fig. 8). This species is restricted to the Araucaria Forest biome, with its records shared between the Brazilian states of Paraná and São Paulo (Fig. 9).



**Figure 9.** Southeastern Brazil, showing the distribution of *Khazaddum inermis* (Soares & Soares, 1947) comb. nov. Shaded areas on the background are WWF terrestrial eco-regions: NT 0101 (Araucaria moist forests, in dark green) and NT 0160 (Serra do Mar coastal forests, in light green).



# Chapter 6. New synonymies and combinations in the clade K92

by AB Kury & LN de Ázara

Some synonymies established for genera of K92 in the past failed to treat all species included in the junior synonyms, and therefore left several valid species attached to invalid genera. We here address this problem and try to assign those orphan species to valid genera. What allowed us to tackle those issues are several scattered notes on the taxonomy of K92, amassed during the last years, which still have not found their way into the press. As these do not belong in our larger manuscripts they are presented here, serving both taxonomy and nomenclature. Terminology of male leg IV follows Ázara & Kury (2019).

## Gonyleptidae Sundevall, 1833

### *Acanthogonyleptes* Mello-Leitão, 1922

- *Acanthogonyleptes* Mello-Leitão 1922: 336. 🐞 Type-species by original designation: *Acanthogonyleptes pulcher* Mello-Leitão, 1922.

### *Acanthogonyleptes asperulus* (Roewer, 1930) **comb. nov.**

- *Moreira asperula* Roewer 1930: 433, fig. 38.  
*Paragonyleptes asperulus*: Piza 1942b: 414.

**Comment.** *Moreira asperula* was originally described in Moreira Roewer 1930, a genus later (DaSilva & Pinto-da-Rocha 2010: 626) synonymized with *Acanthogonyleptes*. This species had been long before incorrectly transferred to *Paragonyleptes* Roewer, 1913 by Piza (1942b: 414), which prevented the discovery of its relationships, as it appeared in Kury's catalogue (2003: 121) as "*Paragonyleptes*", the quotation marks indicating that this species was inertially assigned to an invalid genus. *Paragonyleptes* was synonymized with *Collonychium* Bertkau, 1880 by Kury (2003a: 122) and ultimately *Collonychium* ended up as a synonym of *Gonyleptes* Kirby, 1819 by Pinto-da-Rocha et al. (2012: 41). We concur with all those synonymies, except the fact that we deem *M. asperula* to be a female of

*Acanthogonyleptes* as evidenced mainly by the developed armature on free tergites I–III, coxa IV armed with retroapical and proapical acuminate tubercles, straight femur IV armed with dorsal and retrolateral row of acuminate apophyses, as exemplified here by *Acanthogonyleptes fulvigranulatus* (Mello-Leitão, 1922), (Fig 10d).

### *Acanthogonyleptes dubius* (Mello-Leitão, 1932) **comb. nov.**

- *Paragonyleptes dubius* Mello-Leitão 1932b: 322, fig. 191.

**Comment.** *Paragonyleptes dubius* was described by Mello-Leitão based on a male from Mato Grosso [doubtful locality] and provided a photograph of the holotype (Fig. 10c). Despite not confirming the locality, this species possess morphological traits that match the other members of *Acanthogonyleptes*, as transversal proapical apophysis on coxa IV, armature on free tergites and dorsal scutum with gamma pyriform shape (γP). Therefore, we transfer *Paragonyleptoides dubius* to *Acanthogonyleptes* creating the new combination *Acanthogonyleptes dubius* comb. nov.

### *Acanthogonyleptes pygoplus* (Roewer, 1917)

- *Paragonyleptes pygoplus* Roewer 1917: 137, fig. 33.

**Comment.** Roewer described this species based on a female from Santos, SP and provided a schematic drawing of the habitus in dorsal view. As already explained, *Paragonyleptes* ultimately entered in the synonym of *Gonyleptes*, but this species remained in a limbo. Based on the description and the drawing provided by Roewer it is possible to highlight some attributes that indicate this species as belonging to *Acanthogonyleptes*, as (1) developed armature on free tergites I–III, (2) coxa IV armed with retroapical and proapical acuminate tubercles, (3) femur IV armed with dorsal and retrolateral row of acuminate apophyses (Fig. 10d).

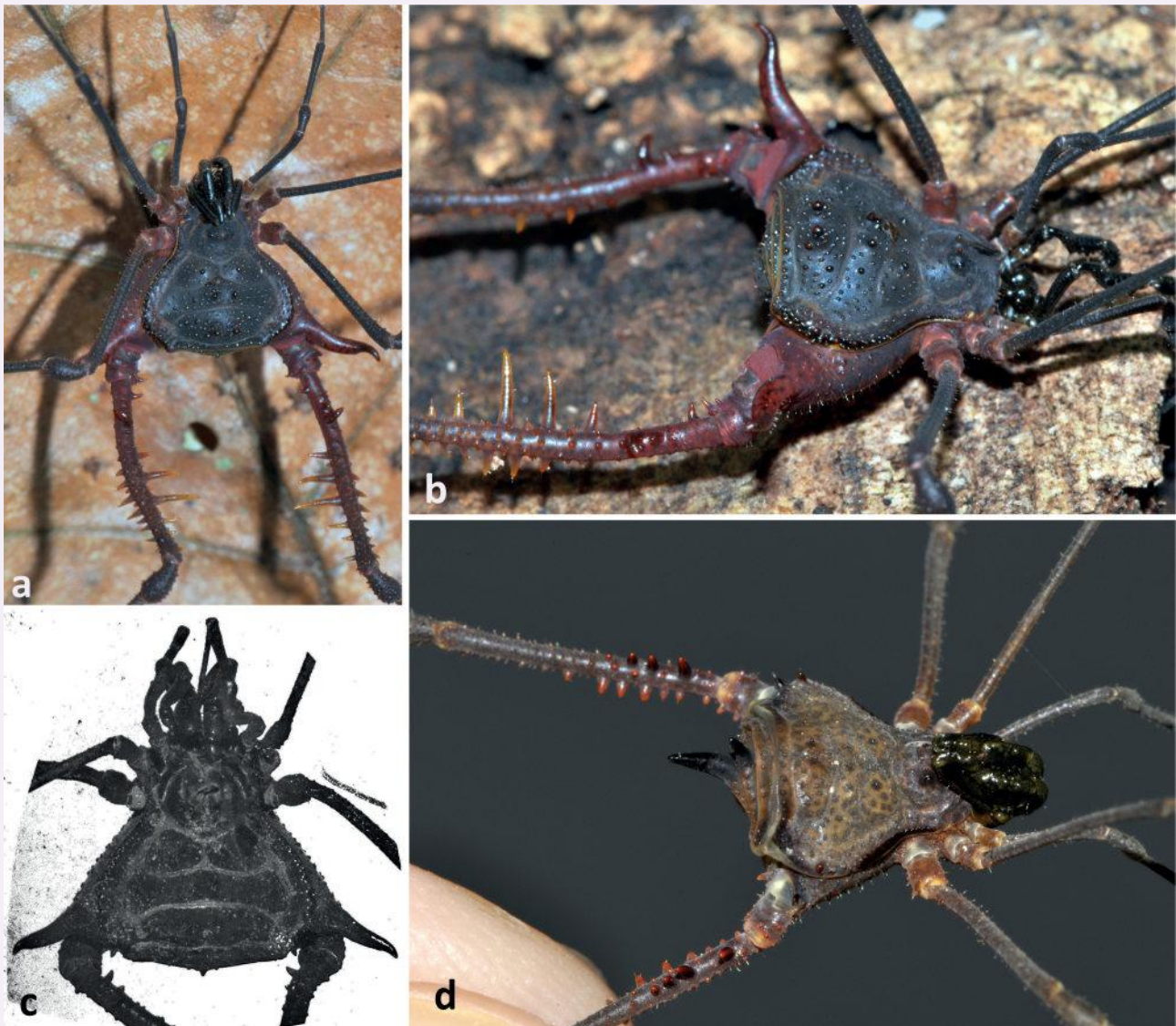
### *Acanthogonyleptes wygodzinskyi* (Soares & Soares, 1954) **comb. nov.**

- *Metagonyleptes wygodzinskyi* Soares & Soares 1954a: 494, fig. 5.

**Comment.** The new combination proposed above is supported due to some features shared with the members of *Acanthogonyleptes*, such as the presence of coxa IV with transversal proapical apophysis, proventral armature of femur IV with row of medium apophyses and the retrolateral sinusoid row of short to long apophyses in median to apical portion of femur IV (Figs 10a, b).

### *Gonyleptoides* Roewer, 1913

- *Gonyleptoides* Roewer 1913: 253. 🐞 Type-species by monotypy: *Ancistrotus acanthoscelis* Bertkau, 1880.



**Figure 10.** Exemplars of species of the clade K92 discussed in this chapter: **(a).** *Acanthogonyleptes wygodzinskyi* (Soares & Soares, 1954) *comb. nov.*, male from Teresópolis, RJ, in vivo, dorsal view; **(b).** Same, dorsolateral view; **(c).** *Acanthogonyleptes dubius* (Mello-Leitão, 1932) *comb. nov.*, male holotype (whereabouts unknown, long lost), dorsal view; original photograph used for the publication; **(d).** *Acanthogonyleptes fulvigranulatus* (Mello-Leitão, 1922), female from Brejo da Lapa, MG, in vivo, dorsolateral view. Photographs Copyright ©: A.B. Kury (a, b, d), Arquivos do Museu Nacional (c). All used with permission.

***Gonyleptoides albipunctatus* (Roewer, 1943) *comb. nov.***

- *Mitobates albipunctatus* Roewer 1943: 50, pl. 6, fig. 57.
- *Gonyleptoides marumbiensis* B. Soares 1945d: 196, fig. 1. **Syn. nov.**

**Comment.** Examination of the female holotype of *M. albipunctatus* led to the conclusion that this species was not a Mitobatinae, but at the same time, an alternative placement has not been proposed, and the noncommittal formula “*Mitobates*” was used (Kury 2003a). Now, with closer study of the species of Progonyleptoidellinae widely known as *G. marumbiensis*, we realize it is the same species (Figs 11a, b). However, this species is also misplaced in the heterogeneous genus *Gonyleptoides* (Figs 11d–f). Nevertheless, for the moment we cannot confidently supply a sure alternative taxonomic place-

ment. This species shares special similarity with the type-species of *Heliella* B. Soares, 1945 (Fig. 11c), but it is out of the scope of the present note to try to advance something in this respect.

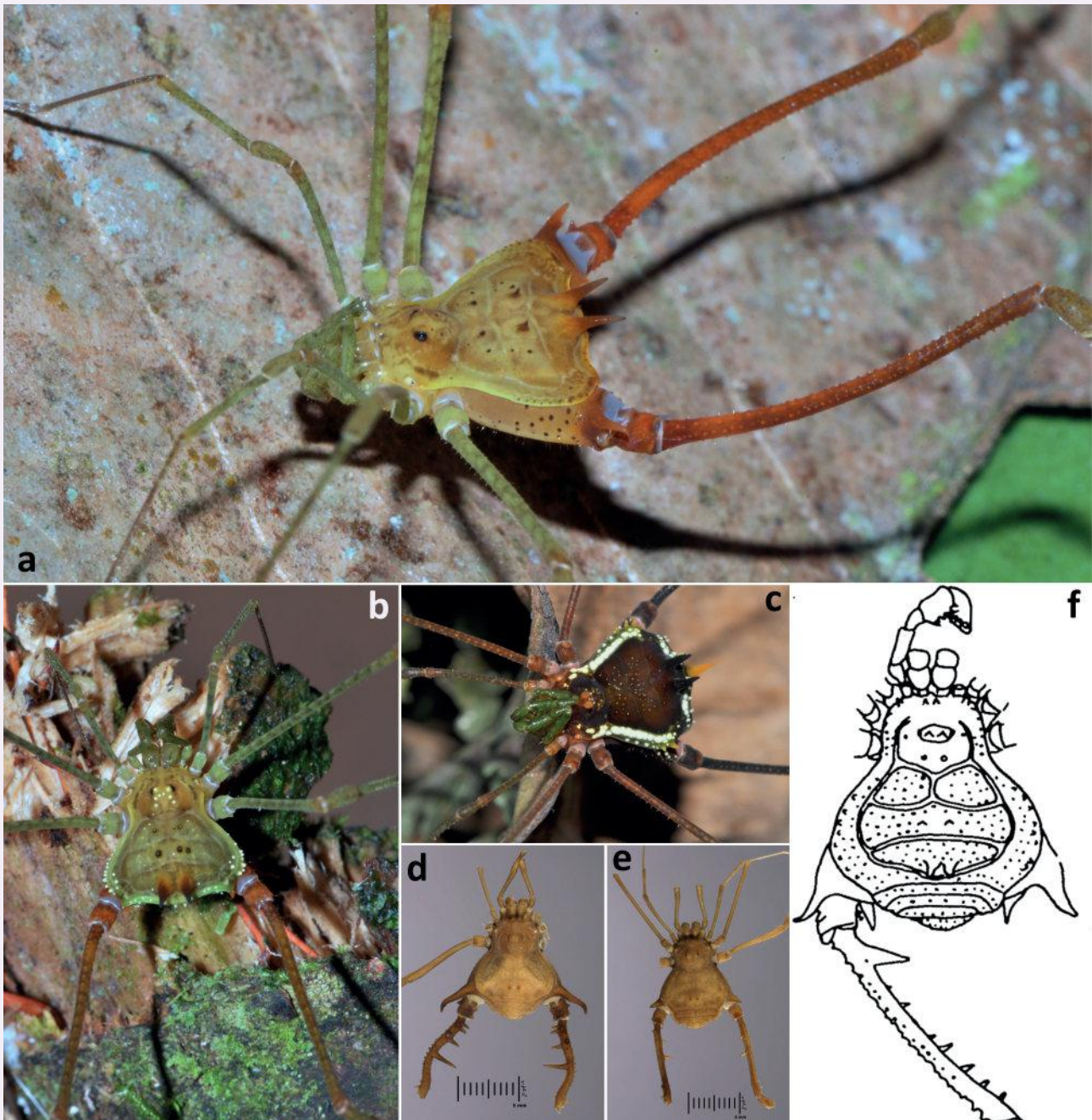
***Gonyleptes* Kirby, 1819**

- *Gonyleptes* Kirby 1819: 450. 🐞 Type-species by subsequent designation of Roewer (1913: 225): *Gonyleptes horridus* Kirby, 1819.

***Gonyleptes auricola* (Mello-Leitão, 1924) *comb. nov.***

- *Paragonyleptes auricola* Mello-Leitão 1924: 183, fig. 4  
“*Paragonyleptes*” *auricola*: Kury 2003a: 121.

**Comment.** Mello-Leitão described this species based on a female from Morro Velho, MG and provided a simplified and unfaithful schematic drawing of the



**Figure 11.** The false *Mitobates* and its alleged relatives: **(a).** *Gonyleptoides albipunctatus* (Roewer 1943) comb. nov., male from Antonina, PR, in vivo; **(b).** Same, female; **(c).** *Heliella singularis* B. Soares, 1945, female from Piraquara, PR, in vivo; **(d).** *Gonyleptoides curvifemur* B. Soares, 1944, male syntype (MZSP 700); **(e).** Same, female; **(f).** *Gonyleptoides acanthoscelis* (Bertkau, 1880), male, from Soares & Soares 1985b: fig. 36. Images Copyright ©: A.B. Kury (a, b, c), Website of Museu de Zoologia da Universidade de São Paulo (d, e), Universidade Estadual Paulista “Júlio de Mesquita Filho”. All used with permission.

habitus in dorsal view. Later, he (1932: 320, fig. 189) provided a photograph of the holotype, of which we reproduce the original here (Fig. 12b), with much better quality than the image that appeared in the publication. As explained above, *Paragonyleptes* ultimately entered in the synonym of *Gonyleptes*, but this species remained in a limbo. In Kury's 2003 catalogue it appeared in “*Paragonyleptes*”. Judging by (1) the massive very light tubercles on laterals of scutum, (2) mesotergal areas I and II each with a slightly arched row of four tiny granules (which are sharply contrasting light colored), and (3) the

conical tubercle on FT II, this is a species closely related to *Gonyleptes perlatus* (Mello-Leitão, 1935) (Fig. 12a).

***Gonyleptes gonypernoides* (Piza, 1943) comb. nov.**

• *Gonyleptilus gonypernoides* Piza 1943a: 48, fig. 6.

*Paragonyleptes gonypernoides*: Piza 1942b: 415.

**Comment.** *Gonyleptilus* Roewer, 1927 was synonymized with *Megapachylus* Roewer, 1913 by Soares & Soares (1987c: 58), with which we concur. However, this species had been earlier transferred to *Paragonyleptes*



Roewer, 1913 by Piza (1942b: 415), which is now a junior synonym of *Gonyleptes* Kirby, 1819. This species shares affinities with the other species of *Gonyleptes* (e.g. *Gonyleptes gonyleptoides*). Therefore, we transfer *G. gonypernoides* to the genus *Gonyleptes* creating the new combination *Gonyleptes gonypernoides* comb. nov.

#### *Juticus* Roewer, 1943

- *Juticus* Roewer 1943: 26. 🐛 Type-species by monotypy: *Juticus furcidens* Roewer, 1943.

**Comment.** We hypothesize that there is a group of species of Gonyleptinae which appear to be closely related to the monotypic *Juticus* as redescribed in Hara et al. (2018). These are exemplified in Figs. 12 c, d. Therefore, we **expand** here the diagnosis of *Juticus* to accommodate all of them. **Diagnosis.** Ocularium armed with a pair of blunt closely placed tubercles. Mesotergum divided into 4 areas. Posterior margin of area IV with V-shaped cleft caused by intrusion of scutal area V. Areas I, II, III, IV each with pair of globular tubercles. Those tubercles on area I and IV are set more widely apart than the others and sometimes are much reduced. Scutal area V [= posterior margin of scutum] sometimes with median acuminate tubercle. Coxa IV of male with a robust proapical apophysis, mostly oblique, rarely transversal (as in *J. hamiferus*), with short secondary branch. Coxa IV of male also with small acuminate retroapical apophysis. Femur IV of male with robust two-branched DO2, with strongly developed retrolateral branch. Main rows of tubercles are: PV4–10, strong spines more or less subequally spaced (pectinate); RL, with only a few more robust units, mainly RL6 to RL9. **Penis:** ventral plate trapezoid with mid-ventral bulge and deep parabolic cleft. Glans with solea only shortly projecting distally; MS A1–A3 very short, sturdy, disposed in arch; MS C1–C3 inserted close together, with tip spatulate. Microsetae of type 1 extending extensively into truncus.

#### *Juticus calcaratus* (Roewer, 1930) comb. nov.

- *Paragonyleptes calcaratus* Roewer 1930: 380, fig. 13.

**Comment.** *Paragonyleptes* Roewer, 1913 was synonymized with *Collonychium* Bertkau, 1880 by Kury (2003a: 122) and in turn, *Collonychium* was synonymized with *Gonyleptes* Kirby, 1819 by Pinto-da-Rocha et al. (2012: 41). We concur with both synonymies. However, this species conforms in general traits with genus *Juticus* Roewer, 1943, as diagnosed above. Therefore, we transfer *P. calcaratus* to the genus *Juticus* creating the new combination *Juticus calcaratus* comb. nov.

#### *Juticus frontalis* (Roewer, 1930) comb. nov.

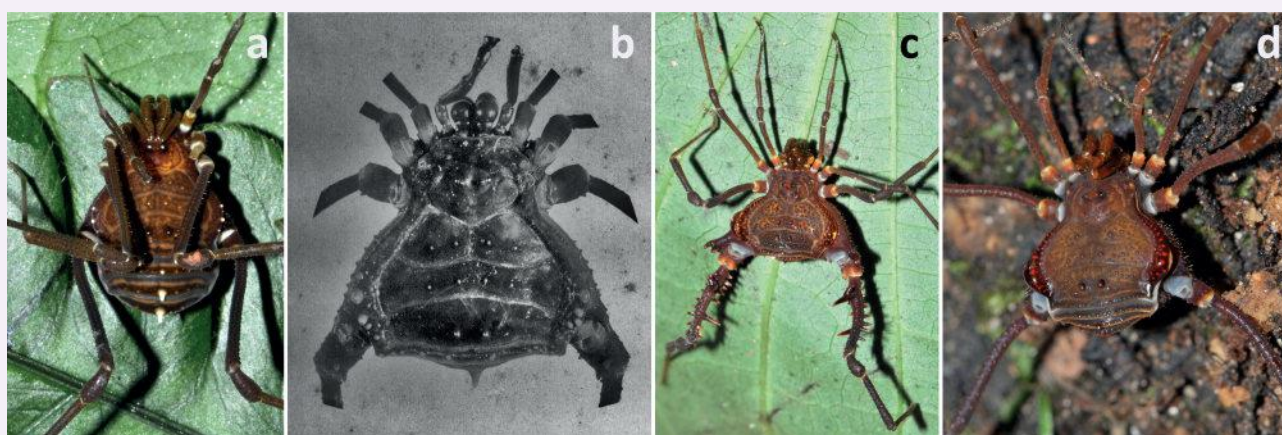
- *Moreira frontalis* Roewer 1930: 434, fig. 39.  
*Paragonyleptes frontalis*: Piza 1942b: 414.

**Comment.** Roewer described *Moreira frontalis* based on two males and one female from Matosinhos, MG and provided a schematic drawing of the habitus in dorsal view. Based on that, it is possible to assign this species to *Juticus* according to the generic diagnosis proposed above. Accordingly, we transfer *Moreira frontalis* to the genus *Juticus* forming the new combination *Juticus frontalis* comb. nov.

#### *Juticus hamiferus* (Mello-Leitão, 1935) comb. nov.

- *Gonyleptilus hamiferus* Mello-Leitão 1935c: 400, fig. 24.

**Comment.** *Gonyleptilus* Roewer, 1927 was synonymized with *Megapachylus* Roewer, 1913 by Soares & Soares (1987c: 58), with which we concur. However, this species conforms in general traits with genus *Juticus* Roewer, 1943, as defined above. Therefore, we transfer *M. hamiferus* to the genus *Juticus* creating the new combination *Juticus hamiferus* comb. nov.



**Figure 12.** *Gonyleptes* and *Juticus*: (a). *Gonyleptes perlatus* (Mello-Leitão, 1935), female from Conceição de Macabu, RJ, in vivo; (b). *Gonyleptes auricola* (Mello-Leitão, 1924) comb. nov., female holotype (MNRJ 1491), dorsal view, original photograph used for the publication; (c). *Juticus* sp., male from Iperó, SP, in vivo, dorsal view; (d). Same, female. Photographs Copyright ©: A.B. Kury (a, c, d), Arquivos do Museu Nacional (b). All used with permission.



# Chapter 7. On a tricommatine from Paraná described in two different families (Laniatores: Gonyleptidae)

by L Caramori & AB Kury

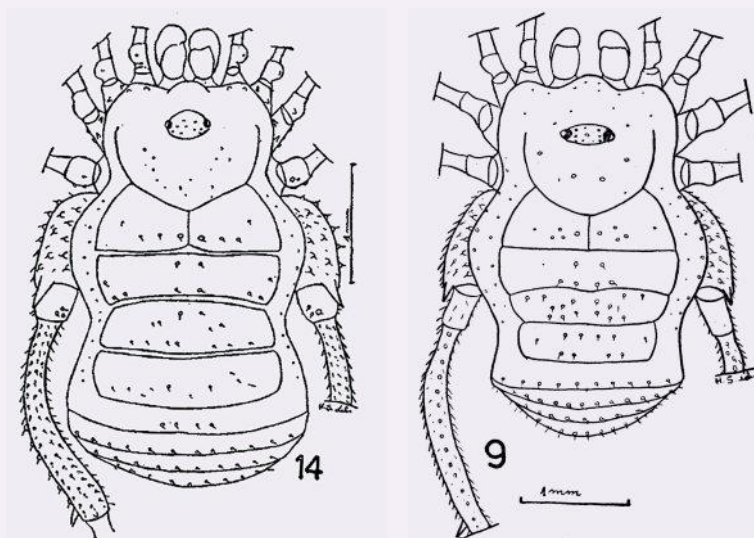
Subfamily Tricommatinae Roewer, 1912

*Tricommatus fulvus* H. Soares, 1966

- *Tricommatus fulvus* H. Soares 1966a: 99, fig. 14.
- *Prosontes brasiliensis* H. Soares 1966a: 96 fig. 9. **II** Junior secondary homonym of *Tricommatus brasiliensis* Roewer, 1912. **Syn. nov.**

**Comment.** *Prosontes* Goodnight & Goodnight, 1945 was described as a monotypic genus from Mexico, which somehow was originally allocated to Gonyleptidae: Pachylinae. Kury & Cokendolpher (2000: 154) transferred it to Cosmetidae. We concur with this transfer, which has never been challenged. The second species of *Prosontes*, *P. brasiliensis*, was described on the basis

of one female from southern Brazil (which is an area strongly dissonant with Mexico, not having a single genus of harvestmen in common). Kury (2003: 153) already dismissed *P. brasiliensis* allocation in *Prosontes* or even Cosmetidae, placing it in Pachylinae, with “*Prosontes*”, between quotation marks. Most probably, the fact that *T. fulvus* and *P. brasiliensis* were originally described in different families strayed subsequent authors from noticing their obvious identity. Both the general morphology and the locality of *P. brasiliensis* are consistent with genus *Tricommatus* Roewer, 1912, and specially with *Tricommatus fulvus* (Fig. 13). The holotypes of both species were collected in May 1948 by Carlos Gofferjé in the same locality: “Serrinha”, PR [= abandoned Railway Station in Balsas Novas Municipality; -25.597263°, -49.712807°]. It seems that the misinterpretation about the presence of a pseudonychium led to confusion as to their taxonomic allocation, given that the original descriptions of both species are extremely similar. The small differences consist of the number of tarsal segments [4-5/6-5-5 in *P. brasiliensis* and 4-5-5-5 in *T. fulvus*] and pedipalpus spines [3-4 in the tibia and 3-3 in the tarsus of *P. brasiliensis* and 4-4 in the tibia and 2-2 in the tarsus of *T. fulvus*], besides an anteriorly elevated cephalothorax in *T. fulvus*, mentioned by H. Soares. Therefore, the presence of pseudonychium would be the only significant difference between the species, mentioned by H. Soares as a characteristic that “pushes” the specimen to be placed in Gonyleptidae. Helia Soares (pers. comm. to ABK, 1988) considered any setiferous tubercle located between the insertions of the claws as a pseudonychium, and this way, the interpretation of presence vs absence of this structure could be very flimsy. Herewith, we propose that *Prosontes brasiliensis* is a junior synonym of *Tricommatus fulvus* **syn. nov.**



**Figure 13.** Gonyleptidae: Tricommatinae, *Tricommatus fulvus* H. Soares, 1966, female holotypes of the two nominal species synonymized herein, habitus, dorsal view: (a) *Tricommatus fulvus* H. Soares, 1966; (b) *Prosontes brasiliensis* H. Soares, 1966. **(3a).** From H. Soares (1966a). © Museu de Zoologia da Universidade de São Paulo. Used here with permission. **(3b).** From H. Soares (1966a). © Museu de Zoologia da Universidade de São Paulo. Used here with permission.

# Chapter 8. On the allocation of some Palaeozoic and Tertiary harvestmen

by AB Kury, JA Dunlop & AM Mendes

## Suborder Dyspnoi Hansen & Sørensen, 1904

### † Nemastomoididae Petrunkevitch, 1955

- Nemastomoididae Petrunkevitch 1955: 85. 📖 Type genus: *Nemastomoides* Thévenin, 1901 [stem: *Nemastomoid-*].

### † *Nemastomoides* Thévenin, 1901

- *Nemastomoides* Thévenin 1901: 609. 📖 Type-species by monotypy: *Nemastomoides elaveris* Thévenin, 1901.

### † *Nemastomoides elaveris* Thévenin, 1901

- *Nemastomoides Elaveris* Thévenin 1901: 609, pl. XIII, fig. 2.

**Comment.** The genus *Nemastomoides* was not originally included in any family (Thévenin 1901), but said by its author to be closest to the modern genus *Nemastoma* C.L. Koch, 1836, which is a Dyspnoi. Petrunkevitch (1955) created the new family Nemastomoididae for *Nemastomoides* in the superfamily Troguloidea (Dyspnoi). Dunlop (2007: 255) considered them to be stem Phalangida but also stated “in my opinion the two valid *Nemastomoides* species resemble Eupnoi more than Dyspnoi”.

Now we have reappraised the issue of the subordinal appartenance of Nemastomoididae. The rationale for keeping Nemastomoididae more firmly in Dyspnoi is as follows:

1) The name *Nemastomoides* Thévenin 1901 (originally monotypic) is anchored in the type species *Nemastomoides elaveris* Thévenin, 1901 from the Coal Measures of Commeny, northern France, upper Carboniferous.

2) Thévenin’s original illustration is poor and there are no features that could be retrieved which are useful to elucidate the allocation of *Nemastomoides*. There is no modern taxonomic work on this genus that could

help understand its relationships, nor any recent study of the type material under advanced techniques.

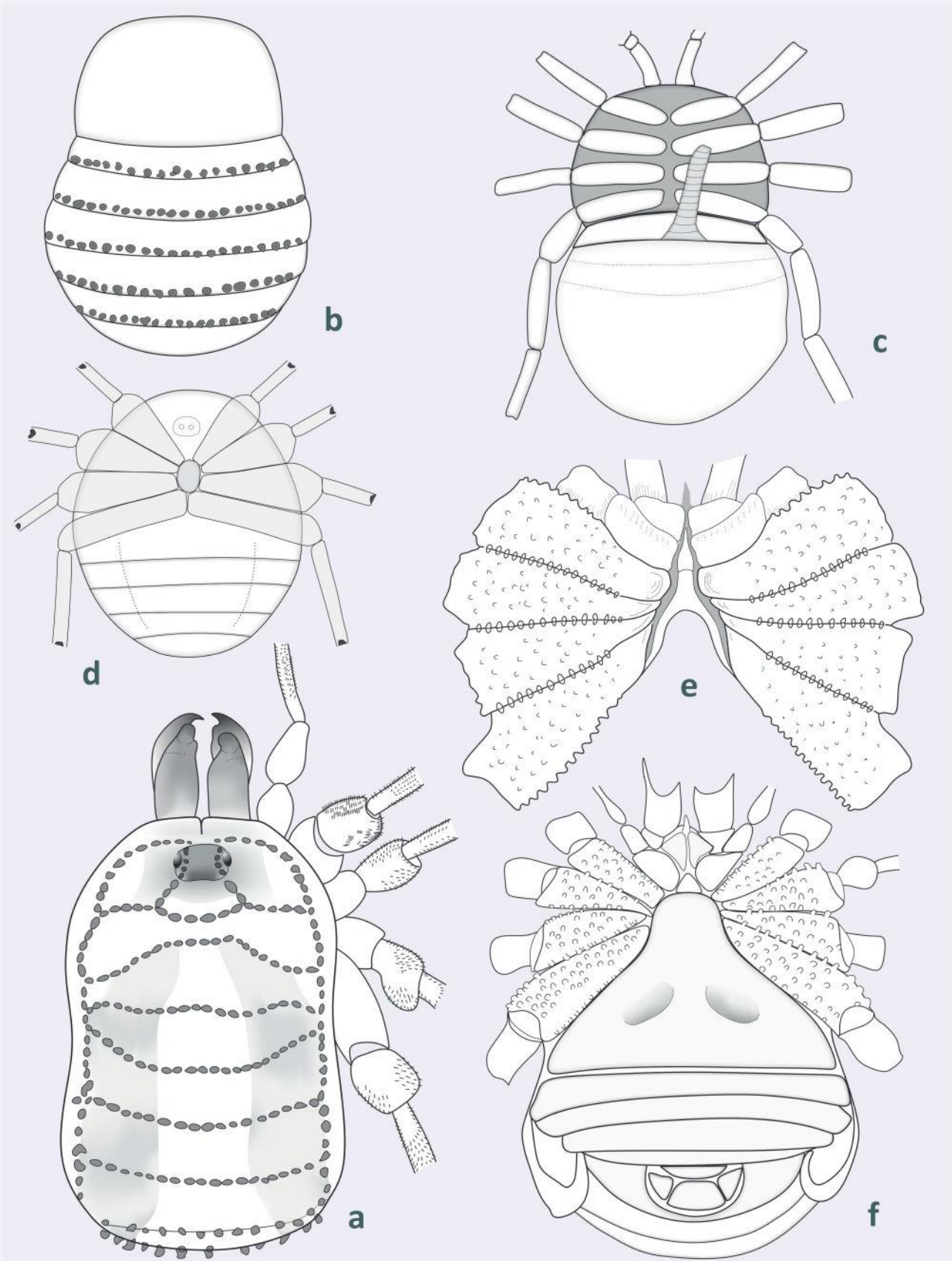
3) Frič (1904: 29) explained that he spent but a short time in Paris and could not find a microscope with which to study Thévenin’s type material. Nevertheless he provided three illustrations of it, made with the aid of a hand-lens (his figs 34 A to C).

4) In Frič’s fig. 34B, in dorsal view *N. elaveris* appears to have a multi-segmented abdominal scutum (Fig. 14b). However, this is most probably an effect caused by the possession of multiple granular rows following the original segment lines, such as present in modern-day nemastomatid *Mitostoma*. The general shape of the scutum is also reminiscent of *Mitostoma* (Fig. 14a).

5) *N. elaveris* appears to have a long segmented ovipositor (Fig. 14c), consistent with Eupnoi (Frič 1904: 29, fig 34C). In Petrunkevitch (1955: 84, fig. 52-1b) there is another interpretation of the ventral surface of *N. elaveris*, in which the “ovipositor” disappears (Fig. 14d). Of course Thévenin’s material should be re-examined with modern technology to definitely discard the presence of such an eupnoan ovipositor.

6) Both Pocock’s (1902) and Kästner’s (1928) illustration of the ventral prosoma of *Nemastoma* sp. (Figs 14e, f) are fairly similar to Petrunkevitch’s rendering. However, what looks like a small oval sternum in Peterunkevitch’s drawing must be considered unlikely as this should not be part of the typical harvestman body plan.

7) In view of the evidence presented, we conclude here that *Nemastomoides* is a Dyspnoi, closely related to Nemastomatinae. We only keep for now the family Nemastomoididae for stability, while modern studies are not able to further clarify their affinities.



**Figure 14.** *Nemastomoides elaveris* Thévenin, 1901 compared with modern Nemastomatinae: **(a)**, *Mitostoma chrysomelas* (Hermann, 1804), habitus, dorsal view, from Spöck (1963); **(b)**, dorsal scutum, dorsal view, from Frič (1904); **(c)**, Habitus, ventral view, with artifactual ovipositor, from Frič (1904); **(d)**, same, without added ovipositor, from Petrunkevitch (1953); **(e)**, *Nemastoma* sp., coxae and sternal region, genital operculum removed, ventral view, from Pocock (1902b); **(f)**, *Paranemastoma quadri-punctatum* (Perty, 1833), habitus, ventral view, from Kästner (1928). All redrawn/adapted from the original sources.

## Suborder Eupnoi Hansen & Sørensen, 1904

### † Kustarachnidae Petrunkevitch, 1949

#### † *Kustarachne* Scudder, 1890

- *Kustarachne* Scudder 1890: 450. 🐛 Type-species by monotypy: *Kustarachne tenuipes* Scudder, 1890.
- *Protopilio* Petrunkevitch 1913: 110. 🐛 Type-species by original designation: *Protopilio longipes* Petrunkevitch, 1913. II Junior subjective synonym of *Nemastomoides* Thévenin, 1901 by Petrunkevitch (1953); **synonymy rejected herein. Syn. nov.**

#### † *Kustarachne longipes* (Petrunkevitch, 1913) **comb. nov.**

- *Protopilio longipes* Petrunkevitch 1913: 110, pl. X, fig. 57; text fig. 68.  
*Nemastomoides longipes*: Petrunkevitch 1953: 46.

**Type data.** Holotype (PMY 171), from USA, Illinois, Mazon Creek.

**Horizon.** Upper Carboniferous, Pennsylvanian, Lower Allegheny.

**Comment.** *Kustarachne* was originally included in Anthracomarti: Architarboidea. It was transferred to its own order Kustarachnae (Petrunkevitch 1913), later renamed Kustarachnida (Petrunkevitch 1955) to conform with the standardized endings adopted by him. Then it was transferred to Eupnoi: Phalangoidea by Dunlop (2004a: 24), who redescribed the type-species, *Kustarachne tenuipes*.

*Protopilio longipes* is currently classified in *Nemastomoides* (Petrunkevitch 1953) as a synonym of *K. album* (Smith, 1850) (see Miller 2005). While the typical *Nemastomoides* from France fits well in Dyspnoi (as explained above), one of the American species – *Nemastomoides longipes* – does not share a single special similarity with it (Fig. 15). On the other hand, it strongly resembles *Kustarachne* in all aspects (after the fictitious characters reported by Petrunkevitch were

disclaimed), therefore it is here brought into this genus. Accordingly, *Protopilio* should be considered as a junior synonym of *Kustarachne*.

### Suborder † Tetrophthalmi Garwood, Sharma, Dunlop & Giribet, 2014

#### Family † Hastocularidae **fam. nov.**

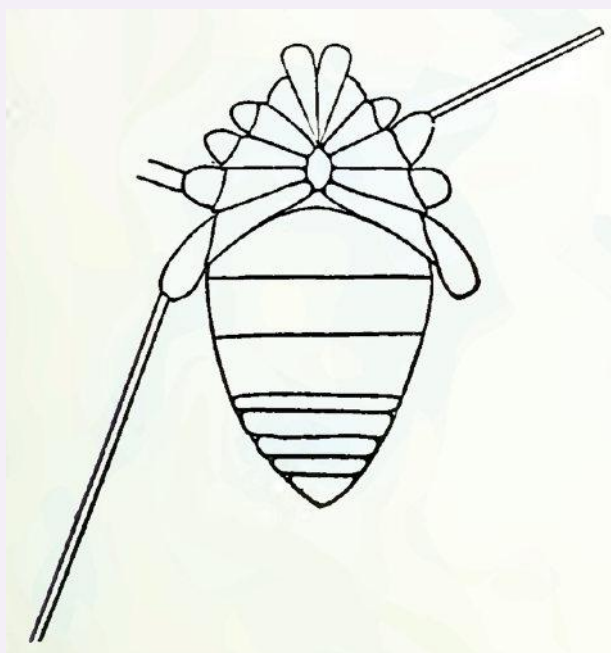
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**Comment.** This family is so far coterminous with the infraorder † Tetrophthalmi, and it is being described here to accomplish the full complement of traditional categories, although it is not mandatory. † Tetrophthalmi has been recently created to include two genera of Devonian and Carboniferous harvestmen (Garwood et al. 2014).

**Included genera.** † Hastocularidae is then composed by † *Eophalangium* Dunlop, Anderson, Kerp & Hass, 2004 from the Devonian of Scotland and † *Hastocularis* Garwood, Sharma, Dunlop & Giribet, 2014 (**type-genus**) from the Carboniferous of France.

**Diagnosis.** Carapace with median ocellum projected as a hook and bearing a pair of eyes; a pair of lateral prosomal mounds also bearing eyes. Dorsal scutum strongly convex in the middle. Tergites configuring a scutum completum. Legs elongate, without extreme incrassation or modifications. Genital operculum absent. Intromittent penis composed of a short shaft and a distal multilobulate region (in † *Eophalangium* the shaft is longer and there is a sclerosomatine-like glans).

**Figure 15.** † *Kustarachne longipes* (Petrunkevitch, 1913) **comb. nov.**, Peabody Mus. No. 171, ventral surface, from Petrunkevitch (1913). Image Copyright © The Connecticut Academy of Arts and Sciences, Yale University. Used here with permission.





# Chapter 9. A new family from the New World (Eupnoi: Phalangioida)

by AB Kury & JC Cokendolpher

## Globipedidae Kury & Cokendolpher fam. nov.

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Phalangiidae [unnamed group]: Gruber 1970: 273 (incl. *Diguëtinus*, *Eurybunus*, *Globipes*, *Metopilio*).

Phalangioida: undescribed family or subfamily Metopilio assemblage: Cokendolpher 1984: 27 (newly incl., *Dalquestia*); Cokendolpher & Cokendolpher 1984: 167.

Sclerosomatidae: Metopilio genus group: Crawford 1992: 4.

Sclerosomatidae: Subfamily unknown (= Metopilio group): Cokendolpher & Lee 1993: 16.

Sclerosomatidae: unnamed assemblage: Cokendolpher & Sissom 2000: 243.

Metopilio group: Giribet et al. 2010: 13; Hedin et al. 2012: 224; Cokendolpher & Holmberg 2018: 6.

**Comment.** This assemblage of Eupnoi has been known for decades as “Metopilio group” and similar monikers, but remained hitherto unnamed. It is important for nomenclature to formally recognize this taxon. We have chosen as type instead of *Metopilio*, the genus *Globipes* because we think the family name is more euphonic. It should be noted that as *Globipes* is derived from Latin *globus* (globe) + *pes*, *pedis* (foot), the stem formed with the singular genitive is *Globiped-*.

They are distributed in southwestern USA, Mexico, and Central America (Chiriquí, Panama north). Examples of living specimens of several genera are shown here in Fig. 16.

**Type genus.** *Globipes* Banks, 1893.

**Included genera.** *Dalquestia* Cokendolpher, 1984, *Diguëtinus* Roewer, 1912, *Eurybunus* Banks, 1893, *Globipes* Banks, 1893, *Lanthanopilio* Cokendolpher & Cokendolpher, 1984 and *Metopilio* Roewer, 1911.

**Diagnosis.** Thick, leathery, granulated cuticle; smoother in *Eurybunus*. **Preocular** area medially often with a small (up to 1/2 height of ocularium) **hump** generally covered with smaller denticles; without trident or

conspicuous large spines/tubercles. **Ozopores** situated near the bases of legs I, clearly visible from above, not occluded by spines or other ornamentation. **Ocularium** low and narrow, essentially round in dorsal view; smooth to denticulate. Strongly arched abdomen (less so in males); tergites 6–7 can be fused with the scutum into a uniform shield. **Tergites** form a scutum parvum. No lateral abdominal sclerites. Tergites mostly with single transverse row of strong acuminate tubercles; modified to two or numerous smaller denticles per row in some *Dalquestia* and *Diguëtinus* and much reduced to absent in *Globipes* and *Eurybunus*, especially in females. **Carapace** small compared to engorged abdominal scutum, especially in females. Abdominal **spiracles** (= **stigmata**) of second abdominal somite bearing occluding **entapophysis** (Šilhavý 1970, fig. 11) without a grill or lattice. Lateral abdominal sclerites absent. Anal operculum with white spot on some species. **Chelicerae** generally monomorphic, weakly developed in both sexes (slightly enlarged and tuberculate in few *Dalquestia*). Basichelicerite with variously developed ventral spur (ranging from distinctly spurred/lobed to rounded bump to smooth). **Pedipalps** monomorphic except palpal tarsus ventromedially bears a row (or a belt) of denticles in males, but not females. Not significantly inflated, clamp-type; no apophysis of any sort on any segment from any age group. Pedipalpal claw smooth, rarely with one to few small rounded processes, not pectinate (= toothed). **Legs** not especially elongate, sexually dimorphic, legs I and III in males very stout and in *Diguëtinus* heavily armed, basically cylindrical but often femur and tibia with polygonal cross-section (especially in *Dalquestia*), with rows of spines or hairs on the edges. Tibia with two accessory spiracles, one near the proximal and one near the apical end. No pseudoarticular nodules on any femora, and with distinct pseudosegments in tibia II (except very rarely in *Dalquestia*). Leg coxae without marginal rows of denticles, coxae II without distally blunt lobes. **Penis**: most diagnostic structures variable and previously hand drawn (Fig. 15, *Globipes* and *Diguëtinus*): *Metopilio* by Gruber (1969); *Lanthanopilio* by Cokendolpher and Cokendolpher (1984); *Dalquestia* Cokendolpher (1984), Cokendolpher and Stockwell (1986), Cokendolpher and Sissom (2000), and *Metopilio* photographed with SEM by Rodriguez, Townsend, Jr. & Proud (2014). Truncus: tubular and sclerotized with a relatively long shaft; slender and lanceolate throughout most of length; or basally wide and bulbed (*Dalquestia*) and shortly afterward tapered to a thinner uniform width. Tendon long, inserted ventrally, at base of glans. Muscle fills about basal half-length of truncus. Distal region (middle region in *Lanthanopilio*) bears lateral transparent plates or alae that are attached to the truncus, large and ventrally curved in *Diguëtinus* and *Metopilio*, reduced in *Dalquestia*, *Globipes* and *Eurybunus*. Alate portion different in *Lanthanopilio* and placed at about mid-length of shaft (see Cokendolpher and Cokendolpher 1984, figs. 3–7). Glans: attenuate spin-



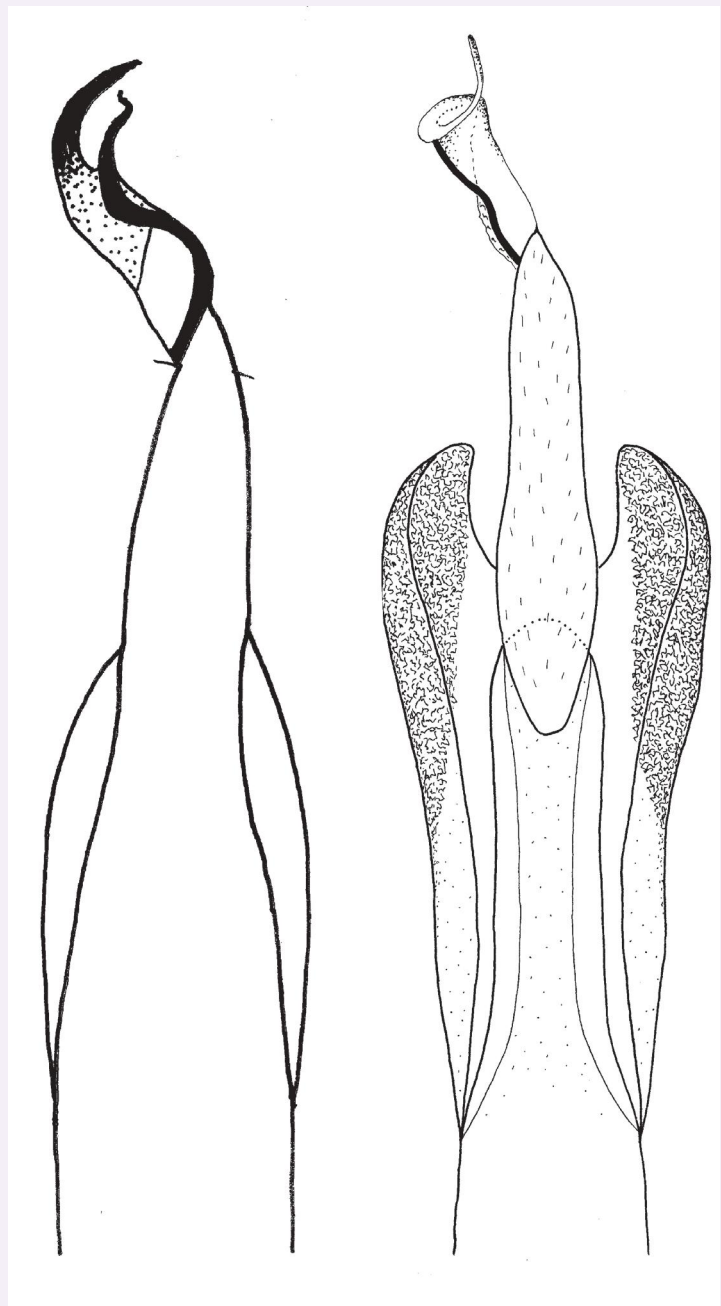
**Figure 16.** Representatives of diverse species of **Globipedidae fam. nov. in vivo.** (a). *Diguetinus raptator* Roewer, 1912, male, from Michoacán, Mexico; (b). *Eurybunus* sp., female from Arizona, USA; (c). *Eurybunus* sp. from Baja California, Mexico. The red pigment near trochanters IV are small parasitic mites; (d). *Metopilio* sp., female from Jalisco, Mexico; (e). *Globipes* sp., female from California, USA. F. *Dalquestia formosa* (Banks, 1910), male, from Texas, USA. Photographs Copyright ©: Ricardo Arredondo T. (a), Marshal Hedin (b), Christian Schwarz (c), Ricardo Olivo R. (d), Cedric Lee (e), Robby Deans (f). All used with permission.



dle-shaped, dorsobasally connected to truncus, rigid with and continuing truncus in a straight line (*Lanthanopilio* apparently flexible and held half-folded over dorsally), weakly delimited ventrobasally by half-ring wrinkle. A reduced, "bulbous" sac is present in the winglets. With two pairs of lateral sensory bristles in the distal 1/3; these bristles often easily observable with high magnification, but can be small or appear to be absent. Penis of *Lanthanopilio* has a significantly different structure with sclerotized "horns" on the glans of *Lanthanopilio* truncus and glans distally with numerous small setae, some *Metopilio* and *Lanthanopilio* have noticeably longer setae (in *Lanthanopilio* also covered with long trichoid "setae" (these structures appear to be outgrowths of the truncus, they do not appear to have sockets at their

bases) and alate portion membranous, not highly sclerotized. Stylus: irregularly spindled and thickened at base, apical third as a thin, sinuous tube (*Dalquestia*, *Globipes*, *Eurybunus*) to coiled (*Diguetinus*, *Metopilio*), inserted dorso-apically to glans, movable, dorsally rugose (or wrinkled) appearance.

**Figure 17.** Left. Distal end of penis, dorsal, *Globipes spinulatus* Banks, 1893 from California, USA. Right. Distal end of penis, ventral, *Diguetinus raptator* Roewer, 1912 from Jalisco, Mexico.





# Chapter 10. Expansion of the MECO clade (Grassatores: Microsetata)

by AB Kury & R Carvalho

## Carunculata **Kury new unranked taxon**

**Comment.** This is an expansion of the MECO clade (Cosmetidae + Metasarcidae) of Gonyleptoidea, to also include Prostygnaeidae (promoted to family in Villarreal & Kury 2020) and the new family Askawachidae.

**Diagnosis.** Podium in situ ca. 1/3 length of VP. Lateral fields of microsetae in VP formed by type-4 scale-bristles prominent, occupying most of ventral plate. Macrosetae D mostly lateral, between A and C. Stylus mostly C-shaped curved, with or without dorsal and ventral processes. Distal portion of stylus strongly compressed, bearing a wattle (or carunculus; ventral flap with serrate margin) and sometimes with lateral rows of barbs. Ejaculatory opening slit-like, shifted to dorsal.

**Cladistic node-based definition.** The last common ancestor of *Oxapampeus* Roewer, 1963 (oldest genus of Askawachidae) and *Cosmetus* Perty 1833 (oldest genus of Cosmetidae), and all descendants of that ancestor.

**Included subtaxa.** Askawachidae, Prostygnaeidae (ranked herein as family following Villarreal & Kury 2020), Metasarcidae and Cosmetidae.

## Askawachidae **Kury & Carvalho fam. nov.**

urn:lsid:zoobank.org:act:760B85CE-B8F1-407F-8337-60474F-2DE298

**Comment.** This is a hitherto undescribed clade of Gonyleptoidea Carunculata from Andean-Amazonian areas. A full study of this taxon was in preparation when the 2018 fire in MNRJ destroyed all the relevant material. However, with the preliminary data obtained thus far, it is possible to clearly characterize this as a new family.

**Type genus.** *Askawachi* Kury & Carvalho gen. nov.

**Included genera.** *Askawachi* Kury & Carvalho gen. nov., *Oxapampeus* Roewer, 1963 transl. nov. (Fig. 18b; originally in Gonyleptinae; transferred to Pachylinae by Pinto-da-Rocha et al. (2012)) and *Thermodontia* Kury & Carvalho gen. nov.

**Diagnosis.** Dorsal scutum outline gamma-type. Sexual dimorphism well-marked, manifested in legs III (Fe with apical spur) and IV (Cx to Ti with apophyses and spines), well-armed in males. Cx IV in males strongly





developed, widely surpassing lateral margin of DS, but not reaching its posterior border. Common ocularium clearly marked but small, armed with pair of tubercles of small spines. Pedipalps short and slender, with articles cylindrical. Macrosetae of penis VP either in the configuration of 3A/0b/3C/1d/2e or more often with MS A and C robust and extremely multiplied in number, occupying all lateral extension of VP. Wattle with well-developed lateral rows of barbs. Dorsal and ventral processes of stylus highly variable.

**Askawachi Kury & Carvalho gen. nov.**

urn:lsid:zoobank.org:act:DF1DDF75-F8AE-4E11-B-988-687889EF8A5D

**Comment.** *Punrunata* seems to be a *bona fide* gonyleptid. However, *P. pulchra* is not closely related to it and thus it is here combined under this new genus.

**Type-species.** *Punrunata pulchra* Soares & Soares, 1979.

**Etymology.** From Cuscan Quechua *aska* (many) + *wachi* (arrow; spear; sting; prick) because of the supernumerary macrosetae on the penis ventral plate. *Askawachi* is singular in Quechua. The plural should be *Askawachikuna*; however, to produce a shorter, more euphonic form, we have omitted the plural suffix. Gender masculine.

**Diagnosis.** Spines of ocularium much reduced. Scutal area I undivided; area III unarmed; area IV present, undivided. Coxa IV, apophysis retroventral distal uniramous. Ventral plate without constriction between proximal and distal parts. Ventral process of glans 1/3 or less of stylus diameter. Numerous supernumerary microsetae A, occupying most of VP length, MS B indistinct (Fig. 18a).

**Askawachi pulcher (Soares & Soares, 1979) comb. nov.**

- *Punrunata pulchra* Soares & Soares 1979a: 397 figs 23–31.

**Figure 18.** Askawachidae fam. nov. (previous page): (a) *Askawachi pulcher* (Soares & Soares, 1979) comb. nov. (MUSM 406), distal half of penis, sinistrolateral view; (b) *Oxapampeus weyrauchi* Roewer, 1963 (FMHN AK 157), distal half of penis, sinistrolateral view; (c) *Thermodontia caramaschii* (Soares & Soares, 1979) comb. nov., distal half of penis, dextrolateral view. Scale bars: 1 mm (d); 100 µm (a, b); 50 µm (c).

**Figure 19.** Askawachidae fam. nov.: *Thermodontia caramaschii* (Soares & Soares, 1979) comb. nov., male (MPEG 2443) habitus, dorsal view. Scale bar: 1 mm

**Thermodontia Kury & Carvalho gen. nov.**

urn:lsid:zoobank.org:act:689B6A5F-204D-4BF3-92F-4-26A530618218

**Comment.** The type species of *Paraluederwaldtia* Mello-Leitão, 1927 is a true gonyleptid from the highlands of Mata Atlântica, entirely unrelated with *Thermodontia*.

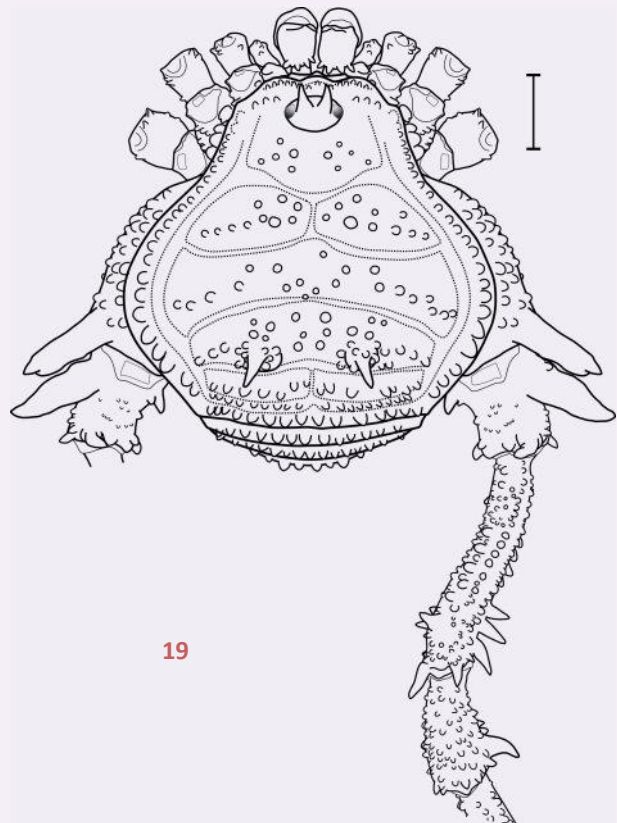
**Type-species.** *Paraluederwaldtia caramaschii* Soares & Soares, 1979.

**Etymology.** From Greek Θερμώδων (Terme River, in Asia Minor), at which banks Herodotus and Strabo place the fierce warrior women Amazons. This refers to the type locality of the type-species in the Amazon drainage. Gender feminine.

**Diagnosis.** Spines of ocularium higher than ocularium height. Scutal area I divided into left and right halves; area III armed with a pair of spines; area IV present, divided medially. Coxa IV without apophysis retroventral distal. Ventral plate with constriction between proximal and distal parts. Ventral process of glans absent. Only two microsetae A; MS B clearly distinct (Figs. 18c, 19)

**Thermodontia caramaschii (Soares & Soares, 1979) comb. nov.**

- *Paraluederwaldtia caramaschii* Soares & Soares 1979c: 258, figs 7–14.



# Chapter 11. On two false South American *Liogyndulus*

by AB Kury & R Pinto-da-Rocha

Family *Gonyleptidae* Sundevall, 1833

*Acrographinotus opacus* (Roewer, 1952) **comb. nov.**

- *Liogyndulus opacus* Roewer 1952: 54.

**Type data.** ♀ holotype (SMF RII 10126/122); 1 ♀ 1 juv. paratypes (WWPC), PERU, Pasco, Laguna Punrún, 4400 m near Cerro do Pasco, in the water system of Junin Lake.

**Comment.** *Liogyndulus* Mello-Leitão 1932 was synonymized with *Cobania* Roewer, 1913 by Soares & Soares (1984: 309). Its type-species, *Liogyndulus luteifemur* Mello-Leitão, 1932 is a synonym of *Cobania picea* (Bertkau, 1880), from the mountains of SE Brazil. We concur with both synonymies. This species, however, is distributed in the water system of Junin Lake, between Peru and Bolivia, which, based on biogeographic features, would make it highly unlikely that it belongs to *Cobania* sensu stricto. Furthermore, *Liogyndulus*

*opacus* (Fig. 20a) matches perfectly the current diagnosis of *Acrographinotus* Holmgren, 1917 (Acosta 2001: 58). Therefore, we herewith transfer *L. opacus* to the genus *Acrographinotus* creating the new combination *Acrographinotus opacus* **comb. nov.**

*Parapachyloides bispinifrons* (Roewer, 1943) **comb. nov.**

- *Liogyndulus bispinifrons* Roewer 1943: 20, pl. 1, fig. 8.

**Type data.** ♀ holotype (SMF RII 5330), BRAZIL, Santa Catarina, Itapocu.

**Comment.** *Liogyndulus* Mello-Leitão 1932 was synonymized with *Cobania* Roewer, 1913 by Soares & Soares (1984: 309). Its type-species, *Liogyndulus luteifemur* Mello-Leitão, 1932 is a synonym of *Cobania picea* (Bertkau, 1880), from the mountains of SE Brazil. We concur with both synonymies. This species, however, hails from southern Brazil. It is known only from a female, but it is clearly not a female of *Cobania* sensu stricto. General morphology, especially the alpha-key-hole scutum outline (Kury & Medrano 2016), praeocular mound and huge size (Fig. 20b) are consistent with *Parapachyloides* Roewer, 1913.

The occurrence of a member of *Parapachyloides* in the coastal mountain range of Santa Catarina is unexpected and does not match the known distribution of the genus. This species has never been recorded again, and it remains to be determined if this is a mislabeling.

In spite of that, we herein transfer *C. bispinifrons* to the genus *Parapachyloides* creating the new combination *Parapachyloides bispinifrons* **comb. nov.**

**Figure 20.** False *Liogyndulus*: (a) *Acrographinotus opacus* (Roewer, 1952) **comb. nov.**, ♀ paratype + label (AMNH) from Peru, habitus, dorsal view; (b) *Parapachyloides bispinifrons* (Roewer, 1943) **comb. nov.**, ♀ holotype + label (SMF RII 5330), from Brazil. Images copyright © Ricardo Pinto-da-Rocha.





# Chapter 12. Pachyli- nae at their sou- thernmost extreme (Laniatores: Gonylep- tidae)

by M Pessoa-Silva, MR Hara, R  
Pinto-da-Rocha & AB Kury

## *Eubalta planiceps* (Guérin-Ménéville, 1842) **comb. nov.**

- *Gonyleptes planiceps* Guérin-Ménéville in Gervais 1842: 2, pl. 2.  
*Sadocus planiceps*: Kury 2003: 191
- *Balta meridionalis* Sørensen, 1902: 22. **Syn. nov.**  
*Eubalta meridionalis*: Roewer 1923: 410, 445 fig. 504.
- *Gonyleptes modestus* Gervais 1847: 576. **Syn. nov.**  
*Discocyrtus modestus*: Roewer 1913: 120.

**Type data.** *Gonyleptes planiceps*: 1 ♂ 1 ♀ syntypes (“collection de M. Guérin-Ménéville”, present whereabouts NHM, examined, ♀ lost), “Détroit de Magellan”. *Balta meridionalis*: 1 ♂ 3 ♀ syntypes (ZMH, “Coll. Mich. 62”, of which 1 ♂ 1 ♀ subsampled by Roewer as SMF RI 805, all examined) “Patagonia meridionalis, Tuesday bay” F. Delfin leg. Aug. 1892; 1 ♂ (ZMUC, ex-MNHN, donated by E. Simon, labeled as *Gonyleptes planiceps*, examined), “Patagonia meridionalis, ad fretum Magellanicum”. *Gonyleptes modestus*: 1 ♀ holotype (whereabouts unknown), “Chili” without further locality data.

**Comment.** *Gonyleptes planiceps* (Figs 21, 22) from Magallanes, Chile has a quite convoluted taxonomic history, being transferred to many genera and/or re-described many times by Gervais (1844; 1849), Sørensen (1902) and Roewer (1913, 1923). Sørensen (1902) and Roewer (1913) noted differences between the text and the illustration provided by Gervais in the redescrptions. For some reason, both authors compared the additional material of *G. planiceps* identified by Simon with *Balta meridionalis* and they agree that these two are distinct species. Perhaps both authors noted the similarities between both species, and wanted to address that issue.

Currently, *G. planiceps* is placed in *Sadocus*. In an ongoing revision of *Sadocus*, MPS, MRH and RPD (from now on “MMR”) tracked down the whereabouts of *G.*



**Figure 21.** *Eubalta planiceps* (Guérin-Ménéville, 1842), *in vivo*, from Reserva Nacional Magallanes, Punta Arenas, Chile; male. Image copyright © Gonzalo Giribet, used with permission.

*planiceps* type material, originally housed at Guérin-Meneville private collection. MMR found it at NHM (male syntype), and we examined it by detailed photographs (as well as stacker imaging by ABK), including those of the vial label, with the inscriptions “type” and “Guérin”. All those were detailed enough to remove it from *Sadocus*, but we had to look for a genus to house it. Considering the type locality and the past systematic issues, we compared it with *Balta meridionalis*. We also had to track the type material of this species, which we finally found deposited at SMF (SMF 805; male and female syntypes removed from the type series in ZMH), ZMH (2 female syntypes), ZMUC (1 male syntype) – all those syntypes match perfectly in number and sex with the type material of Sørensen (1902). Comparing all type material of *B. meridionalis* with that of *G. planiceps*, we concluded that they belong to the same species. MMR will publish a more detailed historical account of this species elsewhere. ABK, during his thorough research of all harvestmen publications to prepare this catalogue, arrived at the same conclusion as MMR. He kindly invited MMR to author this finding herein.

*Gonyleptes modestus* was poorly described by Gervais (1847), even for the standards of that time (Sørensen 1902; Roewer 1913), without any illustration. The type material was already lost by 1913, but Roewer

managed to transfer it to *Discocyrtus* based on further data provided by Prof. Hirst (NHM). The data provided by Hirst, in turn, were based on the specimens identified by Butler (1873). Considering the preliminary data from an ongoing revision of *Discocyrtus* conducted by Rafael N. Carvalho (i.e. Carvalho 2018; Carvalho & Kury 2018; Kury & Carvalho 2016), it is unlikely that the placement proposed by Roewer were correct. It was clear that this species was misplaced in *Discocyrtus* as voiced for example by Ringuélet (1954c: 3). Considering the (very) few characters that Gervais provided, we can infer that it is a female gonyleptid, especially by the phrase: “cuisses un peu courtes portant quelques légères saillies épineuses” [= coxae short, with weak acuminate tubercles]. Gervais also mentioned (1) “tubercules gemmiformes” [= bud-shaped tubercles]; (2) “corselet... ovalaire et un peu en forme de lyre” [= scutum ovoid and roughly lyre-shaped = type gamma of Kury & Medrano (2016)]; (3) “corselet... divisé dans toute sa surface en plusieurs compartiments” [= scutum with several well-defined areas]. Both authors also hint that the lateral margins of dorsal scutum and free tergites I–III are unarmed. Taking the provenance also into account, all those features are consistent with *G. planiceps*.



**Figure 22.** *Eubalta planiceps* (Guérin-Méneville, 1842), *in vivo*, from Reserva Nacional Magallanes, Punta Arenas, Chile; female. Images copyright © Gonzalo Giribet, used with permission.



# Chapter 13. Gonyassamiinae, *Bristoweia*, *Eopachylus* and Project 2559

by AB Kury, AC Mendes & MR Hara

An AracnoLab expedition in 2008 discovered a small bromelicolous species which bridged the morphological gap between *Graphinotus* and *Trichominua*. At the same time, when identifying “pachyline” species for routine work in the collection of AracnoLab, AB Kury came across the obscure genera *Eopachylus* Mello-Leitão, 1931 and *Bristoweia*. *Eopachylus* has two species never rediscovered and which do not share any special similarity, having been originally put together only by superficial traits of the Roewerian system. An attempt to figure out their relationships caused a much larger investigation than initially expected and ultimately led to the discovery of two hitherto undetected subfamilies of Gonyleptidae (Gonyassamiinae *hoc sensu* and another in process of description). AB Kury did extensive **study of type specimens** of potential Gonyassamiinae in many museums and a survey of morphological characters which resulted in a matrix (**MorphoBank Project 2559**). Formal publication of the taxonomic changes of this “Gonyassamiinae Project” were hampered by immense delays in securing material, other projects piling up and ultimately the **destruction of the AracnoLab collection** in 2018. A considerable amount of undescribed species was detected in several genera, but the correspondent material was destroyed. This project is currently **stalled** while we rebuild the fantastic collection of Gonyassamiinae that was burned, but it is expected to be retaken, as a collaboration of AB Kury, AC Mendes and MR Hara. Nevertheless, a fraction of the preliminary results of that project can be advanced herein.

## Gonyassamiinae Soares & Soares, 1988

- Gonyassamiinae [subfamily of Gonyleptidae] Soares & Soares 1988c: 23. 🐞 Type genus: *Gonyassamia* Soares & Soares 1988. Type genus is a junior synonym, which by itself does not affect the validity of the subfamily.

**Diagnosis.** Ocularium narrower than most Gonyleptidae, placed away from anterior border of carapace, bearing long, more or less hooked spiniform

process (except in some *Graphinotus*, which have a small globular tubercle). Mesotergum with 3 or 4 areas, armature variable, either paired or unpaired. Coxa IV of male either with one or 2 dorso-apical apophyses of varied sizes, shapes and directions; retroapical apophysis lacking. Trochanter IV of male mostly with powerful prodorsal-apical apophysis. Femur IV of male variedly armed, always lacking dorso-basal apophysis. Male genitalia (Fig. 22): Pedestal of glans trapezoid in dorsal view; podium not overlapping ventral plate in lateral view; head of stylus with apex subsquare in lateral view, compressed; ventral process present as a small flabellum, sometimes partly or totally fused to stylus (then called **pseudoflabellum**); attachment of stylus to glans erect. It is noteworthy the presence of a few species with reduced sexual dimorphic armature.

**Background and current proposals.** Gonyassamiinae was created by Soares & Soares (1988c: 23) for the single species *Gonyassamia roeweri* (Soares & Soares, 1954). Kury (2003a: 119) expanded the concept of Gonyassamiinae to also include *Phalangochilus* Mello-Leitão, 1938 and *Trichominua* Mello-Leitão, 1938, synonymizing *Gonyassamia* with the latter. Based on preliminary but steady results of a MP analysis of the matrix in Project 2559 is possible to outline a **new ergotaxonomy for the Gonyassamiinae**.

**Included genera.** *Acanthopachyloopsis* Soares & Soares, 1949 transl. nov. (transferred from Pachylinae), *Eugyndes* Roewer, 1923 transl. nov. (transferred from Pachylinae), *Graphinotus* C.L. Koch, 1839 transl. nov. (transferred from Pachylinae), *Gyndulus* Roewer, 1929 transl. nov. (transferred from Pachylinae), *Phalangochilus* Mello-Leitão, 1938, *Progyndes* Roewer, 1917 transl. nov. (transferred from Pachylinae), *Trichominua* Mello-Leitão, 1938.

## *Acanthopachyloopsis* Soares & Soares, 1949

- *Acanthopachyloopsis* Soares & Soares 1949a: 51. 🐞 Type-species by original designation: *Acanthopachyloopsis spectabilis* Soares & Soares, 1949.
- *Oxyrhina* B. Soares 1944c: 181. 🐞 Type-species by original designation: *Oxyrhina zoppeii* B. Soares, 1944. II Junior homonym of *Oxyrhina* Agassiz 1835: 86 (**Elasmobranchii: Lamnidae**). **Syn. nov.**
- *Rhioxyyna* Soares & Bauab-Vianna 1970: 139. 🐞 Type-species automatically (ICZN Code Art. 67.8) the same as for *Oxyrhina*. 📌 Replacement for *Oxyrhina* B. Soares, 1944. **Syn. nov.**

**Placement and composition.** *Acanthopachyloopsis* was hitherto a monotypic genus of Pachylinae. With the current expanded diagnosis it now encompasses five species from the Brazilian Atlantic Forest. These species were not recognized previously as a natural unit probably because of the varied scutal armature.

**Etymology.** From pre-existing genus *Acanthopachylus* Roewer, 1913 + Greek ὄψις (aspect, appearance). Gender feminine.

**Diagnosis.** Armature of mesotergal areas III–IV extremely variable; other areas and free tergites unarmed. Left and right halves of mesotergal area I broadly joined. Chelicerae monomorphic. Coxa IV of male rather short (barely reaching the posterior margin of DS). Trochanter IV of male always with well-developed prodorsal-apical apophysis, with varied orientation. Femur IV of male short, with longitudinal rows of spines. Ventral process of stylus apparently largely fused with it; distal setae of penis ventral plate long and curved. There are 2 unique notable synapomorphies distinguishing *Acanthopachylopsis* from all other Gonyleptidae: (1) antero-lateral corners of area I smooth and glossy, without tuberculation and (2) hook of the ocularium immensely developed and curved, turning downwards until reaching the level of the base. The wide variation of the mesotergal armature prevented the recognition of the species of this genus in the restrictive Roewerian system. *Acanthopachylopsis* is most closely related to *Gyndulus* (by striking genital features), and is easily distinguished of it by the prodorsal-apical apophysis of coxa IV of male normal, instead of being an L-shaped protuberance (which is a unique synapomorphy for *Gyndulus*).

**Included species.** *Eopachylus illectus* Soares & Bauab-Vianna, 1970, *Progyndes iporangae* Soares & Soares, 1945, *Acanthopachylopsis spectabilis* Soares & Soares, 1949, *Rhioxyna unicornis* Soares & Bauab-Vianna, 1970, *Oxyrhina zoppeii* B. Soares, 1944.

***Acanthopachylopsis illecta* (Soares & Bauab-Vianna, 1970) comb. nov.**

- *Eopachylus illectus* Soares & Bauab-Vianna 1970: 136, figs 6–7.

***Acanthopachylopsis iporangae* (Soares & Soares, 1945) comb. nov.**

- *Progyndes iporangae* Soares & Soares 1945c: 266, fig. 11.

***Acanthopachylopsis spectabilis* Soares & Soares, 1949**

- *Acanthopachylopsis spectabilis* Soares & Soares 1949a: 51, figs 3–4.

***Acanthopachylopsis unicornis* (Soares & Bauab-Vianna, 1970) comb. nov.**

- *Rhioxyna unicornis* Soares & Bauab-Vianna 1970: 139, figs 15–16.

***Acanthopachylopsis zoppeii* (B. Soares, 1944) comb. nov.**

- *Oxyrhina zoppeii* B. Soares 1944c: 181, fig. 2.  
*Rhioxyna zoppeii*: Soares & Bauab-Vianna 1970: 140.

***Eugyndes* Roewer, 1923**

- *Gyndes* Sørensen 1884: 637. 🐞 Type-species by monotypy: *Gyndes reinhardi* Sørensen, 1884. ⚔ Junior homonym of *Gyndes* Stål, 1862: 314 (Hemiptera: Rhyparochromidae).
- *Eugyndes* Roewer 1923: 397. 🐞 Type-species automatically (ICZN Code Art. 67.8) the same as for *Gyndes*. 📄 Replacement name for *Gyndes* Sørensen, 1884.
- *Soerenseniopilio* Strand 1942: 396. 📄 Unneeded replacement name for *Gyndes* Sørensen, 1884; first noted by Roewer (1943).

**Etymology.** *Gyndes* from Greek Γύνδης (a tributary of the Tigris, in Assyria). Gender masculine. *Eugyndes* from Greek εὖ (rightful, proper, good) + pre-existing genus *Gyndes*. Gender masculine.

**Placement and composition.** While the type species of *Bristoweia* is entirely unrelated to Gonyassamiinae, the second species, *B. zorodes* (Figs 21g–h), was rediscovered during the course of this project, and a male specimen could be studied before destroyed in 2018. Both the external and genital morphology match well the diagnosis of *Eugyndes* as given below. The type material of both *E. reinhardi* and *E. patellaris* has been examined by ABK and found to be conspecific. Fresh material of *Progyndes trochanteralis* has been studied and it does not match the current diagnosis of *Progyndes*, rather fitting well in *Eugyndes* as defined here.

**Diagnosis.** Dorsal scutum alpha or gamma type. Ocularium narrow, moderately high, with strong spine inclined frontwards. Dorsal scutum and free tergites unarmed, except for area III, armed with 2 paramedian short spines strongly inclined backwards or globular tubercles. Scutal areas III–IV independent, fully developed, parallel to area II. Left and right halves of mesotergal area I broadly joined. Coxa IV of male widely surpassing dorsal scutum from the basal part, its outline sharply changing direction to become almost parallel with scutum; prodorsal-apical apophysis short, truncate, with 2 lobes; without retrolateral-apical apophysis. Trochanter IV of male moderately elongate, with powerful dorso-apical anvil-tipped hook. Femur IV of male short and thick, only slightly sinuous, provided with longitudinal rows of coarse tubercles and distal half with 2 dorsal and 2 retrolateral robust spiniform apophyses pointing to inner side. Patella IV of male coarsely granular, with immensely robust sharply polygonal ventro-distal apophysis. Tibia IV of male incrassate, with longitudinal rows of strong tubercles. Basitarsus I with only one very thick joint; distitarsus I with 3 joints, distal one longest. Tarsal counts: 4(3)/6(3)/6/6. Ventral plate elongate, in some cases extremely so. All macrosetae extremely reduced, except for C1–C3 which are well developed (Figs 22f–g).

Distinguished from *Graphinotus* and *Progyndes* by the erect trochanteral hook (strongly displaced prolaterally).



erally in these genera, forming pincer with coxal apophysis); tibia III slender, without ventral rows of acuminate tubercles or paramedian armature on mesotergal area III. *Graphinotus* may be further distinguished by the areas III-IV merged and partially sunken into area II; pale patches placed on lateral and posterior margins of DS forming a U-shaped lyra; area III with a single stout spine (paramedian granules in *G. flavolimbatus*). *Progyndes* may be further distinguished by the row of numerous very strong prolateral spines of femur IV.

**Included species.** *Gyndes reinhardi* Sørensen, 1884, *Progyndes trochanteralis* Soares & Bauab-Vianna, 1972, and *Bristoweia zorodes* Mello-Leitão, 1924.

**Excluded species.** *Eugyndes flavolimbatus* H. Soares, 1946, herein transferred to *Phalangochilus* (see below).

#### ***Eugyndes reinhardi* (Sørensen, 1884)**

- *Gyndes reinhardi* Sørensen 1884: 638.
- *Eugyndes patellaris* H. Soares 1966b: 107, figs 1 and 4. **Syn. nov.**

#### ***Eugyndes trochanteralis* (Soares & Bauab-Vianna, 1972) comb. nov.**

- *Progyndes trochanteralis* Soares & Bauab-Vianna 1972a: 213, figs 9–13.

#### ***Eugyndes zorodes* (Mello-Leitão, 1924) comb. nov.**

- *Bristoweia zorodes* Mello-Leitão 1924: 180.

#### ***Graphinotus* C.L. Koch, 1839**

- *Graphinotus* C.L. Koch 1839b: 10. 🐞 Type-species by monotypy: *Graphinotus ornatus* C.L. Koch, 1839. 📖 Made available before 1931 as a combined description and without being stated as new (**Code Art. 12.2.6**).
- *Therezopolis* Mello-Leitão 1923c: 133. Type-species by original designation: *Therezopolis therezopolis* Mello-Leitão, 1923. **II** Junior subjective synonym of *Graphinotus* C.L. Koch, 1839 by Soares & Soares (1986: 92).
- *Vitiches* Roewer 1927b: 347. 🐞 Type-species by monotypy: *Vitiches viridilimbata* Roewer, 1927. **II** Junior subjective synonym of *Therezopolis* Mello-Leitão, 1923 by Mello-Leitão (1932: 460). Junior subjective synonym of *Graphinotus* C.L. Koch, 1839 by Soares & Soares (1986: 92).
- *Paraorguesia* Mello-Leitão 1927b: 16. 🐞 Type-species by original designation: *Paraorguesia albiornata* Mello-Leitão, 1927. **II** Junior subjective synonym of *Graphinotus* C.L. Koch, 1839 by Soares & Soares (1986: 92).
- *Wygodzinskyia* Soares & Soares 1945f: 339. 🐞 Type-species by original designation: *Wygodzinskyia viridiornata* Soares & Soares, 1945. **II** Junior subjective synonym of *Graphinotus* C.L. Koch, 1839 by Soares & Soares (1986: 92).

**Etymology.** *Graphinotus* from Greek γράφω (to write, to draw) + νῶτος (back). Gender masculine.

**Placement and composition.** *Graphinotus therezopolis*, from the central mountain range of RJ, has been

described three times. B. Soares (1944i) synonymized the two junior nomina with the senior, with which we concur. Later, Soares & Soares (1986) synonymized all of those with *G. ornatus*. However we recognize herein two very closely related species of *Graphinotus* which differ from all others in the genus by the armature of ocularium being reduced to a globular tubercle. Both species are allopatrid: *G. ornatus* occurs in Nova Friburgo and Petrópolis, while *G. therezopolis* is known so far only from Teresópolis. *G. ornatus* is clearly recognizable by the neon green blots extending also to mesotergal areas I and II (**Fig. 21a**).

**Diagnosis.** Scutum outline gamma-type. Dorsal scutum with yellow/green/orange flattened contiguous patches extending from carapace backwards. Left and right halves of mesotergal area I broadly joined. Mesotergal areas II and IV fused to each other. Scutal area III armed with single median high acuminate spine. Basitarsomeres of leg I of male fused to each other and strongly incrassate. Macrosetae A forming triangle in lateral view. Macrosetae C1–C3 short and straight.

**Included species.** *Wygodzinskyia gratiosa* H. Soares, 1974, *Therezopolis magnifica* Roewer, 1943, *Graphinotus ornatus* C.L. Koch, 1839, *Therezopolis therezopolis* Mello-Leitão, 1923 (herein revalidated from the synonymy of *G. ornatus*), *Therezopolis roseus* Mello-Leitão 1936, and *Wygodzinskyia viridiornata* Soares & Soares 1945

#### ***Graphinotus gratiosus* (H. Soares, 1974)**

- “*Wygodzinskyia*” *gratiosa*: H. Soares 1974b: 359, figs 8–14. 📖 Incorrect subsequent spelling of genus *Wygodzinskyia* Soares & Soares, 1945, which has no nomenclatural effect on the species.  
*Graphinotus gratiosus*: Soares & Soares 1986: 93.

#### ***Graphinotus magnificus* (Roewer, 1943)**

- *Therezopolis magnifica* Roewer 1943: 40, pl. 5, figs 41–41a.
- *Therezopolis franciscoi* Soares & Soares 1945d: 281, figs 1–2. **II** Junior subjective synonym of *Therezopolis magnifica* Roewer 1943 by Kury (2003a: 171).

#### ***Graphinotus ornatus* C.L. Koch, 1839**

- *Graphinotus ornatus* C.L. Koch 1839b: 10, pl. 219, fig. 545.

#### ***Graphinotus roseus* (Mello-Leitão, 1936)**

- *Therezopolis roseus* Mello-Leitão 1936b: 26, figs 21–22.  
*Graphinotus roseus*: Soares & Soares 1986: 93, figs 13–14.
- *Therezopolis sawayai* H. Soares 1946: 387, fig. 3. **Syn. nov.**  
*Graphinotus sawayai*: Soares & Soares 1986: 93, figs 15–16.

#### ***Graphinotus therezopolis* (Mello-Leitão, 1923)**

- *Therezopolis therezopolis* Mello-Leitão 1923c: 133. **II** Junior subjective synonym of *Graphinotus ornatus* C.L. Koch, 1839 by Soares & Soares (1986: 92). **Synonymy rejected here.**



- *Vitiches viridilimbata* Roewer 1927b: 347, fig. 15. **II** Junior subjective synonym of *Therezopolis therezopolis* Mello-Leitão, 1923 by B. Soares (1944i: 169). **II** Junior subjective synonym of *Graphinotus ornatus* C.L. Koch, 1839 by Soares & Soares (1986: 92).

*Therezopolis viridilimbatus*: Mello-Leitão 1932: 460.

- *Paraorguesia albiornata* Mello-Leitão 1927b: 17. **II** Junior subjective synonym of *Therezopolis therezopolis* Mello-Leitão, 1923 by B. Soares (1944i: 170). **II** Junior subjective synonym of *Graphinotus ornatus* C.L. Koch, 1839 by Soares & Soares (1986: 92).

### ***Graphinotus viridiornatus* (Soares & Soares, 1945)**

- *Wygodzinskyia viridiornata* Soares & Soares 1945f: 340, fig. 1.  
*Graphinotus viridiornatus*: Soares & Soares 1986: 94, figs 17–18.

### ***Gyndulus* Roewer, 1929**

- *Gyndulus* Roewer 1929: 218. 🐛 Type-species by monotypy: *Gyndulus trispinifrons* Roewer, 1929.
- *Lussanvira* Mello-Leitão 1935c: 372. Type-species by original designation: *Lussanvira marmorata* Mello-Leitão, 1935. **Syn. nov.**
- *Trochanteroceros* Canals 1935: 6. 🐛 Type-species by monotypy: *Trochanteroceros misionicus* Canals, 1935. **Syn. nov.**
- *Anoplogynus* Piza 1938b: 117. 🐛 Type-species by original designation: *Anoplogynus nasutus* Piza, 1938. **Syn. nov.**
- *Neotrochanteroceros* Canals 1943: 4. 🐛 Type-species by monotypy: *Neotrochanteroceros singularis* Canals, 1943. **II** Junior subjective synonym of *Anoplogynus* Piza, 1938 by B. Soares (1944e: 259). **Syn. nov.**
- *Beckeresia* H. Soares 1970a: 212. 🐛 Type-species by original designation: *Beckeresia beckeri* H. Soares, 1970. **Syn. nov.**
- *Eopachyloides* H. Soares 1970a: 214. 🐛 Type-species by original designation: *Eopachyloides trochanteralis* H. Soares, 1970. **Syn. nov.**
- *Eugyndopsiella* H. Soares 1972: 66. 🐛 Type-species by original designation: *Eugyndopsiella trochanteroceros* H. Soares, 1972. **Syn. nov.**

**Etymology.** *Gyndulus* from pre-existing genus *Gyndes* + Latin diminutive suffix *-ulus*. Gender masculine.

**Placement and composition.** Roewer (1929f) described the new genus *Gyndulus* along with the new species *Gyndulus trispinifrons* from “Mato Grosso” (which today corresponds to two different Brazilian states). The holotype was reported as a male, while Roewer’s drawing clearly allows to recognize it as a female. Roewer often made the same mistake in the case of Pachylinae with some armature on femur IV of females, without having the male (which is even more heavily armed) for comparison. Roewer (1943) added a second species, *Gyndulus curvitibialis*, from Chile, based only on tarsal counts and scutal armature, not surprisingly being entirely unrelated to the first. Soares & Soares (1954b) synonymized *Gyndulus* with *Progyndes*, carrying both species. *Gyndulus curvitibialis* became

a junior secondary homonym, being replaced with *Progyndes roeweri*, an unlucky name, carelessly used by the same authors in the same year for an entirely unrelated species from SE Brazil. Soares & Bauab-Vianna (1972a) briefly revalidated *Gyndulus*, without any meaningful comment. Soares & Soares (1985a) reported an additional female of *G. trispinifrons* from Paraguay (thus extending southwestwards the distribution of this species within the same general ecorregion), and casted doubt on the maleness of Roewer’s holotype. Their illustration allows clear recognition of characters existing in the present concept of Gonyassamiinae. *Lussanvira* was originally in Tricommatinae, then removed to Gonyleptidae incertae sedis by Kury (2014). The other genera synonymized here were originally described in Pachylinae as monotypic and never related to any other genus, except two of them, which were synonymized to each other.

**Diagnosis.** Dorsal scutum gamma type, with narrow ocularium. Left and right halves of mesotergal area I broadly joined. Mesotergal area I with a pair of paramedian tubercles; many species armed on the sides of mesotergal area III. Mesotergal groove II median portion invading mesotergal area II (attenuate in females). Chelicerae monomorphic. Coxa IV of male short, barely reaching mesotergal area IV or visible in dorsal view, with prodorsal apical apophysis doubled, both units forming a straight angle. Basitarsomeres of leg I of male neither incrassate nor fused to each other. Male genitalia: Ventral plate subrectangular (VP basal lobe well-developed, as large as distal lobe) with accentuated bulge on proximal half. Macrosetae A forming triangle in lateral view. Macrosetae C clustered apically on VP. Macrosetae D inserted laterally, between A and C. Pseudoflabellum present.

**Included species.** *Beckeresia beckeri* H. Soares, 1970, *Beckeresia delicata* Soares & Soares, 1970, *Trochanteroceros misionicus* Canals, 1935, *Anoplogynus nasutus* Piza, 1938, *Neotrochanteroceros singularis* Canals, 1943, *Gyndulus trispinifrons* Roewer, 1929, *Eopachyloides trochanteralis* H. Soares, 1970 and *Eugyndopsiella trochanteroceros* H. Soares, 1972.

### ***Gyndulus beckeri* (H. Soares, 1970) comb. nov.**

- *Beckeresia beckeri* H. Soares 1970a: 213, figs. 1–5.

### ***Gyndulus delicatus* (Soares & Soares, 1970) comb. nov.**

- *Beckeresia delicata* Soares & Soares 1970: 344, fig. 6.

### ***Gyndulus misionicus* (Canals, 1935) comb. nov.**

- *Trochanteroceros misionicus* Canals 1935: 6, fig. 3.

### ***Gyndulus nasutus* (Piza, 1938) comb. nov.**

- *Anoplogynus nasutus* Piza 1938b: 118, figs 4–5.



**Figure 23.** Gonyassamiinae *in vivo*: (a). *Graphinotus ornatus* C.L. Koch, 1839, male from Nova Friburgo, RJ; (b). *Phalangochilus flavolimbatus* (H. Soares, 1946), male from Nova Friburgo, RJ; (c). *Graphinotus* sp., male from Macaé, RJ, *in vivo*; (d). *Acanthopachyloopsis spectabilis* Soares & Soares, 1949, male from Piraquara, PR; (e). *Gyndulus delicatus* (Soares & Soares, 1970), male from Brejo da Lapa, MG. Photographs copyright © Arthur Anker (a, b), AB Kury (c-e) used with permission.

***Gyndulus singularis* (Canals, 1943) comb. nov.**

- *Neotrochanteroceros singularis* Canals 1943: 4, figs 2a–b.  
*Anoplogynus singularis*: B. Soares 1944e: 259.

***Gyndulus trispinifrons* Roewer, 1929**

- *Gyndulus trispinifrons* Roewer 1929: 219, fig. 15.  
*Progyndes trispinifrons*: Soares & Soares 1954b: 293.
- *Lussanvira marmorata* Mello-Leitão 1935c: 372, fig. 3. **Syn. nov.**

***Gyndulus trochanteralis* (H. Soares, 1970) comb. nov.**

- *Eopachyloides trochanteralis* H. Soares 1970a: 214, figs 6–10.

***Gyndulus trochanteroceros* (H. Soares, 1972) comb. nov.**

- *Eugyndopsiella trochanteroceros* H. Soares 1972: 66, figs 5–9.

***Phalangochilus* Mello-Leitão, 1938**

- *Phalangochilus* Mello-Leitão 1938a: 139. 🐛 Type-species by original designation: *Phalangochilus luteipes* Mello-Leitão, 1938.

**Etymology.** Not given, but most probably a combination of pre-existing generic names *Phalangodes* + *Pachylus*. Gender masculine.

**Placement and composition.** *Phalangochilus* was originally placed in Phalangodinae, but has been transferred to Gonyassamiinae by Kury (2003a). Males of *P. luteipes* are as yet unknown, but it shares sufficient special similarity with *E. flavolimbatus* to be considered its sister group. The latter does not share any specific similarity with *Eugyndes* and it is here used to build a more complete diagnosis for *Phalangochilus*.

**Diagnosis.** Dorsal scutum gamma type without yellow/green patches on mesotergum, when present restricted to two arches on lateral margins of the abdominal scutum. Ocularium with thick short spine. Left and right halves of mesotergal area I broadly joined. Mesotergal area III armed with pair of closely joined, posteriorly located lighter paramedian small tubercles which deform posterior border of area III making it projects into area IV. Mesotergal area IV divided into two halves. Chelicerae monomorphic. Ventral plate subsquare, not especially elongate, with prominent basal lobes, without lateral constrictions, penetrating in truncus, with microsetae distributed along its length. Macrosetae C1–C3 elongate, curved.

**Included species.** *Phalangochilus flavolimbatus* (H. Soares, 1946) and *Phalangochilus luteipes* Mello-Leitão, 1938.

***Phalangochilus flavolimbatus* (H. Soares, 1946) comb. nov.**

- *Eugyndes flavolimbatus* H. Soares 1946: 386, fig. 2.

***Phalangochilus luteipes* Mello-Leitão, 1938**

- *Phalangochilus luteipes* Mello-Leitão 1938a: 139, fig. 1.

***Progyndes* Roewer, 1917**

- *Progyndes* Roewer 1917: 107. 🐛 Type-species by monotypy: *Progyndes curvibialis* Roewer, 1917.

**Etymology.** From Greek πρό (“before, forth”) + pre-existing generic name *Gyndes*. Gender masculine.

**Placement and composition.** *Progyndes* has a long and convoluted history, having included many unrelated species, which are now in different genera and subfamilies. Roewer (1917) created the new genus *Progyndes* in Pachylinae to include the new species *Progyndes curvibialis*, from “Argentina” (this locality however is incorrect, as this species is consistently and exclusively found in Rio de Janeiro). Mello-Leitão (1935a; 1940c) added a second and a third species *Progyndes basiliscus* and *Progyndes brasiliensis*, both from Rio de Janeiro. Soares & Soares (1945c) added a 4th species, *Progyndes iporangae*, from São Paulo. Soares & Soares (1954b) put *Pachyloidellus* Müller, 1917 and *Gyndulus* Roewer, 1929 in the synonymy of *Progyndes* (both later revalidated), combining many extraneous species under *Progyndes* – *Pachyloidellus fuscus* Müller, 1917, *Pachyloidellus tricalcaratus* Roewer, 1923, *Pachyloidellus crassus* Roewer, 1943, *Gyndulus curvibialis* Roewer 1943 (which thereby became a junior secondary homonym of *Progyndes curvibialis* Roewer, 1917, forcing the creation of the replacement *Progyndes roeweri* Soares & Soares 1954), and *Gyndulus trispinifrons* Roewer 1929. Soares & Soares (1954a) in the same year absent-mindedly created yet another *Progyndes roeweri*, from Rio de Janeiro, homonym of their replacement name. Soares & Bauab-Vianna (1972) described one more species, *Progyndes trochanteralis*, from Minas Gerais, which we herein judge to belong in *Eugyndes* (see that genus), revalidated *Gyndulus* (without explanation, but thereby removing the secondary homonymy), mentioned *Pachyloidellus crassus*, still in *Progyndes*, even after *Pachyloidellus* having been transferred to the synonymy of *Acanthopachylus* (by Ringuet 1959). Soares & Soares (1988c) transferred *Progyndes roeweri* Soares & Soares, 1954 to the new genus *Gonyassamia*, then made type of the senseless new monotypic subfamily Gonyassamiinae. Finally Acosta (1996c) transferred the Chilean *Pachyloidellus crassus* Roewer 1943 to *Pachylus*, leaving *Progyndes* with five species, as appeared in Kury (2003a).

**Diagnosis.** Dorsal scutum gamma type, without yellow/green patches whatsoever. Ocularium with high forward inclined spine. Left and right halves of mesotergal area I broadly joined. Laterals of mesotergal area II not projected anteriorly, front-post asymmetrical. Scutal area IV entire. All mesotergal areas with a transverse row of minute granules. Chelicerae monomor-

phic. Retrodorsal-apical apophysis of femur IV of male truncated. Tibia IV of male often short, stout, sinuous, clavate and densely spined. Basitarsomeres of leg I of male fused to each other and strongly incrassate. Ventral plate subsquare, not especially elongate, without lateral constrictions, with prominent basal lobes, penetrating in truncus, with microsetae distributed along its length. Full complement of stout macrosetae A1–A3. Macrosetae C1–C3 mostly hyper developed, angular, with apex curved and flattened.

**Included species.** *Progyndes brasiliensis* Mello-Leitão, 1940, and *Progyndes curvibtibialis* Roewer, 1917.

#### ***Progyndes brasiliensis* Mello-Leitão, 1940**

- *Progyndes brasiliensis* Mello-Leitão 1940c: 105.

#### ***Progyndes curvibtibialis* Roewer, 1917**

- *Progyndes curvibtibialis* Roewer 1917: 107, fig. 14.
- *Progyndes basiliscus* Mello-Leitão 1935d: 11, fig. 3. **Syn. nov.**

#### ***Trichominua* Mello-Leitão, 1938**

- *Trichominua* Mello-Leitão 1938: 141. 🦋 Type-species by original designation: *Trichominua annulipes* Mello-Leitão, 1938.
- *Gonyassamia* Soares & Soares 1988c: 24. 🦋 Type-species by original designation: *Progyndes roeweri* Soares & Soares, 1954. II Junior subjective synonym of *Trichominua* Mello-Leitão, 1938 by Kury (2003a: 119)



**Figure 24.** Gonyassamiinae; external morphology: (a). *Eugyndes reinhardi* (Sørensen, 1884), male holotype (ZMUC) patella IV, dorsal view; (b). Same, habitus, sinistrolateral view; (c). *Trichominua roeweri* (Soares & Soares, 1954), male (MNRJ 2395), sinistrolateral view; (d). *Eugyndes zorodes* (Mello-Leitão, 1924), male (MNRJ 7225), dorsal view; (e). Same, female. Scale bars: 0.5 mm (a), 1 mm (c, d, e), 2 mm (b).

**Etymology.** *Trichominua* is possibly a combination of pre-existing generic names *Tricommatatus* + *Minua*. Gender masculine. *Gonyassamia* is a combination of pre-existing generic names *Gonyleptes* + *Assamia*. Gender feminine.

**Placement.** *Trichominua* originally in Tricommatinae. Transferred to Gonyassamiinae by Kury (2003a).

**Diagnosis.** Dorsal scutum gamma type, with whitish yellow tubercles on the DS lateral margin. Carapace sexually dimorphic, in males much higher than abdominal scutum and much longer than in females with immense ocularium. Left and right halves of mesotergal area I touching only in a narrow part. All scutal areas entirely unarmed. Chelicerae sexually dimorphic, swollen in male. Dimorphic armature of legs extremely reduced. Coxa IV in both sexes short (barely reaching scutal grooves III/IV, and unarmed. Basitarsomeres of leg I of male fused to each other and strongly incrassate.

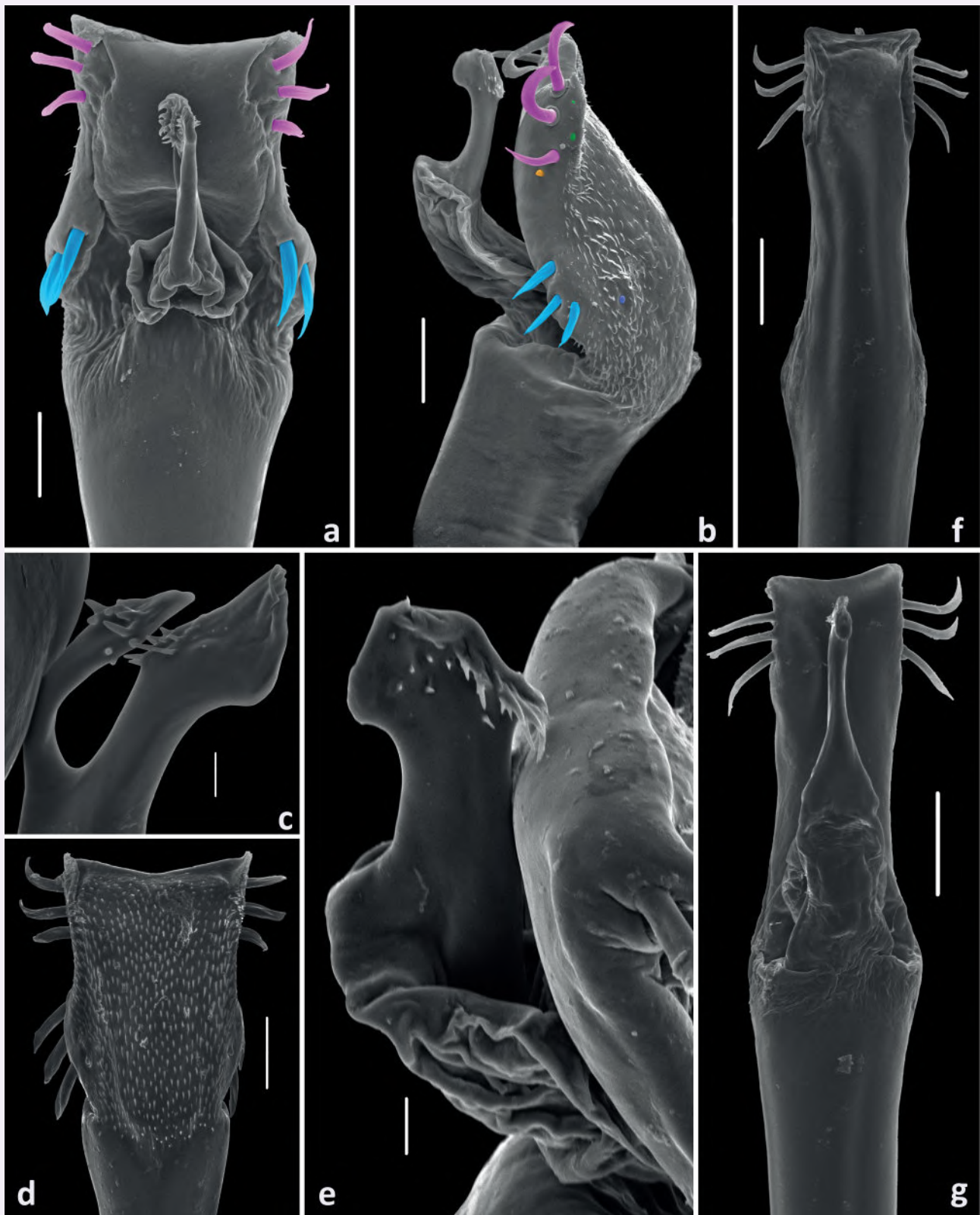
**Included species.** *Trichominua annulipes* Mello-Leitão, 1938 and *Progyndes roeweri* Soares & Soares, 1954.

#### ***Trichominua annulipes* Mello-Leitão, 1938**

- *Trichominua annulipes* Mello-Leitão 1938: 141, fig. 2.

#### ***Trichominua roeweri* (Soares & Soares, 1954)**

- *Progyndes roeweri* Soares & Soares 1954a: 501, fig. 10.  
*Gonyassamia roeweri*: Soares & Soares 1988c: 24, figs 1–9.  
*Trichominua roeweri*: Kury 2003a: 119.






**Figure 25.** Gonyassamiinae; genital morphology of several species, distal part of penis: (a). *Progyndes* sp. (MNRJ 17558), dorsal view; (b). *Gyndulus beckeri* (H. Soares, 1970) (MNRJ 17582), dextralateral view; (c). *Progyndes brasiliensis* Mello-Leitão, 1940 (MNRJ 2083), glans, sinistrolateral view; (d). Same, ventral plate, ventral view; (e). *Gyndulus singularis* (Canals, 1943) (MACN AK 075), glans, dextralateral view; (f). *Eugyndes* sp. (MNRJ 7248), ventral view; (g). Same, dorsal view. Artificial color conventions for ventral plate macrosetae are: A (blue), B (dark blue), C (magenta), D (golden yellow), E (green).

# Chapter 14. False *Gyndulus* from Austral lands (Grassatores: Gonyleptidae)

by BS Sabattini & MR Hara

## *Metagyndes pulchellus* (Loman, 1899)

- *Gyndes pulchellus* Loman 1899: 8  
*Metagyndes pulchella*: Roewer 1913c: 21, fig. 4.   
Incorrect subsequently applied gender declination.
- *Gyndulus curvitibialis* Roewer 1943: 16, pl. 1, fig. 2.  Junior secondary homonym of *Progyndes curvitibialis* Roewer, 1917 in virtue of the subjective synonymy of *Gyndulus* Roewer, 1929 with *Progyndes* Roewer, 1917 established by Soares & Soares (1954b: 291). **Syn. nov.**
- *Progyndes roeweri* Soares & Soares 1954b: 292.  Nomen novum for *Gyndulus curvitibialis* Roewer, 1943; no longer congeneric with *Progyndes curvitibialis* Roewer, 1917 in virtue of the revalidation of *Gyndulus* by Soares & Bauab-Vianna (1972a: 211); in current use (ICZN Art. 59.3). **Syn. nov.**  
*Gyndulus roeweri*: Kury 2003a: 172.  
*Metagyndes roeweri*: Pérez-Schultheiss et al. 2020: 3, figs 1–7.

**Type data.** *Gyndes pulchellus*: 1 male and 3 female syntypes, det. J.C.C. Loman (ZMB 7842, examined), “Chile; Corral”. *Gyndulus curvitibialis*: 3 male syntypes, det. C.-F. Roewer, 1929 (SMF 9901374 - RII/1374/67 - 34, not examined), “Chile (Santiago)”. Lectotype selected by E.A. Maury (1989) but not published.

**Comment.** Loman (1899) described *G. pulchellus* without providing any illustrations, as was typical of that time. Roewer (1913c) first provided the illustration of that species in his Gonyleptidae revision. However, he only illustrated *G. pulchellus* leg’s IV, which closely resembled that of *Metagyndes martensii* (Sørensen in Henriksen 1902).

In an ongoing revision of *Metagyndes* and putative closely related taxa (such as genera *Calcarogyndes* Mello-Leitão, 1932, *Pseudogyndes* Mello-Leitão, 1932 and the species then known as *Gyndulus roeweri* (Soares & Soares, 1954)), BSS and MRH were able to access type material of both *M. martensii* and *G. pulchellus* and check their identity.

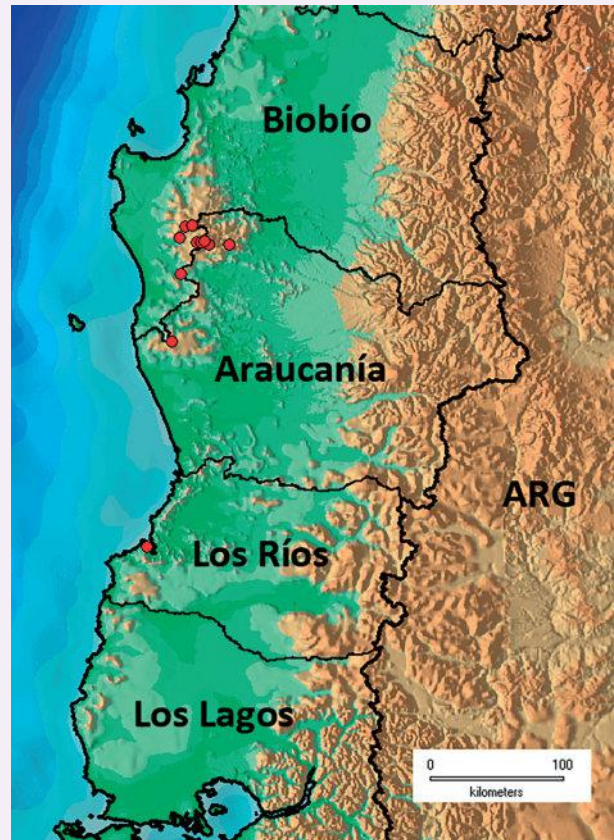
They concluded that *M. martensii* and *G. pulchellus* are indeed two distinct species, but the illustration of the leg IV of *G. pulchellus* provided by Roewer (1913c) was mistaken. Roewer’s illustration of the leg IV of *M. pulchellus* actually represents the leg IV of *M. martensii*. Furthermore, BSS and MRH also verified the identity of *G. roeweri* as part of the above mentioned revision.

The male syntype of *Gyndes pulchellus* is a minor [= beta] male, with a very subtle retrocentral elevation on femur IV which would certainly be dismissed by untrained eyes. Comparing it with the remaining species of *Metagyndes* as well as the closely related putative genera and species, we stumbled upon *Gyndulus curvitibialis* (later renamed *G. roeweri* because of a secondary synonymy). Roewer’s original illustration depicts a major [= alpha] male. We have examined many vials from Nahuelbuta, and verified that there is quite a variation in the retrocentral apophysis on femur IV, ranging from a subtle elevation to a somewhat truncated apophysis, as in Roewer’s classic illustration of *Gyndulus curvitibialis*.

*Gyndulus curvitibialis/roeweri* (Fig. 27) is the second species described for this genus. Despite fitting well in *Gyndulus* according to the Roewerian system, the armature pattern of female leg IV of *G. roeweri* has nothing to do with that of *Gyndulus trispinifrons* Roewer, 1929 (type species of the genus and only known by the female holotype). The dorsal face of the leg IV of female of *G. trispinifrons* is more armed (compared to other “Pachylinae”), as is typical of that genus (for details, see Chapter 13 above on Gonyassamiinae). Other features, such as the shape of scutal groove II and armature on the anterior margin of DS, also corroborate the unlikelihood of the current placement of *G. roeweri*.

The type material of *G. curvitibialis/roeweri* was apparently lost (Acosta 1996: 216). However, this material recently resurfaced (Pérez-Schultheiss et al. 2020: 4). Fortunately, the illustration provided by Roewer (1943) allowed BSS and MRH to identify unequivocally that species in the material deposited in many museums. The redescription of Pérez-Schultheiss et al. (2020) also corroborated the identity of *G. roeweri* as a species that matches the original description by Roewer (1943) (Fig. 26 a, b). BSS and MRH were able to verify that the retrocentral armature on femur IV of *G. roeweri* is somewhat variable, which also included the form observed in *G. pulchellus*. These findings will be treated in detail elsewhere. Meanwhile, in a parallel study, JP-S also noted the resemblance of *G. roeweri* with *Metagyndes* (Pérez-Schultheiss et al. 2019). Recently, Pérez-Schultheiss et al. (2020) formally proposed the transference to that genus based on the male genitalia features that is typical of Pachylinae stricto sensu (Pinto-da-Rocha et al. 2014), and body and male trochanter IV features, which BSS and MRH agree.

We conclude, summing up all this evidence, that *G. roeweri* is indeed a *Metagyndes* and a junior synonym of *G. pulchellus*. Accordingly, most of the published records for *M. pulchellus* should be disregarded, those possibly belonging to *M. martensii* that apparently is widespread. So far, *M. pulchellus* is restricted to Nahuelbuta (Chile) and its vicinities (Pérez-Schultheiss et al. 2019; 2020) (Fig. 26).



**Figure 26.** Centro-southern Chile, showing the revised distribution of *Metagyndes pulchellus* (Loman, 1899).



**Figure 27.** *Metagyndes pulchellus* (Loman, 1899), *in vivo*, from Ex Reserva Contulmo, provincia de Arauco, Región del Biobío, Chile ( $38^{\circ}1'34''\text{S}$   $73^{\circ}11'58''\text{W}$ ): (a). Male; (b). Female. Photographs (a, b) copyright © Jorge Pérez-Schultheiss, used with permission.



## Chapter 15. Acknowledgements

The **OmniPaper** initiative gave this WCO project a great impulse. The names of all the wonderful people who contributed to OmniPaper are all listed there (and it is a long list). The University of São Paulo maintains since 2016 a program called **Alumni USP**, which allows access to important portals and websites such as Web of Science, and from which ABK greatly benefitted.

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## Chapter 16. Complete taxonomic references for Opiliones

Rather than using here only the references explicitly cited in this work, we decided to present here a complete taxonomic bibliography for Opiliones, as it appeared fractured in small portions both in the **OmniPaper** project and in the **Opiliones Wikia**. It is and it will always be a work in progress. This part of our work, then, has not originated in **TaxonWorks**. On the contrary, we are transferring all data from this list to the format used there.

Names of each given author are spelled the same here, regardless if it appeared in different forms in the publications themselves (**as explained in Section 1.2**). Nominal dates, when different from actual dates, are given between square brackets and quotation marks. Actual issue dates are given between parentheses. Whenever we could ascertain a more detailed date, we added that at the end. As for the complete citation of periodicals, as much as we find the report of the name of the city immensely useful, we are constrained by the Zootaxa format, where WCO will be eventually published.

We give here only the publisher's name instead of the complete denomination of the publishing house, therefore: (1) C.H. Zeh instead of C. H. Zeh'schen Buchhandlung, (2) Gustav Fischer instead of Gustav Fischer Verlag. We have opted for presenting references in **chronological order** rather than by author for two reasons: 1) to give historical perspective of the evolution of publication rhythm across the decades and 2) to preserve the Zeitgeist over each individual contribution.

### 1758

Linnaeus, C. (1758) *Systema naturae per regna tria naturae, secundum classes, ordines, genera, species, cum characteribus, differentiis, synonymis, locis*. Editio decima, reformata. Tomus 1. Laurentius Salvius, Holmiae [Stockholm], [iv] + 824 pp. [Issued 1 Jan 1758].



## 1759

Reichard, C. W. E. (1759) *Dissertatio inauguralis medica, De pediculis inguinalibus insectis et vermibus homini molestis von SpulWurm, HertzWurm, St. VeitWurm.* / Stanno Heringiano, Erfurti [Erfurt, Germany], 51 pp. + 3 pls. [Issued 28 Dec. 1759].

## 1763

Scopoli, J.A. (1763) *Entomologia Carniolica exhibens insecta Carnioliae indigena et distributa in ordines, genera, species, varietates. Methodo Linnaeana.* Ioannis Thomae Trattner, Vindobonae [Vienna], [xxxviii] + 419 pp. + 680 figs [43 unnumbered pls.].

## 1767

Linnaeus, C. (1767) *Systema naturae per regna tria naturae, secundum classes, ordines, genera, species, cum characteribus, differentiis, synonymis locis.* Editio duodecima reformata. Tomus 1, pars 2. Laurentius Salvius, Stockholm, 533–1327 + [37] pp.

## 1772

Pallas, P.S. (1772) Phalangia, Araneae, Acari. In: *Spicilegia Zoologica. Continens quadrupedum, avium, amphibiorum, piscium, insectorum, molluscorum aliorumque marinorum.* Vol. 1, Fasc 9. Gottlieb August Lange, Berolini [Berlin], pp. 28–50.

## 1775

Fabricius, J.C. (1775) *Systema Entomologiae, sistens insectorum classes, ordines, genera, species, adiectis synonymis, locis, descriptionibus, observationibus.* Officina Libraria Kortii, Flensburgi et Lipsiae [Flensburg and Leipzig, Germany], xxxii + 832 pp.

## 1776

Müller, O.F. (1776) *Zoologiae Danicae Prodrömus, seu Animalium Daniæ et Norvegiæ Indigenarum characteres, nomina, et synonyma imprimis popularium.* Heineck & Faber [printed by Hallager], Hafniae [Copenhagen], xxxii + 274 pp.

## 1778

De Geer, C. (1778) Des Faucheurs (pp 163–175, pl. 10). In: *Mémoires pour servir à l'histoire des insectes.* Tome 7. Pierre Hesselberg, Stockholm, xii + 950 pp., 49 pl.

## 1779

Fabricius, J.C. (1779) *Reise nach Norwegen mit Bemerkungen aus der Naturhistorie und Oekonomie.* Carl Ernst Bohn, Hamburg, lxiv + 388 + [12] pp.

## 1780

Fabricius, O. (1780) *Fauna Groenlandica: systematice sistens animalia Groenlandiae occidentalis hactenus indagata, quoad nomen specificum, triviale, vernaculumque: synonyma auctorum plurium, descriptionem, locum, victum, generationem, mores.* Ioannis Gottlob Rothe, Hafniae & Lipsiae [Copenhagen & Leipzig], xvi + 452 pp.

## 1781

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